









## 30/05/2016

# Galaxy

Initiation

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Credit to Gildas Le Corguillé - V1.07





## **INTRODUCTION / PROBLEMATIC**



- . Setup TP
  - http://galaxy.sb-roscoff.fr
  - . Account
    - login@sb-roscoff.fr
    - \*\*\*\*\*\*\*\*

```
login@sbr4-1042:~$ ssh -Y login@bioinfo.sb-roscoff.fr
[...]
[login@n0 ~]$ cdprojet
[login@n0 login]$ cd 13-07-29-panda/tmp/mapping
[login@n0 mapping]$ cat tophat.qsub
#!/bin/bash
#$ -S /bin/bash
#$ -M login@sb-roscoff.fr
#$ -m bea
#$ -V
#$ -cwd
#$ -o qsub.out
#$ -e qsub.err
tophat2 panda v121029 ../input/IllR1-1.fq ../input/IllR1-2.fq
-GTF ../input/panda v121029.gtf --b2-sensitive -r 100
-num-threads 8
[login@n0 mapping]$ qsub -q long.q -pe thread 8 tophat.qsub
Your job 5338969 ("tophat.qsub") has been submitted
[login@n0 mapping]$ ls
accepted hits.bam junctions.bed qsub.err unmapped.bam
deletions.bed logs
                           qsub.out
insertions.bed prep reads.info tmp
[login@n0 mapping]$ cd ...
```

[login@n0 mapping]\$ mkdir cufflinks

```
login@sbr4-1042:~$ ssh -Y login@bioinfo.sb-roscoff.fr
\left[ \ldots \right]
[login@n0 ~]$ cdprojet
[login@n0 login]:
[login@n0 mapping
#!/bin/bash
#$ -S /bin/bash
#$ -M login@sb-r(
#$ -m bea
#$ -V
#$ -cwd
#$ -o qsub.out
#$ -e qsub.err
                                                           .R1-2.fq
tophat2 panda v1:
-GTF ../input/pai
-num-threads 8
[login@n0 mapping
```

.qsub

l.bam

prep\_reads.info insertions.bed [login@n0 mapping]\$ cd ... [login@n0 mapping]\$ mkdir cufflinks

Your job 5338969

[login@n0 mappin(

accepted hits.bar

deletions.bed





# Select your level:







## « I want to know the gene expression »







# « I want to map my reads on a reference genome and count them »







## « I want to launch the tools tophat2 and cufflinks. I have fastq files and my genome in fasta and gtf. »







« I want 1TB for my project. I will launch tophat2 through SSH on the cluster in multi-thread mode.

Next I want to submit the bam file to my genome with cufflinks.

Except that, I will manage :P"







## « I have a bunch of cool tools! But I'm the only one who can launch them.

Comments? »

Station Biologique					
ABMS	Introduc	ction			
.gtf	.gff3 .be .wig	d .cas	.ba bam	ai .C COC	oordSorted.bai
.nhr .phr	.fai <sup>.s</sup> .fasta	sam <sup>.</sup> .vcf	bum	.na	ameSorted.bai eSorted.bam
.*.eb .re\ .xm	wt /.*.ebwt .tab I .tgz	.*.bt2 .rev.*.	bt2 .faste	q/.fq	.phred64.fq .fastqsanger

## Introduction

Station Biologique





- Graphical interface click-button tools within windows
  - + very ergonomic
  - too ergonomic  $\rightarrow$  lack of flexibility
  - don't count on it ! Have you ever seen a PhD student having the time to make beautiful green buttons?
  - paying for it!
- Tools available on the internet
  - + very ergonomic
  - too ergonomic  $\rightarrow$  lack of flexibility
  - A small part of the available tools
  - distributed on different universities locations
  - the submission size is often limited
  - must not be paranoid





## Introduction

#### library(xcms)

#### loaddata() polare-"PoCommand line tools

#### noise=250000

xset <- xcmsSet(cdffiles,ppm=ppm, mzdiff=mzwid, peakwidth=peakwidth,\_noise=noise,\_snthresh=snth, method="centWave", fitgauss=TRUE, nSlaves=8)</pre> xset2<-retco+xrepresent almost the majority of scientific tools dev.copy2pdf(devi "), paper="a4", height=9, width=14) xset3<-group(xset2, minfrac = 0.2, bw = bw, minsamp = 1, mzwid = mzwid, max = 50, sleep = 0)</pre> xset5<-fillPea

### + good parameters completeness

reporttab <- diffreport(xset5, filebase =paste(pathResult,"/Rapport\_",expe,"\_",polar, sep=""), mzdec=4, eicmax=5000, metlin = metlin, classeic=levels(xset5@phenoDa

### + can be executed on high performance computers

dir.create(paste(pathResult,"/Rapport\_",expe,"\_",polar,"\_diffreport/", sep=""), showWarnings = FALSE)

write.table(report table) paste(pathResult "Rapport "experimental polar "diffreport/resultat "experimental polar,".xls", sep=""), sep="\t") + g33KS love it, since automatable, workflowsable, ...

#### library(CAMERA)

#### and-annotate(reminimum alinux knowledge is required = 3, maxiso=4, minfrac=0.5, ppm=15, mzabs=0

#### diffreport1<-getPeakl; <sup>e</sup>cruel lack of ergonomics

#diffreport <- annotateDiffreport(xsg,pval\_th=0.05,fc=0.1, nSlaves=8, calcIso=TRUE, calcCaS=FALSE, maxcharge=3, maxiso=4, minfrac=0.5, # ppm=15, mzabs=0.015, quick=FALSE, psg\_list=NULL, rules=NULL, # polarity=polarity, sortpval=FALSE) diffreport<-cbind(reporttab,diffreport1[,c("isotopes", "adduct","pcgroup")])</pre> write.table(diffreport, file=paste(pathResult,"/result\_",expe,"\_",polar,"\_CAMERA\_diffreport-fast.xls",sep=""), row.names=FALSE, sep="\t")

```
library(FactoMineR)
pca3<-PCA(t(matacp), axes=c(1,2))</pre>
pca3<-PCA(t(matacp), axes=c(1,3))</pre>
pca3<-PCA(t(matacp), axes=c(2,3))</pre>
pca4<-PCA(t(matacplog2))</pre>
```

```
png("percentage_of_variance.png", width =800, height = 400);
barplot(resPCA$eig$per,xlab="Components",ylab="percentage of variance");
dev.off()
```

```
png("eigenvalue.png", width =800, height = 400);
barplot(resPCA$eig$eig,xlab="Components",ylab="eigenvalue");
dev.off()
```

```
library(ctc)
if (normalization) {
    data=t(scale(t(data)))
```

3



## **INTRODUCTION / GALAXY**

💳 Galaxy / 4 / M	etabolomics Analyze Data Workflow Shared Data - Visualization - Admin Help - User -		Using -993344424 b
Tools	xcms xcmsSet version 2.0.1	History	C 🌣 🗆
search tools	Choose your inputs method:	search datasets	8
Upload File from your computer	Zip file from your history containing your chromatograms	Sacuri Zip	
Export Data	Zin file:	19 shown	
LC-MS		289.7 MB	
Format Conversion		19:	• A ×
Preprocessing	Extraction method for peaks detect	xset.group.retcor.gr	roup.fillPeaks.anno
Normalisation	matchedFilter :	tate.variableMetadat	ta.tsv (Xdiffreport)
Quality Control		<u>18:</u>	<b>()</b>
Statistical Analysis	Step size to use for profile generation:	xset.group.retcor.gr	roup.fillPeaks.anno
Annotation	0.01	tate.negative.Rdata	
00.110	[step] The peak detection algorithm creates extracted ion base peak chromatograms (EIBPC) on a fixed step size	17:	• A X
GC-MS	Full width at half maximum of matched filtration gaussian model peak:	xset.group.retcor.gr	oup.fillPeaks.anno
Normalization	30	tate.dataMatrix.tsv	
	[fwhm] Only used to calculate the actual sigma	16:	• A X
Statistical Analysis	Advanced options:	xset.group.retcor.gr	oup.fillPeaks.anno
Annotation	hide ‡	tate.variableMetada	ta.tsv
Amotation		15:	
NMR	Execute	xset.aroup.retcor.ar	oup.fillPeaks.RDat
Preprocessing		<u>a</u>	
Normalisation	<b>1</b> Authors Colin A. Smith csmith@scripps.edu. Ralf Tautenbahn.rtautenb@gmail.com. Steffen Neumann.sneumann@ipb-halle.de. Paul	14.	
Quality Control	Benton hpaul.benton08@imperial.ac.uk and Christopher Conley cjconley@ucdavis.edu	xset.group.retcor.gr	roup.Rplots.pdf
Statistical Analysis		Notigrouphotoong	
COMMON TOOLS	If you use this tool, please cite: Smith,C.A. et al.(2006). XCMS: processing mass spectrometry data for metabolite profiling using nonlinear peak alignment matching, and identification. Anal. Chem. 78, 779–787	<u>13:</u>	۲ 🖉 🕐
Data Handling	For details about this tool, please go to <u>http://www.bioconductor.org/packages/release/bioc/html/xcms.html</u>	xset.group.retcor.gr	roup.RData
Text Manipulation		<u>12:</u>	🖲 🖋 🗙
Filter and Sort	<b>Galaxy integration</b> ABIMS TEAM, Station biologique de Roscoff.	xset.group.retcor.BF	PCs_corrected.pdf
Join, Subtract and Group		11:	
<	Contact <u>support@workflow4metabolomics.org</u> for any questions or concerns about the Galaxy implementation of this tool.		>

- Galaxy it's ...
  - No need to execute a command line through a terminal
  - Programming or scripting skills are not required
  - Submission of jobs is transparent through a high performance computer cluster
  - Secure histories and data manager
  - A data and protocols sharing system
  - Tool-boxes of several bioinformatics fields
    - NGS

- Chemistry
- Metabolomics Image analysis
- Statistics Etc ...
- A web-based interface





# Why Galaxy ? –Accessibility –Reproductibility –Transparency



## **MR.GEEK**





[login@n0 mapping]\$ qsub -q long.q -pe thread 8 tophat.qsub Your job 5338969 ("tophat.qsub") has been submitted





- ogin@n0 ~]\$ cdpro
- [login@nU login]\$ cd l3-U/-29-panda/tmp/map
- #!/bin/bash
- #\$ -S /bin/bas
- #\$ -M login@sb-roscoff.fr
- \$ -m be
- #\$ −V
- #\$ -Cv
- \$ -o qsub.
- \$ -e qsub.er

tophat2 panda v121029 ../input/IllR1-1.fq ../input/IllR1-2.fq -GTF ../input/panda\_v121029.gtf --b2-sensitive -r 100 -num-threads 8

[login@n0 mapping]\$ qsub -q long.q -pe thread 8 tophat.qsub Your job 5338969 ("tophat.qsub") has been submitted











[lecorguille@n0 ~]\$ e-PCR --help e-PCR: invalid option -- usage: [-hV] [posix-options] stsfile [fasta ...] [compat-options] where posix-options are: -m ## Margin (default 50) -w ## Wordsize (default 7) Max mismatches allowed (default 0) -n ## -g ## Max indels allowed (default 0) Use ## discontiguos words, slow if -f ## ##>1 -0 ## Set output file -t ## Set output format: 1 - classic, range (pos1..pos2) 2 - classic, midpoint 3 - tabular 4 - tabular with alignment in comments (slow) -d##-## Set default size range (default 100-350) Turn hits postprocess on/off -p +-Verbosity flags -v ## Use presize alignmens (only if -a a|f gaps>0), slow

> a - Allways or f - as Fallback -x +- Use 5'-end lowercase masking of primers (default -)

-u +- Uppercase all primers (default -)

#### [...]

#### 💳 Galaxy / ABiMS

#### e-PCR (version 1.0.0)

#### STS file:

100: (as tabular) Trinity on data 9..Transcripts

#### Fasta file:

100: Trinity on data 9.. Transcripts 🔺

#### format : fasta

Wordsize (W):

#### 7

Set word size for primers hash (nucleotide positions). Longer word size decreases hash collision rate, but increases memory usage. Also no mismatches are allowed within word size near 'inner' boundary of primers unless one uses discontiguous words, and no gaps are ever allowed in that region.

