



Biogenouest
BIOGÉOINFORMATIQUE



OCEANOMICS



ARBIMS⁴

14/12/2020



Initiation

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Erwan Corre

Credits to Gildas Le Corguillé, Galaxy Training Network

v2.6



- Learning objectives:
 - Familiarize yourself with the basics of Galaxy
 - Learn how to import data
 - Learn how to run tools
 - Learn how histories work
 - Learn how to create a workflow
 - Learn how to share your work
 - Understand and master dataset collections



- Introduction
- Data import
- Tools
- Dataset
- History
- Workflow
- Share

- Introduction
- Data import
- Tools
- Dataset
- History
- Workflow
- Share



Sequence files manipulation



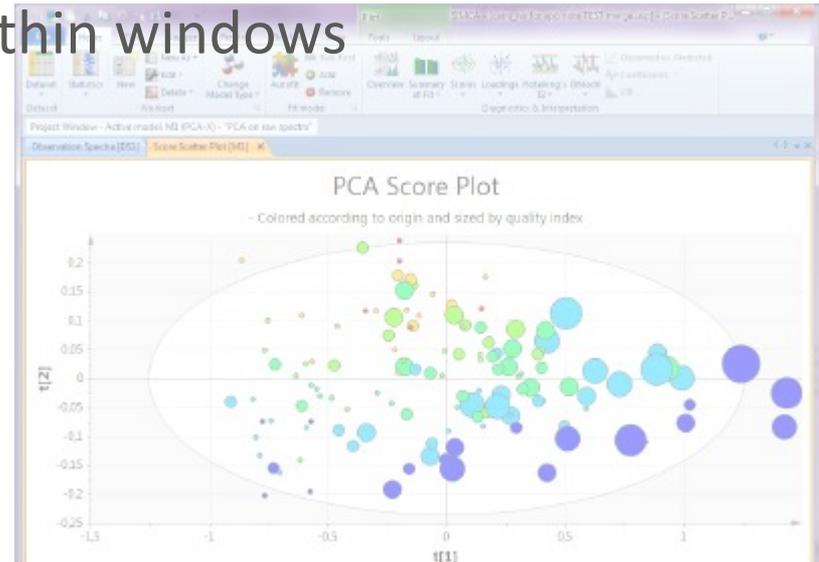
INTRODUCTION / PROBLEMATIC

- In biomedical research, high-throughput technologies produce **large datasets**.
- How to perform analyses of these data **without bioinformatics skills** ?
 - Assemble transcript sequences de novo
 - Determine the gene expression
 - Build a phylogenetic tree
 - Predict subcellular targeting for proteins
 - Identify and quantify metabolites detected by LC-MS
 - ...

Introduction / problematic

- **Graphical interface click-button tools within windows**

- + very ergonomic
- too ergonomic → lack of flexibility
- few
- paying for it!



- **Tools available on the internet**

- + very ergonomic
- too ergonomic → lack of flexibility
- A small part of the available tools
- the submission size /storage is often limited
- must not be paranoid

Introduction / problematic

```
library(xcms)
loaddata()
polar = "Po"
```

● Command line tools

```
noise=250000
xset <- xcmsSet(cdfFiles, ppm=ppm, mzdiff=mzwid, peakwidth=peakwidth, noise=noise, snthresh=snth, method="centWave", fitgauss=TRUE, nSlaves=8)
xset2<-retco
+ represent almost the majority of scientific tools
dev.copy2pdf(device = 2, file = paste(pathResult, "/Ret_Cor-Graph", expe, "_", polar, ".pdf", sep=""), paper="a4", height=9, width=14)
xset3<-group(xset2, minfrac = 0.2, bw = bw, minsamp = 1, mzwid = mzwid, max = 50, sleep = 0)
xset5<-fillPeaks(xset3)
+ good parameters completeness
# rapport final avec statistiques de différences entre les deux classes
reporttab <- diffreport(xset5, filebase =paste(pathResult, "/Rapport_", expe, "_", polar, sep=""), mzdec=4, eicmax=5000, metlin = metlin, classeic=levels(xset5@phenoDa
+ can be executed on high performance computers
#écriture du fichier Excel
dir.create(paste(pathResult, "/Rapport_", expe, "_", polar, "_diffreport/", sep=""), showWarnings = FALSE)
write.table(reporttab, paste(pathResult, "/Rapport_", expe, "_", polar, "_diffreport/resultat_", expe, "_", polar, ".xls", sep=""), sep="\t")
+ automatable, workflowsable, ...
```

```
library(CAMERA)
#annotation version rapide?
an<-annotate(xset5, nSlaves=8, calcIso=TRUE, calcCaS=FALSE, maxcharge=3, maxiso=4, minfrac=0.5,
ppm=15, mzabs=0.015, quick=FALSE, psg_list=NULL, rules=NULL,
polarity=polarity)
diffreport1<-getPeaklist(an)
```

- minimum linux knowledge is required

- cruel lack of ergonomics

```
#diffreport <- annotateDiffreport(xsg, pval_th=0.05, fc=0.1, nSlaves=8, calcIso=TRUE, calcCaS=FALSE, maxcharge=3, maxiso=4, minfrac=0.5,
# ppm=15, mzabs=0.015, quick=FALSE, psg_list=NULL, rules=NULL,
# polarity=polarity, sortpval=FALSE)
diffreport<-cbind(reporttab, diffreport1[,c("isotopes", "adduct", "pcgroup")])
write.table(diffreport, file=paste(pathResult, "/result_", expe, "_", polar, "_CAMERA_diffreport-fast.xls", sep=""), row.names=FALSE, sep="\t")
```

```
library(FactoMineR)
pca3<-PCA(t(matacp), axes=c(1,2))
pca3<-PCA(t(matacp), axes=c(1,3))
pca3<-PCA(t(matacp), axes=c(2,3))
pca4<-PCA(t(matacplog2))
```

```
# -- output png --
# Percentage of variance
png("percentage_of_variance.png", width =800, height = 400);
barplot(resPCA$eig$per, xlab="Components", ylab="percentage of variance");
dev.off()
```

```
png("eigenvalue.png", width =800, height = 400);
barplot(resPCA$eig$eig, xlab="Components", ylab="eigenvalue");
dev.off()
```

```
library(ctc)
# -- Normalization: logratio --
if (normalization) {
  data=t(scale(t(data)))
```

```
login@sbr4-1042:~$ ssh -Y login@bioinfo.sb-roscoff.fr
```

```
[...]
```

```
[login@n0 ~]$ cd projet
```

```
[login@n0 login]$ cd 13-07-29-panda/tmp/mapping
```

```
[login@n0 mapping]$ cat tophat.qsub
```

```
#!/bin/bash
```

```
#$ -S /bin/bash
```

```
#$ -M login@sb-roscoff.fr
```

```
#$ -m bea
```

```
#$ -V
```

```
#$ -cwd
```

```
#$ -o qsub.out
```

```
#$ -e qsub.err
```

```
tophat2 panda_v121029 ../input/I11R1-1.fq ../input/I11R1-2.fq  
-GTF ../input/panda_v121029.gtf --b2-sensitive -r 100  
-num-threads 8
```

```
[login@n0 mapping]$ qsub -q long.q -pe thread 8 tophat.qsub
```

```
Your job 5338969 ("tophat.qsub") has been submitted
```

```
[login@n0 mapping]$ ls
```

```
accepted_hits.bam junctions.bed qsub.err unmapped.bam
```

```
deletions.bed logs qsub.out
```

```
insertions.bed prep_reads.info tmp
```

```
[login@n0 mapping]$ cd
```

```
login@sbr4-1042:~$ ssh -Y login@bioinfo.sb-roscoff.fr
[...]
[login@n0 ~]$ cdprojet
[login@n0 login]:
[login@n0 mapping]
#!/bin/bash
#$ -S /bin/bash
#$ -M login@sb-ro
#$ -m bea
#$ -V
#$ -cwd
#$ -o qsub.out
#$ -e qsub.err
```