#### Use ## discontinuos words (F):



Set discontiguous word count for primers hash (1 means 'use contiguous words'). Discontiguous words increase number of hash tables and decrease 'effective' word size (thus increasing hash collision rate), so make search significantly slower, but increase sencitivity by allowing mismatches within word size. Reasonable values are 1 (contiguous words) and 3.

#### Margin (M):

	-	-
50		

Set maximal allowed deviation of hit product size from expected STS size.

#### Set default sts lower size (D):



Set ddefault STS size range - values used for STSs that have no size associated in file.

#### Set default sts higher size (D):

400

Set ddefault STS size range - values used for STSs that have no size associated in file.

#### Max mismatches allowed (N):

0

Set maximal number of mismatches allowed in primer-to-sequence alignment (per primer!).

#### Max indels allowed (G):

Set maximal number of gaps allowed in primer-to-sequence alignment (per primer!).

#### Set output format (T):

tabular

Output formats

Execute



### xcmsSet.matchedFilter(object, fwhm = 30, sigma = f

#### Arguments

object	xcmsRaw object
fwhm	full width at half maximum of matched filtration gaussian model j used to calculate the actual sigma, see below.
sigma	standard deviation (width) of matched filtration model peak
max	maximum number of peaks per extracted ion chromatogram
snthresh	signal to noise ratio cutoff
step	step size to use for profile generation
steps	number of steps to merge prior to filtration
mzdiff	minimum difference in m/z for peaks with overlapping retention tim
index	return indicies instead of values for m/z and retention times
sleep	number of seconds to pause between plotting peak finding cycles
scanrange	scan range to process

#### Galaxy / 4 / Metabolomics

Execute

Analyze Data Workflow

Zip file Zip file	e your inputs method e from your history containing your chromatograms
Zip file Zip fi	e from your history containing your chromatograms
Zip fi	
Ľ	ile
	省 🗅 No no_unzip.zip dataset available.
xtrac	tion method for peaks detection
matche	edFilter
metho	d] See the help section below
Step	size to use for profile generation
0.01	1
[step]	] The peak detection algorithm creates extracted ion base peak chromatograms (EIBPC) on a f
Full v	width at half maximum of matched filtration gaussian model peak
30	
[fwhn	n] Only used to calculate the actual sigma
Adva	inced options
show	v
Max	ximum number of peaks per extracted ion chromatogram
5	
[ma	ax]
Sig	nal to noise ratio cutoff
10	D
[snt	thresh]
Nur	mber of steps to merge prior to filtration
2	

💳 Galaxy / 4 / Metabo	<b>Olomics</b> Analyze Data Workflow Shared Data - Visualization - Admin Help - User -		Using -993344424 b
Tools	Batch_correction (version 2.0.0)	History	C 🕈 🗆
search tools	Data Matrix file : 🗅 🖓	search datasets	8
Upload File from your computer	17: xset.group.retcor.group.fillPeaks.annotate.dataMatrix.tsv   2	Sacuri Zip	
Export Data	Sample metadata file : 🗅 🖄	19 shown	
LC-MS	3: sampleMetadata.tsv ‡	289.7 MB	
Format Conversion	must contain at least the three following columns: 'batch' + 'injectionOrder' + 'sampleType'	<u>19:</u>	• / ×
Preprocessing	Variable metadata file : 🗅 🖓	xset.group.retcor.gr	oup.fillPeaks.anno
Normalisation	16: xset.group.retcor.group.fillPeaks.annotate.variableMetadata.tsv 🛫	tate.variableMetadat	a.tsv (Xdiffreport)
Batch_correction Corrects	Type of regression model :	<u>18:</u>	• 🖋 🗙
intensities for signal drift and batch-	linear ‡	xset.group.retcor.gro	oup.fillPeaks.anno
Determine, batch, correction to	To select between linear or non-linear (lowess or loess) methods to be used in Van der Kloet algorithm ; when using loess, you can	tate.negative.Rdata	
choose between linear, lowess and	choose to use pools or samples to model batch effect.	<u>17:</u>	۰ 🖋 🗶
loess methods	Factor of interest :	xset.group.retcor.gro	oup.fillPeaks.anno
Transformation Transforms the	batch		
dataMatrix intensity values	column name of factor of interest (often a biological factor); if none, leave 'batch'	<u>16:</u>	۷ ک
Quality Control	Level of details for plots :	xset.group.retcor.gro	oup.fillPeaks.anno
Statistical Analysis	basic ‡	tate.variablemetadata	a.tsv
Annotation	Amount of plots in the pdf file output. See Help section for more details.	<u>15:</u>	• * ×
GC-MS	Execute	xset.group.retcor.gro	oup.fillPeaks.RDat
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Quality Control	1 Authors	xset.group.retcor.gro	oup.Rpiots.pdf
Statistical Analysis	Jean-Francois Martin - PF MetaToul-AXIOM ; INRA ; MetaboHUB (for original version of this tool and overall development of the R	<u>13:</u>	• 💉 🗙
Annotation	script)	xset.group.retcor.gro	oup.RData
NMR	1 Contributors	<u>12:</u>	• # ×
Preprocessing	Melanie Petera - PFEM ; INRA ; MetaboHUB (for R wrapper and R script improvement)	xset.group.retcor.BP	PCs_corrected.pdf
Normalisation		11:	@ # ¥ ·
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### Menu

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Tools	Batch_correction (version 2.0.0)	History	; <b>*</b> 🗆
search tools	3 Data Matrix file : 🗅 🗠	search datasets	8
Upload File from your computer	17: xset.group.retcor.group.fillPeaks.annotate.dataMatrix.tsv Sample metadata file: Pt 企	Sacuri Zip 19 shown	
LC-MS Format Conversion Preprocessing Normalisation Batch_correction Corrects intensities for signal drift and batch effects Determine_batch_correction to choose between linear, lowess and loess methods Transformation Transforms the dataMatrix intensity values	3: sampleMetadata.tsv   must contain at least the three following columns: 'batch' + 'injectionOrder' + 'sampleType'   Variable metadata file :    Variable metadata file :    16: xset.group.retcor.group.fillPeaks.annotate.variableMetadata.tsv   ilinear   inose for regression model :   Income   Income   Income   Income   Income   Income   Variable metadata file :    Income    Income   Incom	289.7 MB	<ul> <li>×</li> <li>×&lt;</li></ul>
Quality Control Statistical Analysis Annotation GC-MS	Level of details for plots :         basic       ;         Amount of plots in the pdf file output. See Help section for more details.         Execute	xset.group.retcor.group.fillPea tate.variableMetadata.tsv 15: xset.group.retcor.group.fillPea a	ks.anno
Preprocessing Normalisation Quality Control Statistical Analysis Annotation	Authors     Jean-Francois Martin - PF MetaToul-AXIOM ; INRA ; MetaboHUB (for original version of this tool and overall development of the R     script)	14:     Image: Comparison of the sector of the	<ul> <li> <i>P</i> <li> <i>P</i> <li> <i>P</i> </li> </li></li></ul>
NMR Preprocessing Normalisation	Contributors     Melanie Petera - PFEM ; INRA ; MetaboHUB (for R wrapper and R script improvement)     Etienne Thevenot - LIST/LADIS ; CEA ; MetaboHUB (for R script and wrapper concerning "all loess pool" and "all loess sample"     methods)	12: xset.group.retcor.BPCs_correc 11:	ted.pdf

### Tool list

Batch_correction (version 2.0.0)	istory	<i>C</i> 🕈 🗆	נ
search tools 😢 🗍 Data Matrix file : 🗅 🖄	search datasets	8	
Upload File from your computer       17: xset.group.retcor.group.fillPeaks.annotate.dataMatrix.tsv       Sau         Export Data       Sample metadata file : D       2	<b>acuri Zip</b> 9 shown		
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### Web forms / visualization / diverse information

🗧 Galaxy / 4 / M	etabolomics Analyze Data Workflow Shared Data - Visualization - Admin Help - User -		Using -993344424 t
Fools	Batch_correction (version 2.0.0)	History	C 🕈 🗆
search tools	Data Matrix file : D	search datasets	
Upload File from your computer Export Data LC-MS Format Conversion Preprocessing Normalisation Batch_correction Corrects intensities for signal drift and ba effects Determine_batch_correction to choose between linear, lowess loess methods	17: xset.group.retcor.group.fillPeaks.annotate.dataMatrix.tsv   Sample metadata file :    2:   3: sampleMetadata.tsv   must contain at least the three following columns: 'batch' + 'injectionOrder' + 'sampleType'   Variable metadata file :    2:   16: xset.group.retcor.group.fillPeaks.annotate.variableMetadata.tsv :   Type of regression model :   linear :   To select between linear or non-linear (lowess or loess) methods to be used in Van der Kloet algorithm ; when using loess, you can choose to use pools or samples to model batch effect.	Sacuri Zip 19 shown 289.7 MB <u>19:</u> <u>xset.group.retcor.gro tate.variableMetadata</u> <u>18:</u> <u>xset.group.retcor.gro tate.negative.Rdata</u> <u>17:</u> <u>xset.group.retcor.gro tate.dataMatrix.tsv</u>	Image: Second state
<u>Transformation</u> Transforms the dataMatrix intensity values <u>Quality Control</u> <u>Statistical Analysis</u> <u>Annotation</u> GC-MS	column name of factor of interest (often a biological factor); if none, leave 'batch'  Level of details for plots :  basic ;  Amount of plots in the pdf file output. See Help section for more details.  Execute	16:         xset.group.retcor.group.ret	
Preprocessing Normalisation Quality Control Statistical Analysis Annotation	Authors     Jean-Francois Martin - PF MetaToul-AXIOM ; INRA ; MetaboHUB (for original version of this tool and overall development of the R     script)	<u>14:</u> xset.group.retcor.gro <u>13:</u> xset.group.retcor.gro	<ul> <li>Image: A state of the state of the</li></ul>
NMR Preprocessing Normalisation	Contributors     Melanie Petera - PFEM ; INRA ; MetaboHUB (for R wrapper and R script improvement)     Etienne Thevenot - LIST/LADIS ; CEA ; MetaboHUB (for R script and wrapper concerning "all loess pool" and "all loess sample"     methods)	12: xset.group.retcor.BP	Cs_corrected.pdf

### History

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Tools	Batch_correction (version 2.0.0)	History	C 🕈 🗆	
search tools	Data Matrix file : 🗅 🖄	search datasets	8	1
Upload File from your computer	17: xset.group.retcor.group.fillPeaks.annotate.dataMatrix.tsv	Sacuri Zip		
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Normalisation	16: xset.group.retcor.group.fillPeaks.annotate.variableMetadata.tsv 💲	tate.variableMetadata	.tsv (Xdiffreport)	
Batch_correction Corrects	Type of regression model :	<u>18:</u>		
effects	linear ‡	xset.group.retcor.grou	up.fillPeaks.anno	
Determine batch correction to	To select between linear or non-linear (lowess or loess) methods to be used in Van der Kloet algorithm ; when using loess, you can choose to use pools or samples to model batch effect	47		
choose between linear, lowess and		<u>17:</u>	🕑 🖋 🗙	
loess methods	batch	tate.dataMatrix.tsv	ap.mireaks.amo	
Transformation Transforms the	column name of factor of interest (often a biological factor); if none, leave 'batch'	16:		
	Level of details for plots :	xset.group.retcor.grou	up.fillPeaks.anno	
Statistical Analysis	basic t	tate.variableMetadata	<u>.tsv</u>	٢
Annotation	Amount of plots in the pdf file output. See Help section for more details.	<u>15:</u>	• / ×	
		xset.group.retcor.grou	up.fillPeaks.RDat	
GC-MS	Execute	<u>a</u>		
Normalisation		<u>14:</u>	• 🖋 🗙	
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Statistical Analysis	Jean-Francois Martin - PF MetaToul-AXIOM ; INRA ; MetaboHUB (for original version of this tool and overall development of the R	<u>13:</u>	👁 🖋 🗙	
Annotation	script)	xset.group.retcor.grou	up.RData	
NMR	Contributors	<u>12:</u>	• / ×	
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## **GET HELP**

Station Biologique Roscoff Gethelp

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NGS: Picard	Colour is an open web based platform for data intensive biamedical research. The Colour team is a part of BV at Depp		
NGS: QC and manipulation	State, and the Biology and Mathematics and Computer Science departments at Emory University. The Galaxy Project is		
NGS: SAM Tools	supported in part by <u>NHGRI</u> , <u>NSF</u> , <u>The Huck Institutes of the Life Sciences</u> , <u>The Institute for CyberScience at Penn State</u> ,		
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NGS: Picard	Colovy is an open, web based platform for data intensive biomedical research. The Colovy team is a part of PV at Depp		
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## **DATA IMPORT**



# DATA IMPORT < 2 GO</pre>

### Data import < 2 Go



### Data import < 2 Go



### Data import < 2 Go

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# STEP 1: CHOOSE A FTP CLIENT



# **STEP 1: CHOOSE A FTP CLIENT**



# **STEP 1: CHOOSE A FTP CLIENT**





# Step 2: Easy! DATA IMPORT > 2 GO











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# Exercise DATA IMPORT







- Exercise
  - Fetch this file

# http://tinyurl.com/GI-input0

- 1. First, as you want
- 2. Then, consider that it is >2 Go


## **DATA IMPORT**

#### Data import

#### For HUGE public resources: genome, databank ...

#### --> Make a request to the support team

💳 Galaxy / ABiMS	Analyze Data Workflow Shared Data - Visualization - Help - User -		Using 42%
Tools	NCBI BLAST+ blastn (version 0.0.17)	-	History 2 •
search tools			Unnamed history
<u>Get Data</u>			0 bytes 🖉 📄
<ul> <li><u>Upload File</u> from your computer</li> </ul>	Subject database/sequences:		Your history is empty. Click 'Get Data' on the left pane to start
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FASTA manipulation			
Join, Subtract and Group	Execute		
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<u>NCBI BLAST+ blastn</u> Search	Note. Database searches may take a substantial amount of time. For large input datasets it is advisable to allow overnight processing.		
nucleotide database with nucleotide query sequence(s)	L		
<u>NCBI BLAST+ blastp</u> Search	What it does		
protein database with protein query sequence(s)	Search a nucleotide database using a nucleotide query, using the NCBI BLAST+ blastn command line tool. Algorithms include blastn, megablast, and discontiguous megablast.		
<ul> <li><u>NCBI BLAST+ blastx</u> Search protein database with translated nucleotide query sequence(s)</li> </ul>	You can also search against a FASTA file of subject nucleotide sequences. This is not advised because it is slower (only one CPU is used), but more importantly gives e-values for pairwise searches (very small e-values which will look overly significant). In most cases you should instead turn the other FASTA file into a database first using makeblastdb and search against that.		
NCBI BLAST+ tblastn Search			
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# TOOLS

## Tools - panel

💳 Galaxy / ABiMS	Analyze Data Workflow Shared Data - Visualization - Help - User -		Using 41%
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search tools	Nucleotide query sequence(s):	Human protein study 5.3 MB	0 🖻
<ul> <li><u>Upload File</u> from your computer</li> </ul>	1: human_protein.fas	2: chr22 check.gff3	• / %
ABIMS WORKFLOWS	FASTA file from your history (see warning note below)	<u>1: human_protein.fas</u>	• / %
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Join, Subtract and Group Filter and Sort NCBI BLAST+	Note. Database searches may take a substantial amount of time. For large input datasets it is advisable to allow overnight processing.		
<ul> <li><u>NCBI BLAST+ blastn</u> Search nucleotide database with nucleotide query sequence(s)</li> </ul>	What it does Search a protein database using a translated nucleotide guery, using the NCBI BLAST+ blastx command line tool.		
<ul> <li><u>NCBI BLAST+ blastp</u> Search protein database with protein query sequence(s)</li> </ul>	You can also search against a FASTA file of subject protein sequences. This is not advised because it is slower (only one CPU is used), but more importantly gives e-values for pairwise searches (very small e-values which will look overly significiant). In most cases you should instead turn the other FASTA file into a database first using makeblastdb and search against that.		
<ul> <li><u>NCBI BLAST+ blastx</u> Search protein database with translated nucleotide query sequence(s)</li> </ul>	Output format		
<u>NCBI BLAST+ tblastn</u> Search	Because Galaxy focuses on processing tabular data, the default output of this tool is tabular. The standard BLAST+ tabular output contains 12 columns:	76 /	
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## Tools - panel

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<ul> <li><u>NCBI BLAST+ blastp</u> Search protein database with protein query sequence(s)</li> </ul>	<ul> <li>26-04-13: DESeq de</li> <li>26-04-13:</li> </ul>	RNASeq : sam2counts is now available to count the reads coverage by transcrit. It's also a requirement for novo. Metabolomic : Workflow Metabolomic by ABiMS, updated to version 2.0.0 (2013_04_18)	<u>1: human protein.fas</u>	• ( %
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	<u>Galaxy</u> is an open, web-t <u>Biology</u> and <u>Mathematics</u> <u>The Huck Institutes of the</u>	based platform for data intensive biomedical research. The <u>Galaxy team</u> is a part of <u>BX</u> at <u>Penn State</u> , and the <u>a and Computer Science</u> departments at <u>Emory University</u> . The <u>Galaxy Project</u> is supported in part by <u>NHGRI, NSF,</u> <u>Life Sciences, The Institute for CyberScience at Penn State</u> , and <u>Emory University</u> .		
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🗧 Galaxy / 4 / Me	tabolomics Analyze Data Workflow Shared Data - Visualization - Admin Help - User -		Using -993344424 I
Tools	xcms xcmsSet version 2.0.1	History	2 <b>*</b> 🗆
search tools	Choose your inputs method:	search datasets	8
Upload File from your computer	Zip file from your history containing your chromatograms 💲	Sacuri Zip	
Export Data	Zip file:	1 shown	
LC-MS	1: sacuri.zip ‡	191.3 MB	
Format Conversion	Extraction method for peaks detection:	<u>1: sacuri.zip</u>	۵ 🖋 🗶
Preprocessing	matchedFilter :		
xcms.xcmsSet Filtration and Pea	[method] See the help section below		
from xcms R package to preproce	Step size to use for profile generation:		
LC/MS data for relative	0.01		
quantification and statistical analy	sis [step] The peak detection algorithm creates extracted ion base peak chromatograms (EIBPC) on a fixed step size		
xcms.group Group peaks togethe	Full width at half maximum of matched filtration gaussian model peak:		
across samples using overlapping	30		
smoothed peak distributions in	[fwhm] Only used to calculate the actual sigma		
chromatographic time.	Advanced ontions:		
xcms.retcor Retention Time	hide 1		
Correction using retcor function fi	om		
xcms R package	Execute		
xcms.fillPeaks Integrate the signa	lin		
the region of that peak group not			
represented and create a new pe	Authors Colin A. Smith <u>csmith@scripps.edu</u> , Ralf Tautenhahn <u>rtautenh@gmail.com</u> , Steffen Neumann <u>sneumann@ipb-halle.de</u> , Paul		
CAMERA.annotate CAMERA	Benton <u>hpaul.benton08@imperial.ac.uk</u> and Christopher Conley <u>cjconley@ucdavis.edu</u>		
annotation results (isotope peaks	If you use this tool, please cite: Smith,C.A. et al. (2006). XCMS: processing mass spectrometry data for metabolite profiling using		
adducts and fragments) and a	nonlinear peak alignment, matching, and identification. Anal. Chem., 78, 779–787.		
diffreport if more than one conditi	For details about this tool, please go to http://www.bioconductor.org/packages/release/bioc/html/xcms.html		
CAMERA.combinexsAnnos			
Wrapper function for the	<b>Galaxy integration</b> ABIMS TEAM, Station biologique de Roscoff.		
combinexsAnnos CAMERA funct	on. Contact support@workflow4metabolomics.org for any guestions or concerns about the Galaxy implementation of this tool.		
C			>