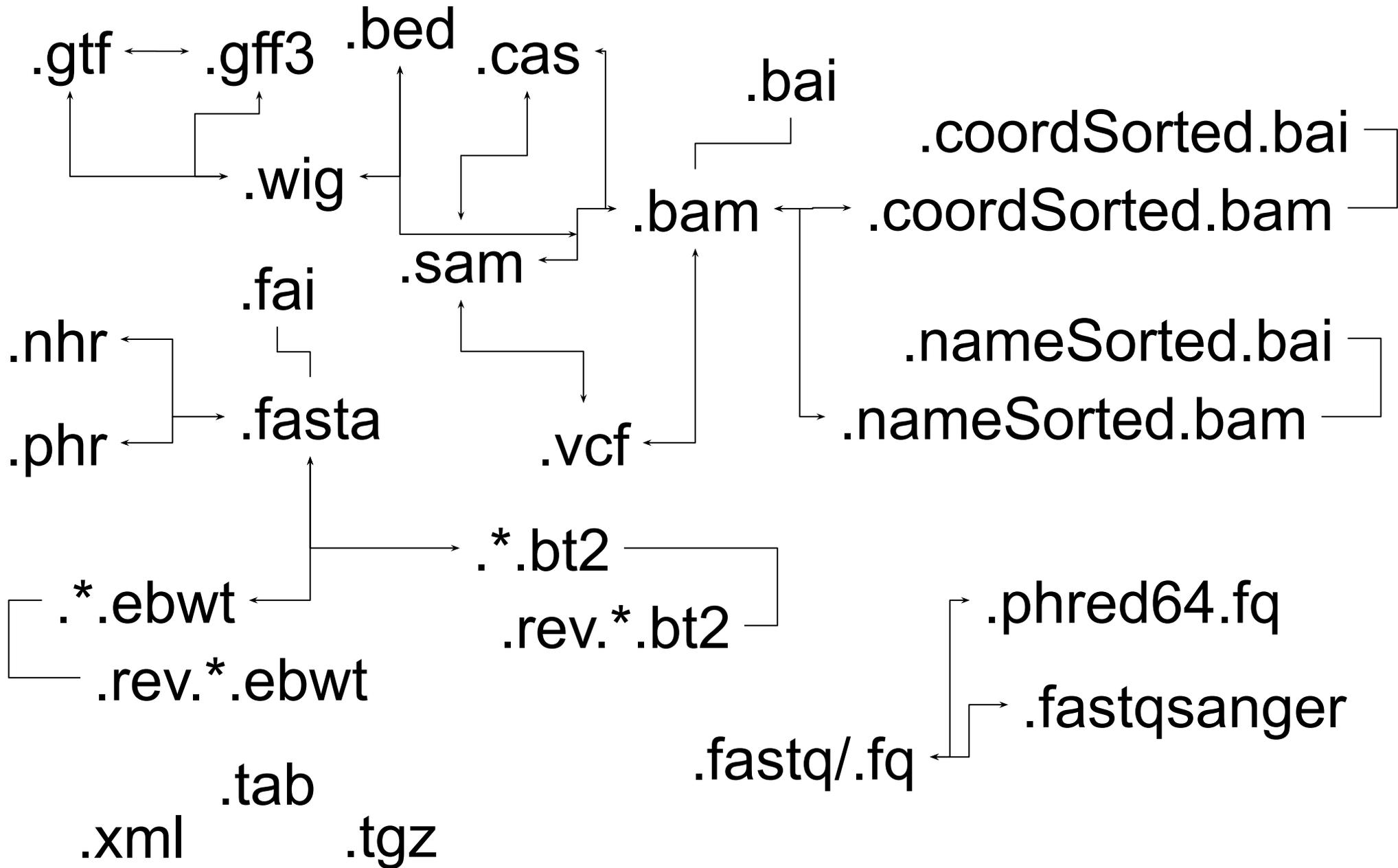


```
tophat2 panda_v12 .R1-2.fq
-GTF ../input/pa
-num-threads 8
```

```
[login@n0 mapping] :.qsub
Your job 5338969
[login@n0 mapping]
accepted_hits.bam junctions.bed qsub.err unmapped.bam
deletions.bed logs qsub.out
insertions.bed prep_reads.info tmp
[login@n0 mapping]$ cd
```

NOOOOOOOO!

Introduction / problematic





Select your level:

Level 1



« I want to know the gene expression »

Level 2



« I want to map my reads on a reference genome and count them »

Level 3



« I want to launch the tools tophat2 and cufflinks.
I have fastq files and my genome in fasta and gtf. »

Level 4



« I want 1TB for my project. I will launch tophat2 through SSH on the cluster in multi-thread mode.

Next I want to submit the bam file to my genome with cufflinks.

Except that, I will manage :P”

Level 5



« I have a bunch of cool tools!
But I'm the only one who can launch them.

Comments? »



- Web-based platform for computational biomedical research (analysis and data integration)
 - Developed at Penn State, Johns Hopkins, OHSU and Cleveland Clinic with substantial outside contributions
 - Open source under Academic Free License
- More than 6,500 citations
- More than 125 [public Galaxy resources](#)
 - 100+ public servers, many more non-public
 - Both general-purpose and domain-specific



INTRODUCTION / GALAXY

Why Galaxy ?

- Accessibility
- Reproductibility
- Transparency

- Galaxy it's ...
 - A web-based interface
 - No need to execute a command line through a terminal
 - Programming or scripting skills are not required
 - Submission of jobs is transparent through a high performance computer cluster
 - Secure histories and data manager
 - A sharing system for data and protocols
 - Tool-boxes of several bioinformatics fields
 - NGS
 - Chemistry
 - Metabolomics
 - Image analysis
 - Statistics
 - Etc ...