Galaxy / 4 / Metabolomics Analyze Data Workflow Shared Data - Visualization - Admin Help - User -			Using -993344424
Tools	<b>~</b>	History	S 🕈 🗆
search tools  Choose your inputs method:		search datasets	8
Upload File from your computer		Sacuri Zip	
Export Data		1 shown	
		191.3 MB	
Ec-INS		1: sacuri.zip	@ / X
Extraction method for peaks detection:			
xcms.xcmsSet Filtration and Peak			
Identification using xcmsSet function			
from xcms R package to preprocess Step size to use for profile generation:			
LC/MS data for relative 0.01			
[step] The peak detection algorithm creates extracted ion base peak chromatograms (EIBPC) on a fixed step size			
xcms.group Group peaks together across samples using overlapping			
m/z bins and calculation of 30			
smoothed peak distributions in [fwhm] Only used to calculate the actual sigma			
chromatographic time. Advanced options:			
xcms.retcor Retention Time hide			
Correction using retcor function from			
xcms R package			
xcms.fillPeaks Integrate the signal in			
the region of that peak group not			
Authors Colin A. Smith <u>csmith@scripps.edu</u> , Ralf Tautenhahn <u>rtautenh@gmail.com</u> , Steffen Neumann <u>sneumann@ipb-halle.de</u> , Pa	aul		
CAMERA.annotate CAMERA Benton hpaul.benton08@imperial.ac.uk and Christopher Conley cjconley@ucdavis.edu			
annotation results (isotope peaks.			
adducts and fragments) and a nonlinear peak alignment, matching, and identification. Anal. Chem., 78, 779–787.			
diffreport if more than one condition. For details about this tool, please go to http://www.bioconductor.org/packages/release/bioc/html/xcms.html			
CAMERA.combinexsAnnos			
Wrapper function for the <b>Galaxy integration</b> ABIMS TEAM, Station biologique de Roscoff.			
combinexsAnnos CAMERA function.			
	))	3	

👕 Galaxy / 4 / Metabo	Analyze Data Workflow Shared Data - Visualization - Admin Help - User -		Using -993344424
Tools	xcms.xcmsSet version 2.0.1	History	C 🕈 🗆
search tools	Choose your inputs method:	search datasets	8
Upload File from your computer	Zip file from your history containing your chromatograms $z$	Sacuri Zip	
Export Data	Zip file:	1 shown	
LC-MS	1: sacuri.zip ‡	191.3 MB	
Format Conversion	Extraction method for peaks detection:	<u>1: sacuri.zip</u>	• 🖋 🗙
Preprocessing	matchedFilter :		
xcms.xcmsSet Filtration and Peak	[method] See the help section below		
from xcms R package to preprocess	Step size to use for profile generation:		
LC/MS data for relative	0.01		
quantification and statistical analysis	[step] The peak detection algorithm creates extracted ion base peak chromatograms (EIBPC) on a fixed step size		
xcms.group Group peaks together	Full width at half maximum of matched filtration gaussian model peak:		
across samples using overlapping m/z bins and calculation of	30		
smoothed peak distributions in	[fwhm] Only used to calculate the actual sigma		
chromatographic time.	Advanced options:		
xcms.retcor Retention Time	hide ‡		
xcms R package			
yoms fillDeaks Integrate the signal in	Execute		
the region of that peak group not			
represented and create a new peak	1 Authors Colin A. Smith csmith@scripps.edu, Ralf Tautenhahn rtautenh@gmail.com, Steffen Neumann sneumann@ipb-halle.de, Paul		
CAMERA.annotate CAMERA	Benton hpaul.benton08@imperial.ac.uk and Christopher Conley cjconley@ucdavis.edu		
annotate function. Returns	If you use this tool, please cite: Smith C.A. et al. (2006). XCMS: processing mass spectrometry data for metabolite profiling using		
adducts and fragments) and a	nonlinear peak alignment, matching, and identification. Anal. Chem., 78, 779–787.		
diffreport if more than one condition.	For details about this tool, please go to http://www.bioconductor.org/packages/release/bioc/html/xcms.html		
CAMERA.combinexsAnnos			
Wrapper function for the	Galaxy integration ABIMS TEAM, Station biologique de Roscoff.		
	Contact support@workflow4metabolomics.org for any questions or concerns about the Galaxy implementation of this tool.		
		$\rightarrow$	

#### Tools can have some advanced options

👕 Galaxy / 4 / Metabo	Diomics Analyze Data Workflow Shared Data - Visualization - Admin Help - User -		Using -993344424
Tools	xcms.xcmsSet_version 2.0.1	History	C 🕈 🗆
search tools	Choose your inputs method:	search datasets	8
Upload File from your computer	Zip file from your history containing your chromatograms 💲	Sacuri Zip	
Export Data	Zip file:	1 shown	
LC-MS	1: sacuri.zip ‡	191.3 MB	
Format Conversion	Extraction method for peaks detection:	<u>1: sacuri.zip</u>	• 🖋 🗙
Preprocessing	matched Filter		
xcms.xcmsSet Filtration and Peak	[method] See the help section below		
Identification using xcmsSet function from xcms R package to preprocess	Step size to use for profile generation:		
LC/MS data for relative	0.01		
quantification and statistical analysis	[step] The peak detection algorithm creates extracted ion base peak chromatograms (EIBPC) on a fixed step size		
xcms.group Group peaks together	Full width at half maximum of matched filtration gaussian model peak:		
across samples using overlapping	30		
smoothed peak distributions in	[fwhm] Only used to calculate the actual sigma		
chromatographic time.			
xcms.retcor Retention Time			
Correction using retcor function from	Snow -		
xcms R package	Maximum number of peaks per extracted ion chromatogram:		
xcms.fillPeaks Integrate the signal in	5		
the region of that peak group not	[max]		
represented and create a new peak	Signal to noise ratio cutoff:		
CAMERA.annotate CAMERA			
annotate function. Returns	[onthroch]		
annotation results (isotope peaks,	Isumesul		
adducts and fragments) and a	Number of steps to merge prior to filtration:		
diffreport if more than one condition.	2		
CAMERA.combinexsAnnos	[steps] The peak identification algorithm combines a given number of FIBPCs prior to filtration and peak detection, as defined by the		
Wrapper function for the	steps argument		
combinexsAnnos CAMERA function.			
<	Execute		2

🗧 Galaxy / 4 / Metab	olomics Analyze Data Workflow Shared Data - Visualization - Admin Help - User -		Using -993344424 b
Tools	xcms xcmsSet version 2.0.1	History	C 🕈 🗆
search tools	Choose your inputs method:	search datasets	8
Upload File from your computer	Zip file from your history containing your chromatograms	Sacuri Zip	
Export Data	Zin file:	1 shown	
LC-MS	1: sacuri.zip ‡	191.3 MB	
Format Conversion	Extraction method for peaks detection:	<u>1: sacuri.zip</u>	• 🖋 🗙
Preprocessing	matchedEiter *		
xcms.xcmsSet Filtration and Peak	[method] See the help section below		
Identification using xcmsSet function from xcms R package to preprocess	Step size to use for profile generation:		
quantification and statistical analysis	0.01 [step] The peak detection algorithm creates extracted ion base peak chromatograms (EIBPC) on a fixed step size		
<u>xcms.group</u> Group peaks together across samples using overlapping m/z bins and calculation of	Full width at half maximum of matched filtration gaussian model peak:         30		
smoothed peak distributions in	[fwhm] Only used to calculate the actual sigma		
chromatographic time.	Advanced options:		
xcms.retcor Retention Time	show ¢		
xcms R package	Maximum number of peaks per extracted ion chromatogram:		
xcms.fillPeaks Integrate the signal in the region of that peak group not	5		
represented and create a new peak	[IIIdX]		
CAMERA.annotate CAMERA	Signal to noise ratio cutoff:		
annotate function. Returns	[snthresh]		
annotation results (isotope peaks,			
diffreport if more than one condition.	Number of steps to merge prior to filtration:		
<u>CAMERA.combinexsAnnos</u> Wrapper function for the combinexsAnnos CAMERA function.	[steps] The peak identification algorithm combines a given number of EIBPCs prior to filtration and peak detection, as defined by the steps argument		
<	Execute		>

💳 Galaxy / 4 / Metabo	OMICS Analyze Data Workflow Shared Data   Visualization    Admin Help    User	Us	sing -993344424 b
Tools		History	C 🕈 🗆
search tools	A job has been successfully added to the queue - resulting in the following datasets:	search datasets	8
Upload File from your computer Export Data	3: sampleMetadata.tsv	Sacuri Zip 1 shown	
LC-MS	4: xset.TICs_raw.pdf	191.3 MB	
Format Conversion Preprocessing	5: xset.BPCs_raw.pdf	<u>6: xset.log.txt</u>	
xcms.xcmsSet Filtration and Peak Identification using xcmsSet function	6: xset.log.txt	O <u>4: xset.TICs_raw.pdf</u>	• / ×
from xcms R package to preprocess LC/MS data for relative quantification and statistical analysis	You can check the status of queued jobs and view the resulting data by refreshing the <b>History</b> pane. When the job has been run the status will change from 'running' to 'finished' if completed successfully or 'error' if problems were encountered.		<u>.</u> • * ×
xcms.group Group peaks together		O 2: xset.RData	• 🖋 🗙
m/z bins and calculation of		<u>1: sacuri.zip</u>	• 🖋 🗙
smoothed peak distributions in chromatographic time.			
<u>xcms.retcor</u> Retention Time Correction using retcor function from xcms R package			
<u>xcms.fillPeaks</u> Integrate the signal in the region of that peak group not represented and create a new peak			
<u>CAMERA.annotate</u> CAMERA annotate function. Returns annotation results (isotope peaks, adducts and fragments) and a diffreport if more than one condition.			
CAMERA.combinexsAnnos Wrapper function for the combinexsAnnos CAMERA function			
<			>





Job is waiting to run

## = the job is in the scheduler « queue »

Duration time of this status depends on the amount of actual queued jobs or on the requested number of processors





Job is currently running

= the job is being executed on the computing cluster

Duration time of this status depends completely on the job's attributes and the computing ressources allocated.

Some programs are executed with several processors

(using 4, 8 or 16 Gb of RAM).

And others are mono-threaded ®





## Job is finished

It's status is OK

but warnings or errors can be hidden behind. Ah hum !





Job is finished but with an error status

= the program sends an error

The error is often explained by the program and sometimes ... not.





## Job is finished but with an error status

Error causes :

- The user :P
- Bad usage : input file, format or option
- Wrong porting of the program through Galaxy ... sorry :/
- Non anticipated crash of the program



# Exercise TOOLS







- Aim of this Exercise:
  - Get blast hits with :
    - Identity > 75%
    - Alignment coverage > 75%
  - Execute and chain example of little Galaxy friendly tools together.





- Aim of this Exercise:
  - Get blast hits with :
    - Identity > 75%
    - Alignment coverage > 75%



Coverage = alignment length / query length





- Fetch these two files (< 2Go)
  - Link1:
    - http://tinyurl.com/GI-Roscoff1
  - Link2:
    - http://tinyurl.com/GI-Roscoff2
- Check their contents and datatypes through Galaxy.



- Identity > 75%
- Alignment coverage > 75%







- Identity > 75%
- Alignment coverage > 75%
- Step 1: use the tool « Compute sequence length »





- Identity > 75%
- Alignment coverage > 75%
- Step 1: use the tool « Compute sequence length »
- Step 2: use the tool « Join Two Datasets »





- Identity > 75%
- Alignment coverage > 75%
- Step 1: use the tool « Compute sequence length »
- Step 2: use the tool « Join Two Datasets »
  - help : join with column 1 and column 1





- Identity > 75%
- Alignment coverage > 75%
- Step 1: use the tool « Compute sequence length »
- Step 2: use the tool « Join Two Datasets »
   help : join with column 1 and column 1
- Step 3: use the tool « Compute an expression... »





- Identity > 75%
- Alignment coverage > 75%
- Step 1: use the tool « Compute sequence length »
- Step 2: use the tool « Join Two Datasets »
  - help : join with column 1 and column 1
- Step 3: use the tool « Compute an expression... »
   help : ((c8-c7+1)/c14)\*100





- Identity > 75%
- Alignment coverage > 75%
- Step 1: use the tool « Compute sequence length »
- Step 2: use the tool « Join Two Datasets »
  - help : join with column 1 and column 1
- Step 3: use the tool « Compute an expression... »
   help : ((c8-c7+1)/c14)\*100
- Step 4: use the tool « Filter »





- Identity > 75%
- Alignment coverage > 75%
- Step 1: use the tool « Compute sequence length »
- Step 2: use the tool « Join Two Datasets »
  - help : join with column 1 and column 1
- Step 3: use the tool « Compute an expression... »
   help : ((c8-c7+1)/c14)\*100
- Step 4: use the tool « Filter »

– help : c3>=75 and c15>=75



# Part II TOOLS

🗧 Galaxy / ABiMS	Analyze Data Workflow Shared Data - Visualization - Admin Help - User -		Using 19.1 GB
Tools	Online	History	0 0
Search tools		Copper Stress v3 133.9 MB	47 🖻
ABIMS WORKFLOWS Workflow RNA-seq de novo by ABIMS Workflow RNA-seq with reference by ABIMS	<ul> <li>07-06-13: Metabolomic : Workflow 4 Metabolomics, updated to version 2.1.0 (2013_06_07)</li> <li>30-04-13: RNASeq : DESeq is now available for RNASeq expression data with reference (with gtf input).</li> <li>26-04-13: RNASeq : DESeq is now available for denovo RNASeq expression data (without gtf input).</li> <li>26-04-13: RNASeq : sam2counts is now available to count the reads coverage by transcrit. It's also a requirement for DESeq denovo.</li> </ul>	<u> </u>	● Ø X roup.retcor.gro eport.data ma bular
Workflow 4 Metabolomics ABIMS TOOLS	26-04-13: Metabolomic : Workflow Metabolomic by ABIMS, updated to Version 2.0.0 (2013_04_18)	<u>6:</u> mzXML_copper_stress.g up.fillPeaks.annotateDiffr	
Primer RNASeq InterEsil	4	<u>5:</u> mzXML copper stress.g up.fillPeaks.RData	
<u>Statistics</u> <u>Utils</u> <u>Phylogenetics</u> Debug	ADIM5	<u>4:</u> mzXML copper stress.g up.RData	● Ø X roup.retcor.gro
COMMON TOOLS <u>Text Manipulation</u>	Analyses and Bioinformatics for Marine Science	3: mzXML copper stress.R 2: sampleInfo.tab	<u>(Data</u>
Join, Subtract and Group Filter and Sort	Station Biologique	<u>1:</u> mzXML copper stress.m	● Ø X ns.zip
NCBI BLAST+ NGS: QC and manipulation	Roscoff	6	
NGS: Mapping NGS: Picard (beta) NGS: SAM Tools	Information For any question or request for tools or account, send an email at support.abims 'AT' sb-roscoff.fr		
NGS: GATK Tools (beta) SVDetect VarScan	<u>Galaxy</u> is an open, web-based platform for data intensive biomedical research. The <u>Galaxy team</u> is a part of <u>BX</u> at <u>Penn State</u> , and the <u>Biology</u> and <u>Mathematics and Computer Science</u> departments at <u>Emory University</u> . The <u>Galaxy Project</u> is supported in part by <u>NHGRI</u> , <u>NSF</u> , The Huck Institutes of the Life Sciences. The Institute for CyberScience at Penn State, and Emory University.		

Galaxy / ABiMS	Analyze Data Workflow Shared Data - Visualization - Admin Help - User -	Using 19.1 G	в
Tools	Online	History C 4	•
search tools		Copper Stress v3 133.9 MB	
ABIMS WORKFLOWS Workflow RNA-seq de novo by ABIMS Workflow RNA-seq with reference by ABIMS Workflow 4 Metabolomics ABIMS TOOLS Primer RNASeq InterEsil Statistics	<ul> <li>07-06-13: Metabolomic : Workflow 4 Metabolomics, updated to version 2.1.0 (2013_06_07)</li> <li>30-04-13: RNASeq : DESeq is now available for RNASeq expression data with reference (with gtf input).</li> <li>26-04-13: RNASeq : DESeq is now available to count the reads coverage by transcrit. It's also a requirement for DESeq denovo.</li> <li>26-04-13: Metabolomic : Workflow Metabolomic by ABiMS, updated to version 2.0.0 (2013_04_18)</li> </ul>	© 7: mzXML_copper_stress.group.retcor.gr up.fillPeaks_annotateDiffreport.data m trix.tsv and a pvalue.tabular error An error occurred with this dataset: Fata error: Exit code 10 () ERROR: There is a problem with the group of condition (presence of NA). You may need to use change the mode (column/row) Current groups : NA WA NA NA NA NA NA NA NA NA MA NA NA NA NA NA NA NA NA ma NA NA NA NA NA NA NA NA mathematical statematical sta	
<u>Utils</u> Phylogenetics Debug		<u>6:</u> <u>mzXML copper stress.group.retcor.g</u> <u>up.fillPeaks.annotateDiffreport.Rdata</u>	× ro
Text Manipulation FASTA manipulation	Analyses and Bioinformatics for Marine Science	5: mzXML copper stress.group.retcor.g up.fillPeaks.RData	× <u>ro</u>
Filter and Sort Graphics NCBI BLAST+	Station Biologique Roscoff	4:      The second seco	× ro
NGS: QC and manipulation NGS: RNA Analysis		<u>3:</u> mzXML copper stress.RData ● Ø	×
NGS: Mapping NGS: Picard (beta) NGS: SAM Tools NGS: GATK Tools (beta)	Information For any question or request for tools or account, send an email at support.abims 'AT' sb-roscoff.fr	2: sampleInfo.tab     ●     ∅       1:     ●     ∅       mzXML copper stress.ms.zip     ●	×
SVDetect VarScan	<u>Galaxy</u> is an open, web-based platform for data intensive biomedical research. The <u>Galaxy team</u> is a part of <u>BX</u> at <u>Penn State</u> , and the <u>Biology</u> and <u>Mathematics and Computer Science</u> departments at <u>Emory University</u> . The <u>Galaxy Project</u> is supported in part by <u>NHGRI</u> , <u>NSF</u> , The Work Institutes of the Life Sciences. The Institute for CuberScience at Paper State, and Emory University.		

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🗧 Galaxy / ABil	MS	Analyze Data Workflow Shared Data - Visualization - Admin Help - User -			Using 19.1	GB
Tools		Dataset generation errors	ÔН	listory	0	4
search tools	0	Dataset 7: mzXML_copper_stress.group.retcor.group.fillPeaks.annotateDiffreport.data_matrix.tsv_anova_pvalue.tabular	c	Copper Stress v3		
Get Data		Tool execution generated the following error message:	1	33.9 MB	4	2
		Fatal error: Exit code 10 ()	0	<u>37:</u>	• 6	2 1
Workflow RNA-seq de novo	by ABiMS	ERROR: There is a problem with the group of condition (presence of NA). You may need to use change the mo	<u>n</u>	nzXML copper stre	ss.group.retcor	r.g
Workflow RNA-seq with ref	erence by	Current groups :	E tr	pinipeaks.annotate	<u>Diffreport.data</u>	
ABIMS		NA NA NA NA		rror	<u>c.tubutu</u>	
Workflow 4 Metabolomics		NA NA NA NA NA	A	n error occurred with	this dataset: Fa	ital
		NA NA NA NA NA	e	rror: Exit code 10 () E	RROR: There is	s a
ABIMS TOOLS		NA NA NA NA	p	roblem with the grou	p of condition	
Primer		NA NA NA NA	(1	presence of NA). You	may need to us	e
RNASeq		NA NA NA NA	CI	hange the mode (col	umn/row) Currer	nt
InterEsil		NA NA NA NA		JA NA NA NA NA NA NA	NA NA NA NA NA	VA
Statistics		NA NA NA NA NA Na Na Na Na	A	A NA NA NA NA NA	NA NA NA	
Litile		NA NA NA NA	R	R 🖸 🕗		
Ouis		NA NA NA NA	5			
Phylogenetics		NA NA NA NA NA		new or report this error	• (	2 :
Debug		NA NA NA NA	<u>n</u>	nzXML copper stre	ss.group.retcor	r.g
COMMON TOOLS		NA NA NA NA	<u>u</u>	p.fillPeaks.annotate	Diffreport.Rdata	<u>a</u>
Text Maninulation		NA NA NA NA NA	5		@ /	0
Text Manipulation		NA NA NA NA	<u> </u>	<u>.</u> azXML copper stre	ss group retcor	r a
FASTA manipulation		NA NA NA NA	u u	p.fillPeaks.RData	oolgrouphotool	- Second
Join, Subtract and Group		NA NA NA NA				
Filter and Sort		NA NA NA NA	4	<u>a</u>	•	2 :
Graphics		NA NA NA NA	<u>n</u>	nzXML copper stre	ss.group.retcor	r.gr
NCBI BLAST+		NA NA NA NA NA	<u>u</u>	<u>p.RData</u>		
NGS: OC and manipulation		NA NA NA NA	2		@ /	0 4
NGS: RNA Analysis		NA NA NA NA	n n	- nzXML copper stre	ss.RData	1
NCS: Manning		NA NA NA NA				
NGS: Mapping		NA NA NA NA	2	: sampleInfo.tab	• (	2 :
NGS: Picard (beta)		NA NA NA NA NA				
NGS: SAM Tools		NA NA NA NA NA	1	<u>:</u>	•	2 :
NGS: GATK Tools (beta)		NA NA NA NA	m	IZXML copper stre	<u>ss.ms.zip</u>	
SVDetect		NA NA NA NA				
VarScan		NA NA NA NA				
Searching sequence tools		NA NA NA NA				
		NA NA NA NA				
- laurel	determet (	NA NA NA NA NA		11		T