MR. GEEK



```
[login@n0 ~]$ cdprojet
[login@n0 login]$ cd 13-07-29-panda/tmp/mapping
[login@n0 mapping]$ cat tophat.qsub
#!/bin/bash
#$ -S /bin/bash
#$ -M login@sb-roscoff.fr
#$ -m bea
#$ -V
#$ -cwd
#$ -o qsub.out
#$ -e qsub.err

tophat2 panda_v121029 ../input/IllR1-1.fq ../input/IllR1-2.fq
-GTF ../input/panda_v121029.gtf --b2-sensitive -r 100
-num-threads 8

[login@n0 mapping]$ qsub -q long.q -pe thread 8 tophat.qsub
Your job 5338969 ("tophat.qsub") has been submitted
```



Introduction / Galaxy

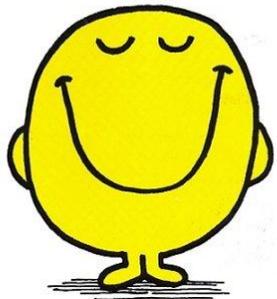
```
[login@n0 ~]$ cdprojet
[login@n0 login]$ cd 13-07-29-panda/tmp/mapping
[login@n0 mapping]$ cat tophat.qsub
#!/bin/bash
#$ -S /bin/bash
#$ -M login@sb-roscoff.fr
#$ -m bea
#$ -V
#$ -cwd
#$ -o qsub.out
#$ -e qsub.err

tophat2_panda_v121029 ../input/I11R1-1.fq ../input/I11R1-2.fq
-GTF ../input/panda_v121029.gtf --b2-sensitive -r 100
-num-threads 8

[login@n0 mapping]$ qsub -q long.q -pe thread 8 tophat.qsub
Your job 5338969 ("tophat.qsub") has been submitted
```



MR. HAPPY
By Roger Hargreaves



```

[lecorguille@n0 ~]$ e-PCR --help
e-PCR: invalid option -- -
usage: [-hV] [posix-options] stsfile [fasta ...]
[compat-options]
where posix-options are:
  -m ##      Margin (default 50)
  -w ##      Wordsize (default 7)
  -n ##      Max mismatches allowed (default 0)
  -g ##      Max indels allowed (default 0)
  -f ##      Use ## discontinuous words, slow if
              ##>1
  -o ##      Set output file
  -t ##      Set output format:
              1 - classic, range (pos1..pos2)
              2 - classic, midpoint
              3 - tabular
              4 - tabular with alignment in comments
                 (slow)
  -d##-##    Set default size range
              (default 100-350)
  -p +-      Turn hits postprocess on/off
  -v ##      Verbosity flags
  -a a|f     Use presize alignmens (only if
              gaps>0), slow
              a - Always or f - as Fallback
  -x +-      Use 5'-end lowercase masking of
              primers (default -)
  -u +-      Uppercase all primers (default -)

[...]
```

Galaxy / ABiMS

e-PCR (version 1.0.0)

STS file:

 format : tabular

Fasta file:

 format : fasta

Wordsize (W):

Set word size for primers hash (nucleotide positions). Longer word size decreases hash collision rate, but increases memory usage. Also no mismatches are allowed within word size near 'inner' boundary of primers unless one uses discontinuous words, and no gaps are ever allowed in that region.

Use ## discontinuous words (F):

Set discontinuous word count for primers hash (1 means 'use contiguous words'). Discontinuous words increase number of hash tables and decrease 'effective' word size (thus increasing hash collision rate), so make search significantly slower, but increase sensitivity by allowing mismatches within word size. Reasonable values are 1 (contiguous words) and 3.

Margin (M):

Set maximal allowed deviation of hit product size from expected STS size.

Set default sts lower size (D):

Set ddefault STS size range - values used for STSs that have no size associated in file.

Set default sts higher size (D):

Set ddefault STS size range - values used for STSs that have no size associated in file.

Max mismatches allowed (N):

Set maximal number of mismatches allowed in primer-to-sequence alignment (per primer!).

Max indels allowed (G):

Set maximal number of gaps allowed in primer-to-sequence alignment (per primer!).

Set output format (T):

 Output formats

Tools ☆ ↑

search tools ✕

Get Data

Send Data

Collection Operations

COMMON TOOLS

Text Manipulation

Filter and Sort

Join, Subtract and Group

Convert Formats

Extract Features

Fetch Sequences

Statistics

Graph/Display Data

Fasta Fastq Manipulation

Operate on Genomic Intervals

Visualization

NCBI SRA Tools

NGS: deepTools

COMMON NGS TOOLS

NGS: Samtools

✓ Welcome to galaxy.sb-roscoff.fr

⚠ 14-10-2020

Dear users, we plan to eventually close this <https://galaxy.sb-roscoff.fr> instance in favour of <https://usegalaxy.fr>. So, if you start a new analysis, please consider migrating to the new national instance <https://usegalaxy.fr>. This instance is more modern, supported by the IFB Core Cluster, maintained in part by ABiMS and the IFB NNCR Cluster TaskForce. If tools are missing, do not hesitate to request them on <https://community.france-bioinformatique.fr/>.

i Information

For any question or request for tools or account, send an email at support.abims@sb-roscoff.fr

By logging in and using this Galaxy instance, you accept both **CNRS Charter** and **IFB Term of Usage**



Analyses and Bioinformatics for Marine Science



Account

- ▼ Changelog
- 28-04-20: UPDATE - Trinity suite (2.9.1)
 - 26-09-19: NEW - Genome assembler tools: SPAdes (3.12.0) and ABySS (2.2.1)

History ↻ + 📄 ⚙

search datasets ✕

Public - Galaxy initiation training 2020 - blast hands-on

13 shown, 12 hidden

154.24 MB

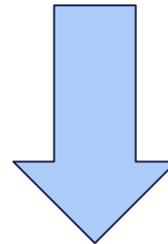
- 18: NCBI BLAST+ blastn across collection 17 ✕
a list with 5 items
- 17: queries ✕
a list with 5 items
- 11: blastx query4.fa vs protein BLAST database from data 2 👁 ✎ ✕
- 10: megablast query5.fa vs nucleotide BLAST database from data 1 👁 ✎ ✕
- 9: protein BLAST database from data 2 👁 ✎ ✕
- 8: nucleotide BLAST database from data 1 👁 ✎ ✕
- 7: query5.fa 👁 ✎ ✕

Galaxy ABiMS --> Galaxy France

<https://galaxy.sb-roscoff.fr/>



The screenshot shows the Galaxy ABiMS interface. The top navigation bar includes 'Galaxy / ABiMS', 'Analyze Data', 'Workflow', 'Visualize', 'Shared Data', 'Admin', 'Help', and 'User'. A 'Using 34%' indicator is in the top right. The left sidebar lists 'Tools' with a search box and categories like 'Get Data', 'Send Data', and 'Collection Operations'. The main content area features a green 'Welcome to galaxy.sb-roscoff.fr' message and an orange warning box dated 14-10-2020 stating: 'Dear users, we plan to eventually close this https://galaxy.sb-roscoff.fr instance in favour of https://usegalaxy.fr. So, if you start a new analysis, please consider migrating to the new national instance https://usegalaxy.fr. This instance is more modern, supported by the IFB Core Cluster, maintained in part by ABiMS and the IFB NNCR Cluster TaskForce. If tools are missing, do not hesitate to request them on https://community.france-bioinformatique.fr/'. The right sidebar shows 'History' with a search box and a list of datasets, including 'Galaxy initiation training 2019 - tools hands-on'.