#### Sent to the support team

🗕 Galaxy / ABiMS	Analyze Data Workflow Shared Data - Visualization - Admin Help - User -		Using 18.1 GB	
Tools	NA NA NA NA	f	History 2 🕈	l
search tools	NA NA NA NA NA NA NA NA		Copper Stress v3	1
<u>Get Data</u>	NA NA NA NA NA NA NA		133.9 MB 🖉 🖻	
ABIMS WORKFLOWS Workflow RNA-seq de novo by ABiMS Workflow RNA-seq with reference by ABIMS Workflow 4 Metabolomics ABIMS TOOLS Primer RNASeq InterEsil Statistics Utils	NA NA NA NA NA NA NA NA NA NA		Image: Construct on the system of the sy	
Phylogenetics	Report this error to the Galaxy Team			=
Debug COMMON TOOLS Text Manipulation	The Galaxy team regularly reviews errors that occur in the application. However, if you would like to provide additional information (suc as what you were trying to do when the error occurred) and a contact e-mail address, we will be better able to investigate your problem and get back to you.	ch a	<u>6:</u> <u>mzXML copper stress.group.retcor.g</u> <u>roup.fillPeaks.annotateDiffreport.Rdat</u> <u>a</u>	
FASTA manipulation	Error Report		<u>5:</u>	
Filter and Sort Graphics NCBI BLAST+ NGS: QC and manipulation	Your email <pre>lecorguille@sb-roscoff.fr</pre> Message		mzXML copper stress.group.retcor.g         roup.fillPeaks.RData         4:         mzXML copper stress.group.retcor.g         roup.RData	
<u>NGS: RNA Analysis</u> <u>NGS: Mapping</u>			<u>3:</u> ■ Ø Ø X mzXML copper stress.RData	
NGS: Picard (beta) NGS: SAM Tools		Ξ	2: sampleInfo.tab	
NGS: GATK Tools (beta)         SVDetect         VarScan         Muscle			<u>1:</u> ● Ø X <u>mzXML copper stress.ms.zip</u> data format: ms_zip, database: <u>?</u>	
RAXML	Report	1	uploaded ms_zip file	•



# HISTORY

## History panel

#### Both inputs and outputs

🗧 Galaxy / 4 / Me	tabol	OMICS Analyze Data Workflow Shared Data → Visualization → Admin Help → User →		Using -993344424	4 t
Tools	1	Batch_correction (version 2.0.0)	History	S 🕈 🗆	]
search tools	8	Data Matrix file : 🗅 🖓	search datasets	8	ŕ
Upload File from your computer		17: xset.group.retcor.group.fillPeaks.annotate.dataMatrix.tsv	Sacuri Zip		
Export Data		Sample metadata file : 🗅 🖓	19 shown		
LC-MS		3: sampleMetadata.tsv ‡	289.7 MB		
Format Conversion		must contain at least the three following columns: 'batch' + 'injectionOrder' + 'sampleType'	<u>19:</u>	👁 🖋 🗙	
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Normalisation		16: xset.group.retcor.group.fillPeaks.annotate.variableMetadata.tsv	tate.variableMetada	ata.tsv (Xdiffreport)	
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## History panel

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## DATASET

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## Informations

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COMMON TOOLS		M110T55	2572.37712822047	13158.4785864824	2853.71333606008	16036.855922733	0	7393.124	xset.group.retcor.group	o.RData	
Data Handling		M111T273	16799.0249907129	130599.823855273	42009.1613629325	78842.8338411576	3967.62968023854	29231.52	10.		
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## Renaming and annotation

Galaxy / 4 / Metabolomics Analyze Data Workflow Shared Data - Visualization - Admin Help - User -		Using -993344424 b
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Edit Attributes         Name:         xset group.retor.group.fillPeaks.annotati         Into:         addings bio_vs_blank.annot.tsv         (deflated 52%)         (deflated 52%)         Annotation / Notes:         Add an annotation or notes to a dataset; annotations are available when a history is viewed.         Database/Build:         unspecified (?)         Save         Auto-detect         This will inspect the dataset and attempt to correct the above column values if they are not accurate.	search datasets Sacuri 19 shown 289.7 MB 19: xset.group.retcor.group tate.variableMetadata.ts 18: xset.group.retcor.group tate.negative.Rdata 7.5 MB format: rdata.camera.ne database: ? adding: bio_vs_blank.bd adding: bio_vs_blank.bd (deflated 52%) adding: bio_vs_blank.bd (deflated 23%) addi	Image: system of the system of th
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## Change the Datatype of the Dataset

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		Q	set but <i>not</i> modify it	s contents. Use t	his if Galaxy I	nas incorrectly gue	essed the type of y	our dataset.		13:	• * *	
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## Graphics



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• Data processing (re	quires 'charts' tool fi	rom Toolshed)				
		I I I				

## Graphics



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Dataset

J.

## Re-run a job

💳 Galaxy / 4 / Me	tabol	Omics Analyze Data Workflow Shared Data - Visualization - Admin Help - User -	===	Using -993344424
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galaxy4metabolomics.sb-roscoff.fr/tool_run	nner/rerun?id=	[ppm] =89e90f36310e084c		



# Cleanup DATASET



## Dataset / Cleanup

### Delete a dataset

💳 Galaxy / 4 / Me	etabol	OMICS Analyze Data Workflow Shared Data - Visualization - Admin Help - User -			Using -993344424 I
Tools	1	CAMERA.annotate (version 2.0.0)	<b>→</b> Î	History	C 🕈 🗆
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Filter and Sort		5		Aser.group.recol.BPC	os_corrected.pdf
Join, Subtract and Group		[ppm]		11:	

## Dataset / Cleanup

## The dataset isn't really deleted

## It's in the Trash

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Join, Subtract and Group	<b>.</b>	[ppm]		



## Dataset / Cleanup

### "Empty Trash" : to free up disk space





# WORKFLOW



• A workflow is a sequence of tool operations and parameters

• Can match the experiment protocol

 A workflow is built to be replayed (more or less strict)



• Our workflow



#### Our workflow with Galaxy



#### From a history

RAXML

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#### From a history



## From a history

💳 Galaxy / 4 / Meta	abo	lomics	Analyze Data Wor	kflow Shared Data		Visualization – Admin Help – User –			Using -993344424	4 b
Tools	1	The following list contains	s each tool that was r	un to create the datas	sets	in your current history. Please select those that you wish to include in	9	History	C 🕈 🗆	J
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### Workflow manager

💳 Galaxy / 4 / Metabolomics	Analyze Data Workflow	Shared Data  ▼ Visualization  ▼	Admin Help▼ User▼		Using -993344424 t
Your workflows	1			Create new workflow	1 Upload or import workflow
Name				# of Step	S
LS-MS -				7	
Copy of 'gigaXml' shared by 'ethevenot@sb-roscoff.fr' -				13	
Workflow LC/MS -				6	
Community -				10	
Full_workflow -				19	
Workflow XCMS -				8	

#### Workflows shared with you by others

Name	Owner	# of Steps
demo_workflow_06_annotation -	mlandi@sb-roscoff.fr	6
<u>cohort</u> •	ethevenot@sb-roscoff.fr	15
gigaRaw-convert ▼	ethevenot@sb-roscoff.fr	1

#### **Other options**

Configure your workflow menu

#### Edit a workflow

<b> Galaxy / 4 /</b> ]	Metabolomics	Analyze Data	Workflow	Shared Data <del>-</del>	Visualization <del>-</del>	Admin	Help 🗸	User <del>-</del>			Using -993344424 t
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Share or Publish									6		
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Rename									10		
View									19		
Delete									8		

#### Workflows shared with you by others

Name	Owner	# of Steps
demo_workflow_06_annotation	mlandi@sb-roscoff.fr	6
<u>cohort</u> •	ethevenot@sb-roscoff.fr	15
gigaRaw-convert ▼	ethevenot@sb-roscoff.fr	1

#### **Other options**

Configure your workflow menu

## Edit a workflow : drag and drop

Galaxy / METABO	Analyze Data	Workflow	Shared Data 🕶	Visualization	← Admin	Help <del>-</del>	User▼			Using 7.8 MB
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3-Normalisation										
4-Quality Control										
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## Edit a workflow : drag and drop

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<u>4-Quality Control</u> <u>5-Statistical Analysis</u>		output (rdata) rplots (pdf)				
<u>6-Annotation</u>		tics_cor (pdf)	00			
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Text Manipulation						
Filter and Sort	xcms.xcmsSet 🗙	xcms.group	×	xcms.group	×	xcms.fillPeaks
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## Edit a workflow : delete a noodle

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Search tools         Get Data         WORKFLOW 4 METABOLOMICS         2.Preprocessina         3-Normalisation         4-Daalily Control         S-Statistical Analysis         6-Annotation         COMMON TOOLS         Text Manipulation         Filter and Sort         Doin. Subtract for Group         Statistics         Graph/Display. Data         Multiple regression         Workflow control         Inputs	Tools	Workflow Canvas   Workflow XCM	IS		0
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## Edit a workflow : add a tool

💳 Galaxy / METABO	Analyze Data Wor	kflow Shared Data	🕶 Visualization 🕶 Admin Help 🕶 User 🕶	 Using 7.8 MB
Tools	Workflow Canvas   Workflow XCMS	5		¢
<ul> <li>search tools</li> <li><u>Get Data</u></li> <li>WORKFLOW 4 METABOLOMICS</li> <li><u>2-Preprocessing</u></li> <li><u>xcms.xcmsSet</u> Filtration and Peak Identification using xcmsSet function from xcms R package to preprocess LC/MS data for relative quantification and statistical analysis</li> <li><u>xcms.qroup</u> Group peaks together across samples process and the process</li> </ul>	xcms.xcmsSet ×	xcms.retcor RData file output (rdata) rplots (pdf) tics_cor (pdf) log (txt) xcms.group		xcms.fillPeaks
<ul> <li>using overlapping m/z bins and calculation of smoothed peak distributions in chromatographic time.</li> <li>xcms.retcor Retention Time Cot 2ction using retcor</li> </ul>	output (rdata) O output_info (tabular) O tics_raw (pdf) O log (txt) O	<ul> <li>RData file</li> <li>output (rdata)</li> <li>rplots (pdf)</li> <li>log (txt)</li> </ul>	xcms.retcor     x       %     loading tool info       rplots (pdf)     0       log (txt)     0	RData file output (rdata) log (txt)
<ul> <li>xcms.fillPeaks Integrate the signal in the region of that peak group not represented and create a new peak</li> <li>xcms.diffreport A report showing the most statistically significant differences in analyte intensities</li> <li>CAMERA.annotateDiffreport</li> </ul>				
Wrapper function for the galaxy4metabolomics.sb-roscoff.fr/workflow/ed	litor?id=c1992d25d70f8c1f#			<
		ahi shaki shaki shaki	a santa santa santa banta banta panta panta panta panta panta. Pan	

## Edit a workflow : add a noodle

Galaxy / METABO	Analyze Data <b>Wo</b>	rkflow Shared Data <del>-</del> Visuali	zation <del>▼</del> Admin Help <del>▼</del> User	-	Using 7.8 MB
Tools	Workflow Canvas   Workflow XCM	IS			0
Search tools		xcms.retcor 🗙	xcms.retcor 🗙		
2-Preprocessing • <u>xcms.xcmsSet</u> Filtration and Peak Identification using xcmsSet function from xcms R package to preprocess LC/MS data for relative quantification and statistical analysis		RData file output (rdata) rplots (pdf) tics_cor (pdf) log (txt)	RData file output (rdata) rplots (pdf) tics_ror (pdf) log (txt)		
<ul> <li>xcms.group Group peaks together across samples using overlapping m/z bins and calculation of smoothed peak distributions in chromatographic time.</li> <li>xcms.retcor Retention Time Correction using retcor function from xcms R package</li> <li>xcms.fillPeaks Integrate the signal in the region of that</li> </ul>	xcms.xcmsSet     ×       output (rdata)     2       output_info (tabular)     2       tics_raw (pdf)     2       log (txt)     2	xcms.group × RData file output (rdata) rplots (pdf) log (txt)	xcms.group RData file output (rdata) rplots (pdf) log (txt)	xcms.group x RData file output (rdata) 0 rplots (pdf) 0 log (txt) 0 x	xcms.fillPeaks RData file output (rdata) log (txt)
<ul> <li>signal in the region of that peak group not represented and create a new peak</li> <li>xcms.diffreport A report showing the most statistically significant differences in analyte intensities</li> <li><u>CAMERA.annotateDiffreport</u> Wrapper function for the </li> </ul>					

Edit a workflow : set or release a parameter



## Run a workflow

💳 Galaxy / METABO	Analyze Data Workflow Shared Data - Visualization - Admin Help - User -				Jsing 7.8 M	в
Tools	Rupping workflow "I S-MS"		History		0	¢
search tools	Step 1: years years Set (yearsion 2.0.1)		sacuri			
<u>Get Data</u>	Choose your inputs method		0 bytes		Q 🗹 📎 🦻	
WORKFLOW 4 METABOLOMICS	Zip file from your history containing your chromatograms		1 This	history is empt	y. You can	
2-Preprocessing	Zip file		load	your own data	or <u>get</u>	
3-Normalisation	1: sacuri.zip 🗸	111	Udla	a nom an exten	lai source	
4-Quality Control	Extraction method for peaks detection					
5-Statistical Analysis	matchedFilter					
6-Annotation	Step size to use for profile generation					
COMMON TOOLS						
Text Manipulation	Full which at half maximum of matched filtration gaussian model peak $4 \left[ \mathscr{E} \right]$					
Filter and Sort	Advanced entions					
Join, Subtract and Group	show					
Statistics	Maximum number of neaks per extracted ion chromatogram					
Graph/Display Data						
Multiple regression	Signal to noise ratio cutoff					
Workflows	3 🕜					
<ul> <li>All workflows</li> </ul>	Number of steps to merge prior to filtration					
	2 🗷					
	Step 2: xcms.group (version 2.0.1)					
	Output dataset 'xsetRData' from step 1					
	Method to use for grouping					
	density					
	Bandwidth					
	30					
	Minimum fraction of samples necessary		1000			
	0.3 🖉	-				>

## Run a workflow : HOP!

🚾 Galaxy / METABO	Analyze Data Workflow Shared Data - Visualization - Admin Help - User -			Using 7.8 MB
Tools		Â	History	C \$
search tools	Successfully ran workflow "Workflow XCMS". The following datasets have been added to the queue:		sacuri	
<u>Get Data</u>	2: sampleMetadata.tsv		0 bytes	Q 🗹 📎 🗩
WORKFLOW 4 METABOLOMICS	3: xset.TICs_raw.pdf			
2-Preprocessing	4: xset.log.txt			
3-Normalisation	5: xset.group.RData			
4-Quality Control	6: yest aroun Rolats odf			
5-Statistical Analysis	7. veck group log byt			
6-Annotation	7: xset.group.log.txt			
COMMON TOOLS	8: xset.group.retcor.RData			
Text Manipulation	9: xset.group.retcor.TICs_corrected.pdf			
Filter and Sort	10: xset.group.retcor.log.txt			
Join, Subtract and Group	11: xset.group.retcor.group.RData	Ε		
Statistics	12: xset.group.retcor.group.Rplots.pdf			
Graph/Display Data	13; xset.group.retcor.group.log.txt			
Multiple regression	14: xset.group.retcor.group.retcor.RData			
Workflows	15: xset.group.retcor.group.retcor.TICs_corrected.pdf			
All workflows	16: xset.group.retcor.group.retcor.log.txt			
	17: xset.group.retcor.group.retcor.group.RData			
	18: xset.group.retcor.group.retcor.group.Rplots.pdf			
	19: xset.group.retcor.group.log.txt			
	20: xset.group.retcor.group.retcor.group.fillPeaks.RData			
	21: xset.group.retcor.group.retcor.group.fillpeaks.log.txt			
	22: xset.group.retcor.group.retcor.group.fillPeaks.annotateDiffreport.variableMetadata.tsv			
	23: xset.group.retcor.group.retcor.group.fillPeaks.annotateDiffreport.dataMatrix.tsv			
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<	25: xset.group.retcor.group.retcor.group.fillPeaks.annotateDiffreport.log.txt	-	111	3











# **SHARE**



## $biologist \leftrightarrow biologist$

- Sharing histories or datasets
  - With or without linked workflow


# $bioanalyst \leftrightarrow biologist$

- Sharing workflows
  - Pre-configured parameters
  - With or without release parameters (set at runtime)
    - According to the user-end knowledge



# $bioinformatician \leftrightarrow bioinformatician$

- Sharing tools ,scripts and wrappers
  - Toolshed



## Share

### Datasets

💳 Galaxy / METABO	Analyze Data	Workflow	Shared	Data <del>+</del>	Visualization	+ Help+	User+			Using 47.0 MB
Tools	Saved Histories								History	2 <b>¢</b>
search tools	search history names and tags	٩							Preprocessing	
<u>Get Data</u>	Advanced Search								45.6 MB	Q 🗹 📎 🗩
WORKFLOW 4 METABOLOMICS	Name	Datasets	Tags	Sharing	Size on Disk	<u>Created</u>	Last Updated	<u>Status</u>	<u>8:</u>	
2-Preprocessing 3-Normalisation 4-Quality Control		8 1	<u>0 Taqs</u>		45.6 MB	~18 hours	~less than	current history	xset.group.retco ks.diffreport.tsv	tabular
5-Statistical Analysis	Switch					ago	590	instory	<u>xset.group.retco</u>	r.group.fillPea
6-Annotation	PRC Share or Publish		<u>0 Taqs</u>		0 bytes	~2 days ago	~2 minutes ago		ks.diffreport.RDa	ita.rdata
Text Manipulation Filter and Sort	Rename test Delete	1	<u>0 Taqs</u>		4.0 KB	Apr 28, 2014	~4 minutes ago		<u>xset.qroup.retco</u> ks.diffreport.dat	r.group.fillPea a matrix.tsv.t
<u>Join, Subtract and Group</u> <u>Statistics</u> Granh/Display Data	After_Preprocessing   •	з	<u>0 Taqs</u>		1.4 MB	~37 minutes ago	~7 minutes ago		5: bio vs blank bo	● / × x/050.pnq
Multiple regression	Unnamed history   +		<u>0 Taqs</u>		0 bytes	Apr 28, 2014	Apr 28, 2014		<u>4:</u> xset.group.retco	🗶 🖋 🗙 r.group.fillPea
<ul> <li><u>All workflows</u></li> </ul>	For 0 selected histories: Re	ename C	Delete	Delete P	ermanently	Undelete			<u>ks.annotateDiffr</u> ar	<u>eport.tsv.tabul</u>
	Histories that have been deleted permanently deleted.	for more th	an a tim	e period s	pecified by th	ie Galaxy a	dministrator(s)	may be	<u>3:</u> xset.group.retco ks.annotateDiffr ata	• X r.qroup.fillPea eport.Rdata.rd
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## Share

### Workflow



## Share

### Mode

Calaxy / METABO
Analyze Data
Workflow
Shared Data + Visualization + Help + User +
Using 4.0 KB

Share or Publish Workflow 'complete\_workflow\_RFMF'
Make Workflow Accessible via Link and Publish It
This workflow is currently restricted so that only you and the users listed below can access it. You can:
Make Workflow Accessible via Link
Generates a web link that you can share with other people so that they can view and import the workflow.
Make Workflow Accessible and Publish
Makes the workflow accessible via Link (see above) and publishes the workflow to Galaxy's Published Workflows section, where it is publicly listed and searchable.

#### Share Workflow with Individual Users

You have not shared this workflow with any users.

Back to Workflows List



## • Get shared histories

💳 Galaxy / METABO		Analyze Data	Workflow Shared Da	ita <del>-</del> Visualization <del>-</del> F	lelp → User →		Ⅲ	Using 216.	1 MB
Tools	Histories shared	with vou	ı bv others			н	story	C	;
search tools		,,				in	HISTORY	/ LISTS	- 1
<u>Get Data</u>	Name	Datasets	<u>Created</u>	<u>Last Updated</u> ↑	<u>Shared by</u>	6!	Histories	Shared with Me	ר
WORKFLOW 4 METARODULC9	🔲 🥌 minonsoor 🗸	6	Apr 28, 2014	~2 days ago	mmonsoor@sb-roscoff.fr	24	CURRENT	T HISTORY	<b>,</b>
2-Preprocessing	lual					xs	Create N	lew .	
<u>3-Normalisation</u>	For 0 selected histories:	Copy Unsha	re			<u>5.</u>	Copy His	story	
4-Quality Control						23	Copy Da	tasets	
E Statistical Analysis							Share or	Dublich	

💳 Galaxy / METABO	Analyze Data Workflow	Shared Data - Visualization	r≠ Help≠ User≠	Using 70.9 MB
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Advanced Search		Published Histories Published Workflows		
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Preprocessing	mlandi	Published Pages		~14 seconds ago
TP1 xcms sacuri	mmonsoor	****		~1 day ago
TP1 xcms sacuri	jfmartin	****		Apr 28, 2014



## • Get shared workflows

💳 Galaxy / METABO	Analyze Data	Workflow	Shared Data <del>-</del>	Visualization <del>-</del>	Help <del>-</del>	User▼			===	Using 216.1 MB
Your workflows			·					Create new workflow	🛉 Upload o	r import workflow
Name							# of Step	os		
complete_workflow_RFMF -							17	Indivi	dua	
Workflows shared with you by others										
Name		Owner						# of Steps		
Workflow mmonsoor -		mmonsoor	r@sb-roscoff.fr					7		

💳 Galaxy / METABO	Analyze Data	Workflow	Shared Data -	Visualization	+ Help+ U	Jser▼		Using 111.4 MB	
Published Workflows	_		Data Libraries Data Libraries	s Beta		Dı	ublic		
search name, annotation, owner, and tags	Q				FUDIIC				
Advanced Search			Published His	tories					
Name	Association	Owner	Published Wo	orkflows	ating	Community Tags	Lact Undat	adl	
name	Annotation	Owner	Published Vis	ualizations	aung	community rags		<u>eu</u> ț	
complete_workflow_RFMF   -		mland	Published Pag	ges			~17 hours	ago	



## • Import shared

Galaxy / METABO	Analyze Data Workflow Shared Data 🕶	Visualization		Using 216.1 MB
Published Histories   mmonsoor   TP1 xcms sacuri			Import history	About this History
TP1 xcms sacuri 65.4 MB	Hist	ories		Author mmonsoor Related Histories
search datasets	Annotation		8	All published histories Published histories by mmonsoor
1: xset.RData	۲			Rating Community
2: sampleMetadata.tsv	۲			(0 ratings, 0.0 average)
<u>3: xset.TICs_raw.pdf</u>	۲			Yours Tags
4: xset.lon.txt				Community: none

Galaxy / METABO	Analyze Data Workflow	N Shared Data <del>-</del>	Visualization <del>-</del>	Help 👻 Use	er▼			Using 216.1 MB
Your workflows		Wc	orkflo	WS	(	📀 Create new workflow	🛉 Upload o	r import workflow
Name					# of Ste	eps		
complete_workflow_RFMF -					17			

#### Workflows shared with you by others

Name		Owner	# of Steps
Workflow	mmonsoo	mmonsoor@sb-roscoff.fr	7
	View		
Other	Run		
Configuro	Сору		
configure	Remove		

TOT





Level 5

• Share of tools and descriptions in the ToolShed

Level 4





- Level 3
  - Launch autonomously tools

Launch autonomously tools

• Use advanced parameters

. Use the Galaxy API

Use workflow more or less presetted

Provide workflow for collegues Level 1-3

Level 2

Use presetted workflow

Level 1

• Share his data to collegues Level 2-5











# END



# BONUS



# How are tools born? BONUS



• How to import a tool in Galaxy?