<https://usegalaxy.fr/>



The screenshot shows the Galaxy France interface. The top navigation bar includes 'Galaxy France', 'Analyze Data', 'Workflow', 'Visualize', 'Shared Data', 'Help', and 'Login or Register'. A 'Using 0%' indicator is in the top right. The left sidebar lists 'Tools' with a search box and categories like 'Get Data', 'Send Data', and 'Collection Operations'. The main content area features a 'Welcome to usegalaxy.fr' message with a graphic of two vertical bars (one blue, one orange) and a text box stating: 'By using this Galaxy instance, we assume that you have read and accept the Term Of Use. For any questions or support: community.cluster.france-bioinformatique.fr/c/galaxy'. The right sidebar shows 'History' with a search box and an 'Unnamed history' section that is empty, with a message: 'This history is empty. You can load your own data or get data from an external source'.

Galaxy interface

Galaxy / 4 / Metabolomics

Analyze Data Workflow Shared Data Visualization Admin Help User

Using -993344424 b

Tools

search tools

Upload File from your computer

Export Data

LC-MS

Format Conversion

Preprocessing

Normalisation

Batch_correction Corrects intensities for signal drift and batch-effects

Determine_batch_correction to choose between linear, lowess and loess methods

Transformation Transforms the dataMatrix intensity values

Quality Control

Statistical Analysis

Annotation

GC-MS

Preprocessing

Normalisation

Quality Control

Statistical Analysis

Annotation

NMR

Preprocessing

Normalisation

Batch_correction (version 2.0.0)

Data Matrix file : 17: xset.group.retcor.group.fillPeaks.annotate.dataMatrix.tsv

Sample metadata file : 3: sampleMetadata.tsv
must contain at least the three following columns: 'batch' + 'injectionOrder' + 'sampleType'

Variable metadata file : 16: xset.group.retcor.group.fillPeaks.annotate.variableMetadata.tsv

Type of regression model : linear
To select between linear or non-linear (lowess or loess) methods to be used in Van der Kloet algorithm ; when using loess, you can choose to use pools or samples to model batch effect.

Factor of interest : batch
column name of factor of interest (often a biological factor); if none, leave 'batch'

Level of details for plots : basic
Amount of plots in the pdf file output. See Help section for more details.

Execute

History

search datasets

Sacuri Zip
19 shown
289.7 MB

19: xset.group.retcor.group.fillPeaks.annotate.variableMetadata.tsv (Xdiffreport)

18: xset.group.retcor.group.fillPeaks.annotate.negative.Rdata

17: xset.group.retcor.group.fillPeaks.annotate.dataMatrix.tsv

16: xset.group.retcor.group.fillPeaks.annotate.variableMetadata.tsv

15: xset.group.retcor.group.fillPeaks.RData

14: xset.group.retcor.group.Rplots.pdf

13: xset.group.retcor.group.RData

12: xset.group.retcor.BPCs_corrected.pdf

11:

1 Authors
Jean-Francois Martin - PF MetaToul-AXIOM ; INRA ; MetaboHUB (for original version of this tool and overall development of the R script)

1 Contributors
Melanie Petera - PFEM ; INRA ; MetaboHUB (for R wrapper and R script improvement)
Etienne Thevenot - LIST/LADIS ; CEA ; MetaboHUB (for R script and wrapper concerning "all loess pool" and "all loess sample" methods)

Galaxy interface

Top menu

The screenshot displays the Galaxy 4 Metabolomics interface. The top navigation bar includes 'Galaxy / 4 / Metabolomics', 'Analyze Data', 'Workflow', 'Shared Data', 'Visualization', 'Admin', 'Help', and 'User'. The main content area shows the configuration for the 'Batch_correction (version 2.0.0)' tool. The configuration includes fields for 'Data Matrix file', 'Sample metadata file', 'Variable metadata file', 'Type of regression model', 'Factor of interest', and 'Level of details for plots'. An 'Execute' button is located at the bottom of the configuration panel. Below the configuration, there is a section for 'Authors' and 'Contributors'. The right-hand side of the interface features a 'History' panel with a search bar and a list of datasets, including 'Sacuri Zip' and various annotated data files.

Galaxy / 4 / Metabolomics Analyze Data Workflow Shared Data Visualization Admin Help User Using -993344424 b

Tools search tools

Upload File from your computer
Export Data

LC-MS
Format Conversion
Preprocessing
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Transformation Transforms the dataMatrix intensity values

Quality Control
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Preprocessing
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Quality Control
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Batch_correction (version 2.0.0)

Data Matrix file : 17: xset.group.retcor.group.fillPeaks.annotate.dataMatrix.tsv

Sample metadata file : 3: sampleMetadata.tsv
must contain at least the three following columns: 'batch' + 'injectionOrder' + 'sampleType'

Variable metadata file : 16: xset.group.retcor.group.fillPeaks.annotate.variableMetadata.tsv

Type of regression model : linear
To select between linear or non-linear (lowess or loess) methods to be used in Van der Kloet algorithm ; when using loess, you can choose to use pools or samples to model batch effect.

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Execute

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History search datasets

Sacuri Zip
19 shown
289.7 MB

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Galaxy interface

Tool list

Galaxy / 4 / Metabolomics Analyze Data Workflow Shared Data Visualization Admin Help User Using -993344424 b

Tools

search tools

Upload File from your computer

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LC-MS

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Normalisation

[Batch_correction](#) Corrects intensities for signal drift and batch-effects

[Determine_batch_correction](#) to choose between linear, lowess and loess methods

[Transformation](#) Transforms the dataMatrix intensity values

Quality Control

Statistical Analysis

Annotation

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Batch_correction (version 2.0.0)

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Sample metadata file :

3: sampleMetadata.tsv

must contain at least the three following columns: 'batch' + 'injectionOrder' + 'sampleType'

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Type of regression model :

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batch

column name of factor of interest (often a biological factor); if none, leave 'batch'

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[xset.group.retcor.group.fillPeaks.annotate.variableMetadata.tsv \(Xdiffreport\)](#)

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[xset.group.retcor.BPCs_corrected.pdf](#)

11:

Galaxy interface

Web forms / dataset visualization / miscellaneous information

Galaxy / 4 / Metabolomics Analyze Data Workflow Shared Data Visualization Admin Help User Using -993344424 b

Tools search tools Upload File from your computer Export Data LC-MS Format Conversion Preprocessing Normalisation

Batch_correction (version 2.0.0)

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19: xset.group.retcor.group.fillPeaks.annotate.variableMetadata.tsv (Xdiffreport)

18: xset.group.retcor.group.fillPeaks.annotate.negative.Rdata

17: xset.group.retcor.group.fillPeaks.annotate.dataMatrix.tsv

16: xset.group.retcor.group.fillPeaks.annotate.variableMetadata.tsv

15: xset.group.retcor.group.fillPeaks.RData

14: xset.group.retcor.group.Rplots.pdf

13: xset.group.retcor.group.RData

12: xset.group.retcor.BPCs_corrected.pdf

11:

Galaxy interface

History

Galaxy / 4 / Metabolomics

Analyze Data Workflow Shared Data Visualization Admin Help User

Using -993344424 b

Tools

search tools

Upload File from your computer

Export Data

LC-MS

Format Conversion

Preprocessing

Normalisation

Batch_correction Corrects intensities for signal drift and batch-effects

Determine_batch_correction to choose between linear, lowess and loess methods

Transformation Transforms the dataMatrix intensity values

Quality Control

Statistical Analysis

Annotation

GC-MS

Preprocessing

Normalisation

Quality Control

Statistical Analysis

Annotation

NMR

Preprocessing

Normalisation

Batch_correction (version 2.0.0)

Data Matrix file : 17: xset.group.retcor.group.fillPeaks.annotate.dataMatrix.tsv

Sample metadata file : 3: sampleMetadata.tsv
must contain at least the three following columns: 'batch' + 'injectionOrder' + 'sampleType'

Variable metadata file : 16: xset.group.retcor.group.fillPeaks.annotate.variableMetadata.tsv

Type of regression model : linear
To select between linear or non-linear (lowess or loess) methods to be used in Van der Kloet algorithm ; when using loess, you can choose to use pools or samples to model batch effect.

Factor of interest : batch
column name of factor of interest (often a biological factor); if none, leave 'batch'

Level of details for plots : basic
Amount of plots in the pdf file output. See Help section for more details.

Execute

1 Authors
Jean-Francois Martin - PF MetaToul-AXIOM ; INRA ; MetaboHUB (for original version of this tool and overall development of the R script)

1 Contributors
Melanie Petera - PFEM ; INRA ; MetaboHUB (for R wrapper and R script improvement)
Etienne Thevenot - LIST/LADIS ; CEA ; MetaboHUB (for R script and wrapper concerning "all loess pool" and "all loess sample" methods)

History

search datasets

Sacuri Zip
19 shown
289.7 MB

19: xset.group.retcor.group.fillPeaks.annotate.variableMetadata.tsv (Xdiffreport)

18: xset.group.retcor.group.fillPeaks.annotate.negative.Rdata

17: xset.group.retcor.group.fillPeaks.annotate.dataMatrix.tsv

16: xset.group.retcor.group.fillPeaks.annotate.variableMetadata.tsv

15: xset.group.retcor.group.fillPeaks.RData

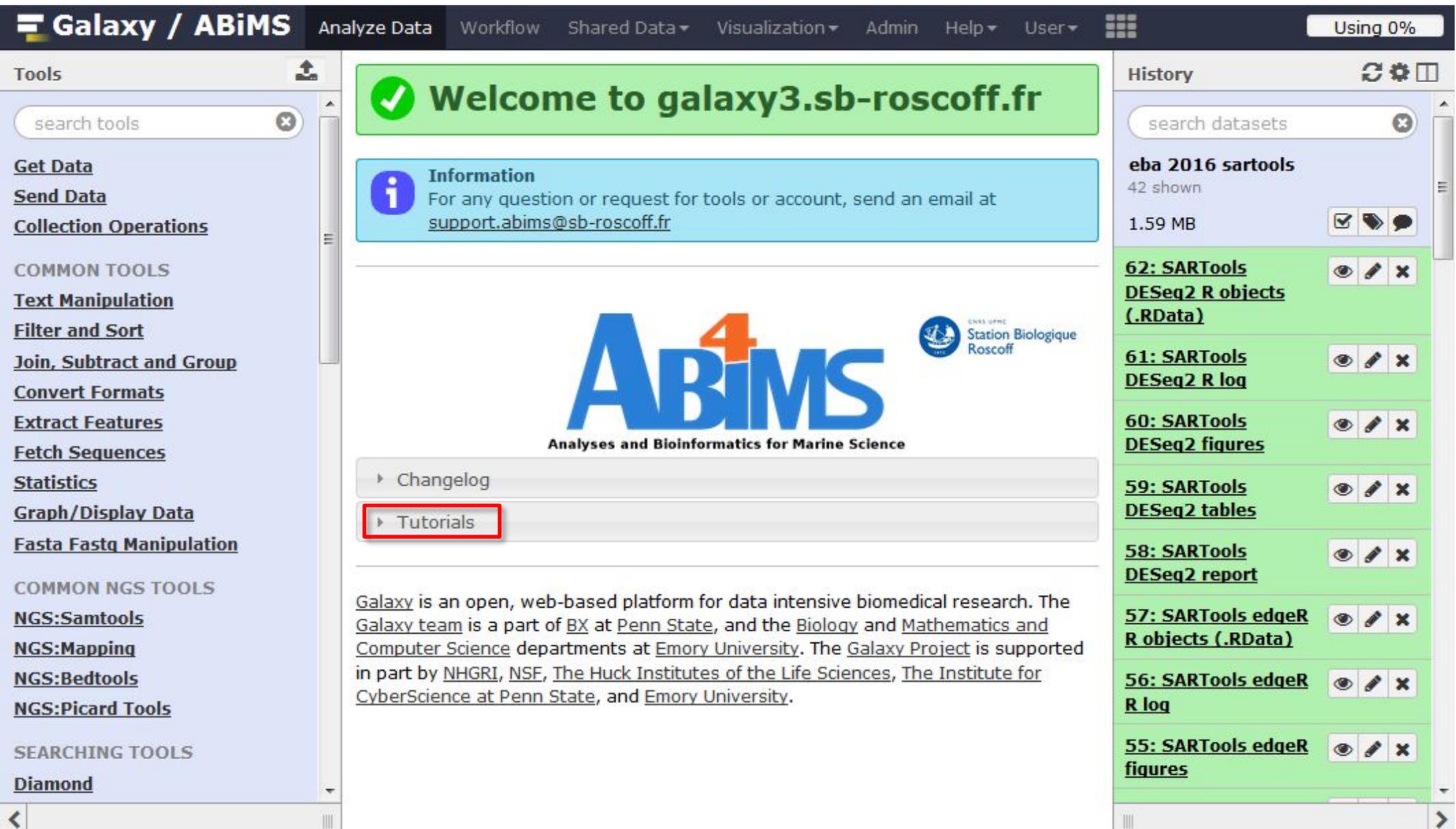
14: xset.group.retcor.group.Rplots.pdf

13: xset.group.retcor.group.RData

12: xset.group.retcor.BPCs_corrected.pdf

11:

GET HELP



Galaxy / ABiMS Analyze Data Workflow Shared Data Visualization Admin Help User Using 0%

Tools

search tools

Get Data
Send Data
Collection Operations

COMMON TOOLS

Text Manipulation
Filter and Sort
Join, Subtract and Group
Convert Formats
Extract Features
Fetch Sequences
Statistics
Graph/Display Data
Fasta Fastq Manipulation

COMMON NGS TOOLS

NGS:Samtools
NGS:Mapping
NGS:Bedtools
NGS:Picard Tools

SEARCHING TOOLS

Diamond

Welcome to galaxy3.sb-roscoff.fr

Information
 For any question or request for tools or account, send an email at support.abims@sb-roscoff.fr

ABiMS
 Analyses and Bioinformatics for Marine Science

Station Biologique Roscoff

Changelog
Tutorials

Galaxy is an open, web-based platform for data intensive biomedical research. The Galaxy team is a part of BX at Penn State, and the Biology and Mathematics and Computer Science departments at Emory University. The Galaxy Project is supported in part by NHGRI, NSF, The Huck Institutes of the Life Sciences, The Institute for CyberScience at Penn State, and Emory University.

History

search datasets

eba 2016 sartools
 42 shown
 1.59 MB

62: SARTools DESeq2 R objects (.RData)

61: SARTools DESeq2 R log

60: SARTools DESeq2 figures

59: SARTools DESeq2 tables

58: SARTools DESeq2 report

57: SARTools edgeR R objects (.RData)

56: SARTools edgeR R log

55: SARTools edgeR figures

Galaxy / ABiMS
Analyze Data
Workflow
Shared Data ▾
Visualization ▾
Admin
Help ▾
User ▾
Using 0%

Tools

Get Data

Send Data

Collection Operations

COMMON TOOLS

Text Manipulation

Filter and Sort

Join, Subtract and Group

Convert Formats

Extract Features

Fetch Sequences

Statistics

Graph/Display Data

Fasta Fastq Manipulation

COMMON NGS TOOLS

NGS:Samtools

NGS:Mapping

NGS:Bedtools

NGS:Picard Tools

SEARCHING TOOLS

Diamond

✓ **Welcome to galaxy3.sb-roscoff.fr**

i **Information**
 For any question or request for tools or account, send an email at support.abims@sb-roscoff.fr



Analyses and Bioinformatics for Marine Science

▸ Changelog

▸ Tutorials

Galaxy is an open, web-based platform for data intensive biomedical research. The Galaxy team is a part of BX at Penn State, and the Biology and Mathematics and Computer Science departments at Emory University. The Galaxy Project is supported in part by NHGRI, NSF, The Huck Institutes of the Life Sciences, The Institute for CyberScience at Penn State, and Emory University.

History

eba 2016 sartools
42 shown
1.59 MB

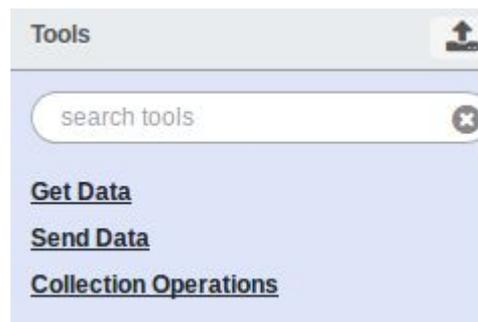
- 62: SARTools DESeq2 R objects (.RData)**
- 61: SARTools DESeq2 R log**
- 60: SARTools DESeq2 figures**
- 59: SARTools DESeq2 tables**
- 58: SARTools DESeq2 report**
- 57: SARTools edgeR R objects (.RData)**
- 56: SARTools edgeR R log**
- 55: SARTools edgeR figures**

DATA IMPORT

- Copy/paste from a file
- Upload data from your local computer
- Upload data from internet using URL
- Upload data from FTP
- Upload data from online databases: EBI ENA, UCSC, BioMart, etc.
- Import from Shared Data (libraries, histories, workflows)

Data import

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- Upload data from your local computer
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- Import from Shared Data (libraries, histories, workflows)