## . How to import a tool in Galaxv?

```
[lecorquille@n0 ~]$ e-PCR --help
e-PCR: invalid option -- -
usage: [-hV] [posix-options] stsfile [fasta ...]
[compat-options]
where posix-options are:
         -m ##
                  Margin (default 50)
                  Wordsize (default 7)
         -w ##
         -n ##
                  Max mismatches allowed (default 0
                  Max indels allowed (default 0)
         -a ##
                  Use ## discontiguos words, slow i
         -f ##
                             ##>1
                  Set output file
         -0 ##
                  Set output format:
         -t ##
                  1 - classic, range (pos1..pos2)
                  2 - classic, midpoint
                  3 - tabular
                  4 - tabular with alignment in com
                            (slow)
         -d##-##
                  Set default size range
                            (default 100-350)
                  Turn hits postprocess on/off
         -p +-
                  Verbosity flags
         -v ##
                  Use presize alignmens (only if
         -a a|f
                            gaps>0), slow
                   a - Allways or f - as Fallback
                  Use 5'-end lowercase masking of
         -x +-
                            primers (default -)
                  Uppercase all primers (default -
         -u +-
```

#### Galaxy / ABiMS

#### e-PCR (version 1.0.0)

#### STS file:

100: (as tabular) Trinity on data 9..Transcripts

#### format : tabular

#### Fasta file:

100: Trinity on data 9.. Transcripts

#### format : fasta

#### Wordsize (W):

Set word size for primers hash (nucleotide positions). Longer word size decreases hash collision rate, but increases memory usage. Also no mismatches are allowed within word size near 'inner' boundary of primers unless one uses discontiguous words, and no gaps are ever allowed in that region.

#### Use ## discontinuos words (F):

#### 1

Set discontiguous word count for primers hash (1 means 'use contiguous words'). Discontiguous words increase number of hash tables and decrease 'effective' word size (thus increasing hash collision rate), so make search significantly slower, but increase sencitivity by allowing mismatches within word size. Reasonable values are 1 (contiguous words) and 3.

#### Margin (M):

#### 50

Set maximal allowed deviation of hit product size from expected STS size.

#### Set default sts lower size (D):

Set ddefault STS size range - values used for STSs that have no size associated in file.

#### Set default sts higher size (D):

400

0

0

100

Set ddefault STS size range - values used for STSs that have no size associated in file.

#### Max mismatches allowed (N):

Set maximal number of mismatches allowed in primer-to-sequence alignment (per primer!).

#### Max indels allowed (G):

Set maximal number of gaps allowed in primer-to-sequence alignment (per primer!).

#### Set output format (T):

tabular Output formats

Execute

>



## • How to import a tool in Galaxy?

#### 

<description>e-PCR parses stsfile in unists format, then reads nucleotide sequence data in FASTA format from files listed in commandline if
any, or from stdin otherwise. For input sequences e-PCR finds matches and prints output in one of three formats./description>

<command>e-PCR -w \$wordsize -f \$wordcnt -m \$margin -d\$sts\_size\_lo-\$sts\_size\_hi -n \$max\_mismatch -g \$max\_gap -t \$output\_format \$infile\_stsfile \$infile\_fasta > \$output</command>

#### <inputs>

- >> <param name="infile\_fasta" type="data" label="Fasta file" format="fasta" help="format : fasta" />
- > <param name="wordsize" type="integer" label="Wordsize (W)" value="7" help="Set word size for primers hash (nucleotide positions).
  Longer word size decreases hash collision rate, but increases memory usage. Also no mismatches are allowed within word size near
  'inner' boundary of primers unless one uses discontiguous words, and no gaps are ever allowed in that region." />
  - > cparam name="wordcnt" type="integer" label="Use ## discontinuos words (F)" value="1" help="Set discontiguous word count for primers
    hash (1 means 'use contiguous words'). Discontiguous words increase number of hash tables and decrease 'effective' word size (thus
    increasing hash collision rate), so make search significantly slower, but increase sencitivity by allowing mismatches within word
    size. Reasonable values are 1 (contiguous words) and 3." />
  - > <param name="margin" type="integer" label="Margin (M)" value="50" help="Set maximal allowed deviation of hit product size from
    expected STS size." />
- > <param name="sts\_size\_lo" type="integer" label="Set default sts lower size (D)" value="100" help="Set ddefault STS size range values
  used for STSs that have no size associated in file." />
  - > </
    - values used for STSs that have no size associated in file." />
  - > <param name="max\_mismatch" type="integer" label="Max mismatches allowed (N)" value="0" help="Set maximal number of mismatches allowed
    in primer-to-sequence alignment (per primer!)." />
  - > <param name="max\_gap" type="integer" label="Max indels allowed (G)" value="0" help="Set maximal number of gaps allowed in primer-tosequence alignment (per primer!)." />
    - <param name="output\_format" type="select" help="Output formats">
      - <label>Set output format (T)</label>
      - >> <option value="1">classic, range (pos1..pos2)</option>
      - » <option value="2">classic, midpoint</option>
      - » <option value="3" selected="true">tabular</option>
    - » <option value="4">tabular with alignment in comments (slow)</option>

#### </param>

#### </inputs>

#### <outputs>

- > <data name="output" format="tabular" />
- </outputs>



## • How to import a tool in Galaxy?

[lecorquille@n0 ~]\$ e-PCRhelp	💳 Galaxy / ABiMS
e-PCR: invalid option	e-PCR (version 1.0.0)
usage: [-hV] [posix-options] stsfile [fasta]	STS file:
[compat-options]	100: (as tabular) Trinity on data 9Transcripts
where posix-options are:	format : tabular
<u> </u>	Fasta file:
-w ## Wordsize (default 7)	100: Trinity on data 9Transcripts
-n ## Max mismatches allowed (default (	Wordsize (W):
-g ## Max indels allowed (default 0)	7
-f ## Use ## discontiguos words, slow :	Set word size for primers hash (nucleotide positions). Longer word size decreases hash collision rate, but increases memory usage. Also no mismatches are allowed within word size near 'inner' boundary of primers unless one uses discontiguous words, and no gaps
##>1	are ever allowed in that region.
-o ## Set output file	Use ## discontinuos words (F):

<tool id="abims\_epcr" name="e-PCR">

<!-- author : lecorguille@sb-roscoff.fr -->

<!-- date : 11-05-12 -->

<description>e-PCR parses stsfile in unists format, then reads nucleotide sequence data in FASTA format from files listed in commandline if
any, or from stdin otherwise. For input sequences e-PCR finds matches and prints output in one of three formats.</description>

<command>e-PCR -w \$wordsize -f \$wordcnt -m \$margin -d\$sts\_size\_lo-\$sts\_size\_hi -n \$max\_mismatch -g \$max\_gap -t \$output\_format \$infile\_stsfile \$infile\_fasta > \$output</command>

#### <inputs>

>	<pre><param format="tabular" help="format : tabular" label="STS file" name="infile_stsfile" type="data"/></pre>
>	param name="infile fasta" type="data" label="Fasta file" format="fasta" help="format : fasta" />
»	<pre><param help="Set word size for primers hash (nucleotide positions).&lt;/pre&gt;&lt;/th&gt;&lt;/tr&gt;&lt;tr&gt;&lt;th&gt;&lt;/th&gt;&lt;th&gt;Longer word size decreases hash collision rate, but increases memory usage. Also no mismatches are allowed within word size near&lt;/th&gt;&lt;/tr&gt;&lt;tr&gt;&lt;th&gt;&lt;/th&gt;&lt;th&gt;'inner' boundary of primers unless one uses discontiguous words, and no gaps are ever allowed in that region." label="Wordsize (W)" name="wordsize" type="integer" value="7"/></pre>
>	<pre><param help="Set discontiguous word count for primers&lt;/pre&gt;&lt;/th&gt;&lt;/tr&gt;&lt;tr&gt;&lt;th&gt;&lt;/th&gt;&lt;th&gt;hash (1 means 'use contiguous words'). Discontiguous words increase number of hash tables and decrease 'effective' word size (thus&lt;/th&gt;&lt;/tr&gt;&lt;tr&gt;&lt;th&gt;&lt;/th&gt;&lt;th&gt;increasing hash collision rate), so make search significantly slower, but increase sencitivity by allowing mismatches within word&lt;/th&gt;&lt;/tr&gt;&lt;tr&gt;&lt;th&gt;&lt;/th&gt;&lt;th&gt;size. Reasonable values are 1 (contiguous words) and 3." label="Use ## discontinuos words (F)" name="wordcnt" type="integer" value="1"/></pre>
>	<param help="Set maximal allowed deviation of hit product size from&lt;/th&gt;&lt;/tr&gt;&lt;tr&gt;&lt;th&gt;&lt;/th&gt;&lt;th&gt;expected STS size." label="Margin (M)" name="margin" type="integer" value="50"/>
>	<pre><param help="Set ddefault STS size range - values&lt;/pre&gt;&lt;/th&gt;&lt;/tr&gt;&lt;tr&gt;&lt;th&gt;&lt;/th&gt;&lt;th&gt;used for STSs that have no size associated in file." label="Set default sts lower size (D)" name="sts_size_lo" type="integer" value="100"/></pre>
>	<pre><param help="Set ddefault STS size range -&lt;/pre&gt;&lt;/th&gt;&lt;/tr&gt;&lt;tr&gt;&lt;th&gt;&lt;/th&gt;&lt;th&gt;values used for STSs that have no size associated in file " label="Set default sts higher size (D)" name="sts_size_hi" type="integer" value="400"/></pre>

charam name="max mismatch" type="integer" label="Max mismatches allowed (N)" value="0" help="Set maximal number of mismatches allowed