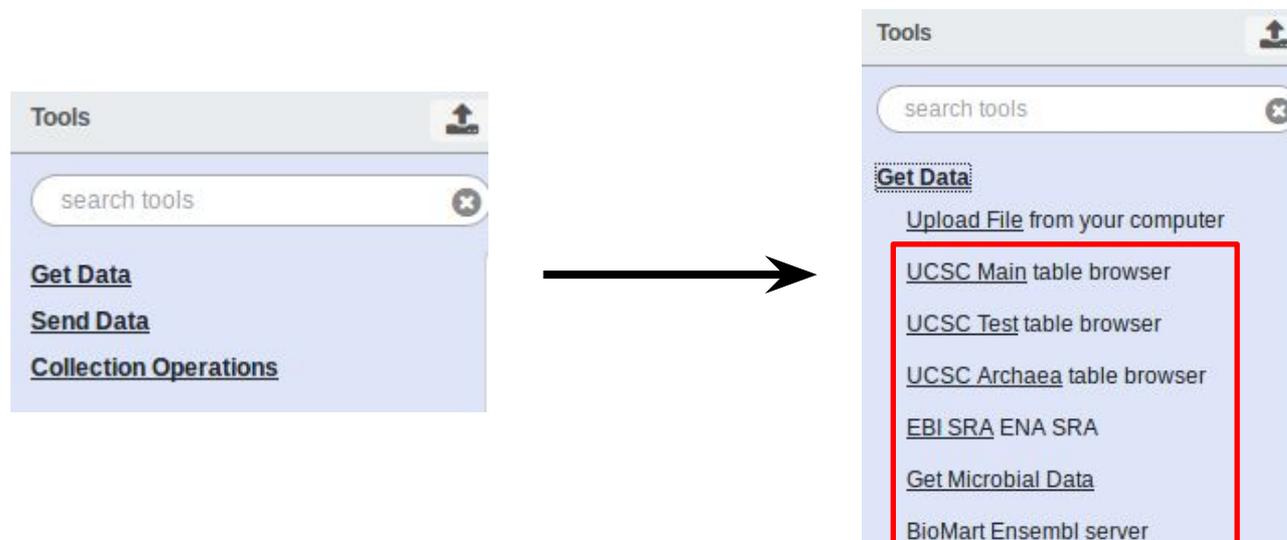
Data import

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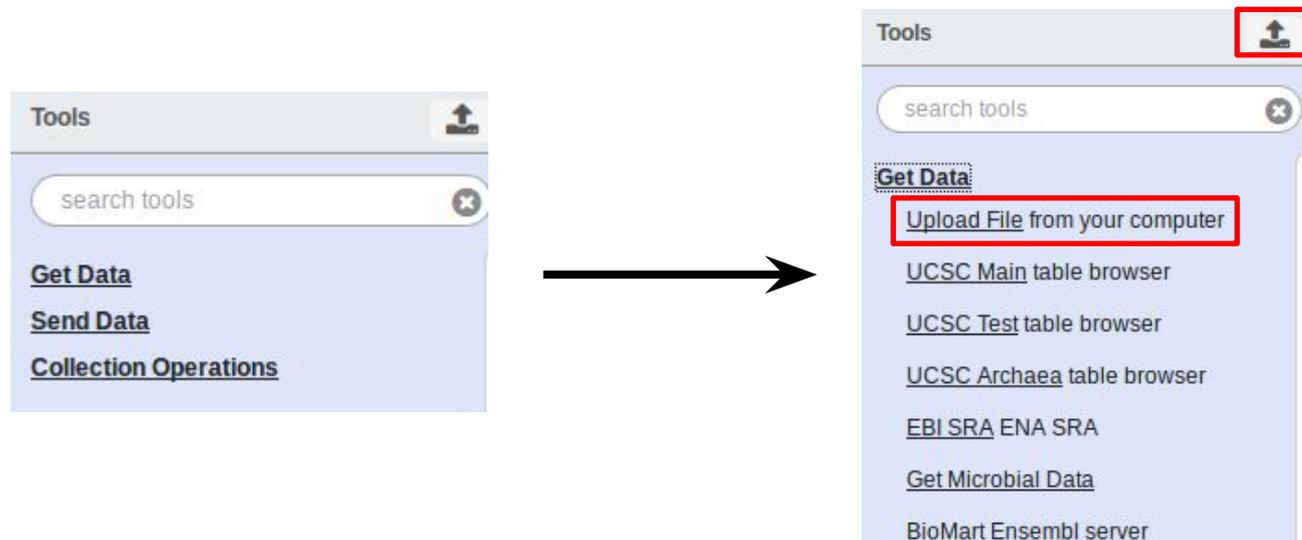
Data import

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Data import

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- Upload data from your local computer
- Upload data from internet using URL
- Upload data from FTP
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- Import from Shared Data (libraries, histories, workflows)



Data import

The screenshot shows the Galaxy 4 / Metabolomics web interface. The browser address bar displays 'galaxy.workflow4metabolomics.org'. The main navigation bar includes 'Galaxy / 4 / Metabolomics', 'Analyze Data', 'Workflow', 'Shared Data', 'Visualization', 'Admin', 'Help', and 'User'. A 'Tools' sidebar on the left lists various analysis categories such as 'Upload File from your computer', 'Export Data', 'LC-MS', 'Preprocessing', 'Normalisation', 'Statistical Analysis', 'Annotation', 'GC-MS', 'NMR', and 'COMMON TOOLS'. A tooltip over the 'Tools' icon reads 'Download from URL or upload files from disk'. The main content area features a green 'Welcome to workflow4metabolomics.org v2.0' banner with a citation: 'Publication: Franck Giacconi, Gildas Le Corguillé, Mishal Monsoor, Marion Landi, Pierre Pericard, Mélanie Pétéra, Christophe Duperier, Marie Tremblay-Franco, Jean-François Martin, Daniel Jacob, Sophie Goulitquer, Etienne A. Thévenot and Christophe Caron (2014). Workflow4Metabolomics: A collaborative research infrastructure for computational metabolomics. Bioinformatics doi:10.1093/bioinformatics/btu813'. Below this is a 'Latest news' section with three entries dated 01/06/2015, 21/09/2015, and 19/12/2014. A 'History' panel on the right shows 'Unnamed history' with 0 bytes and a message: 'This history is empty. You can load your own data or get data from an external source'. The bottom of the interface displays workflow diagrams for 'LC/MS', 'MS', and 'Common'.

Data import

The screenshot shows a web browser window with the URL `galaxy.workflow4metabolomics.org`. The page title is "Galaxy / 4 / Metabolomics". The browser's address bar and navigation icons are visible. The main content area of the browser displays the Galaxy 4 Metabolomics interface, which includes a top navigation bar with "Analyze Data", "Workflow", "Shared Data", "Visualization", "Admin", "Help", and "User" menus. On the left side, there is a "Tools" sidebar with a search bar and a list of tool categories: "Upload File from your computer", "Export Data", "LC-MS", "Preprocessing", "Normalisation", "Statistical Analysis", "Annotation", "GC-MS", "Preprocessing", "Normalisation", "Quality Control", "Statistical Analysis", "Annotation", "NMR", "Preprocessing", "Normalisation", "Quality Control", "Statistical Analysis", "COMMON TOOLS", "Text Manipulation", and "Filter and Sort".

A modal dialog box is centered on the screen with the title "Download data directly from web or upload files from your disk". The dialog contains a large dashed rectangular area for file upload. Below this area, the text reads "You can Drag & Drop files into this box.". At the bottom of the dialog, there are seven buttons: "Choose local file", "Choose FTP file", "Paste/Fetch data", "Start", "Pause", "Reset", and "Close".

The Windows taskbar at the bottom of the screen shows several application icons, including Internet Explorer, File Explorer, Word, Excel, PowerPoint, a document icon, a yellow duck, a small orange figure, and a traffic cone. The system tray on the right shows the language "FR", the Windows logo, and the date and time "10:23 AM 6/5/2015".

Data import

Copy / Paste data

The screenshot shows the Galaxy web interface with the 'Paste/Fetch data' workflow step. The modal window displays the following data:

Name	Size	Type	Genome	Settings	Status
New File	0.9 KB	Auto-detect	unspecified (?)		

The data content is:

```
sampleMetadata class polarity batch  
Blanc15 blank negative 1  
Blanc09 blank negative 1
```

The 'Upload configuration' dialog is open, showing the following options:

- Convert spaces to tabs
- Use POSIX standard

The 'Paste/Fetch data' button is highlighted with a red box and labeled '1'. The data content is highlighted with a red box and labeled '2'. The 'Upload configuration' dialog is highlighted with a red box and labeled '3'.

Data import

From local files

The screenshot shows a web browser window with the URL `galaxy.workflow4metabolomics.org`. The page title is "Galaxy / 4 / Metabolomics". A modal dialog box is open in the center, titled "Download data directly from web or upload files from your disk". The dialog contains a large dashed-line box for file upload, with the text "You can Drag & Drop files into this box." below it. At the bottom of the dialog, there are several buttons: "Choose local file" (highlighted with a red box), "Choose FTP file", "Paste/Fetch data", "Start", "Pause", "Reset", and "Close". The background shows the Galaxy interface with a sidebar of tool categories like "LC-MS", "GC-MS", and "NMR". The Windows taskbar at the bottom shows the time as 10:23 AM on 6/5/2015.

Data import

From local files

The screenshot shows a web browser window with the URL `galaxy.workflow4metabolomics.org`. The browser's address bar and the page's navigation bar are circled in red. The page title is "Galaxy / 4 / Metabolomics". The main content area displays a dialog box titled "Download data directly from web or upload files from your disk". Inside the dialog, there is a large dashed-line box representing a drop zone. A folder icon is being dragged into this box, with a "Move" tooltip visible. Below the drop zone, the text reads "You can Drag & Drop files into this box.". At the bottom of the dialog, there are several buttons: "Choose local file", "Choose FTP file", "Paste/Fetch data", "Start", "Pause", "Reset", and "Close". The background of the browser shows the Galaxy interface with a sidebar of tool categories like "LC-MS", "GC-MS", and "NMR". The Windows taskbar at the bottom shows various application icons and the system clock indicating 10:23 AM on 6/5/2015.

Data import

From local files

Galaxy / 4 / Metabolomics

galaxy.workflow4metabolomics.org

Galaxy / 4 / Metabolomics

Analyze Data Workflow Shared Data Visualization Admin Help User

Tools

search tools

Upload File from your computer

Export Data

LC-MS

Preprocessing

Normalisation

Statistical Analysis

Annotation

GC-MS

Preprocessing

Normalisation

Quality Control

Statistical Analysis

Annotation

NMR

Preprocessing

Normalisation

Quality Control

Statistical Analysis

COMMON TOOLS

Text Manipulation

Filter and Sort

Download data directly from web or upload files from your disk

Name	Size	Type	Genome	Settings	Status
 sacuri.zip	0.2 GB	Auto-det...	unspecified (?)		

You added 1 file(s) to the queue. Add more files or click 'Start' to proceed.

Choose local file Choose FTP file Paste/Fetch data **Start** Pause Reset Close

ARMS

FR 10:23 AM 6/5/2015

Data import

From local files

Galaxy / 4 / Metabolomics

galaxy.workflow4metabolomics.org

Galaxy / 4 / Metabolomics

Analyze Data Workflow Shared Data Visualization Admin Help User

Tools

search tools

Upload File from your computer

Export Data

LC-MS

Preprocessing

Normalisation

Statistical Analysis

Annotation

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Text Manipulation

Filter and Sort

Download data directly from web or upload files from your disk

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sacuri.zip	0.2 GB	Auto-det...	unspecified (?)		

You added 1 file(s) to the queue. Add more files or click 'Start' to proceed.

Choose local file Choose FTP file Paste/Fetch data Start Pause Reset Close

ARMS

FR 10:23 AM 6/5/2015

Data import

From local files

The screenshot shows the Galaxy 4 Metabolomics web interface. A dialog box titled "Download data directly from web or upload files from your disk" is open. The dialog contains a table with the following data:

Name	Size	Type	Genome	Settings	Status
 sacuri.zip	0.2 GB	Auto-det...	unspecified (?)		

Below the table, it says "Please wait...1 out of 1 remaining." At the bottom of the dialog are buttons: "Choose local file", "Choose FTP file", "Paste/Fetch data", "Start" (highlighted by a mouse cursor), "Pause", "Reset", and "Close".

Data import

From local files

Galaxy / 4 / Metabolomics

galaxy.workflow4metabolomics.org

Galaxy / 4 / Metabolomics

Analyze Data Workflow Shared Data Visualization Admin Help User

Tools

search tools

Upload File from your computer

Export Data

LC-MS

Preprocessing

Normalisation

Statistical Analysis

Annotation

GC-MS

Preprocessing

Normalisation

Quality Control

Statistical Analysis

Annotation

NMR

Preprocessing

Normalisation

Quality Control

Statistical Analysis

COMMON TOOLS

Text Manipulation

Filter and Sort

Download data directly from web or upload files from your disk

Name	Size	Type	Genome	Settings	Status
sacuri.zip	0.2 GB	Auto-det...	unspecified (?)		100% ✓

You can Drag & Drop files into this box.

Choose local file Choose FTP file Paste/Fetch data Start Pause Reset Close

ARMS

FR 10:23 AM 6/5/2015

Data import

From local files

The screenshot displays the Galaxy 4 / Metabolomics web interface. The browser address bar shows `galaxy.workflow4metabolomics.org`. The main navigation bar includes 'Galaxy / 4 / Metabolomics', 'Analyze Data', 'Workflow', 'Shared Data', 'Visualization', 'Admin', 'Help', and 'User'. The interface is divided into several sections:

- Tools:** A sidebar on the left with a search bar and categories: 'Upload File from your computer', 'Export Data', 'LC-MS', 'Preprocessing', 'Normalisation', 'Statistical Analysis', 'Annotation', 'GC-MS', 'Preprocessing', 'Normalisation', 'Quality Control', 'Statistical Analysis', 'Annotation', 'NMR', 'Preprocessing', 'Normalisation', 'Quality Control', 'Statistical Analysis', 'COMMON TOOLS', 'Text Manipulation', and 'Filter and Sort'.
- Welcome message:** A green banner with a checkmark icon, titled 'Welcome to workflow4metabolomics.org v2.0'. It includes a publication reference: 'Publication: Franck Giacomoni, Gildas Le Corguillé, Mishari Monsoor, Marion Landi, Pierre Pericard, Mélanie Pétéra, Christophe Duperier, Marie Tremblay-Franco, Jean-François Martin, Daniel Jacob, Sophie Goulitquer, Etienne A. Thévenot and Christophe Caron (2014). Workflow4Metabolomics: A collaborative research infrastructure for computational metabolomics. Bioinformatics doi:10.1093/bioinformatics/btu813' and a support email: 'support@workflow4metabolomics.org'.
- Latest news:** A light blue section with an information icon, titled 'Latest news'. It contains three news items:
 - 01/06/2015 - Workflow4Metabolomics v2.0 starts today - Check the changelog section below
 - 01/06/2015 - The W4M 2.0 release is presented in the June 2015 MetaboNews Spotlight [link]
 - 21/09/2015 - Ecole-chercheurs : Traitement des données métabolomiques sur l'infrastructure online Workflow4Metabolomics (21-25 Sept. 2015) [in French] / Roscoff, France
 - 19/12/2014 - W4M publication in Bioinformatics is now available - Workflow4Metabolomics: A collaborative research infrastructure for computational metabolomics
- Navigation:** Buttons for 'Changelog', 'Tutorials', and 'Past events'.
- History Panel:** On the right, a 'History' panel shows a search bar for datasets. Under 'Unnamed history', it displays '0 bytes' and a single entry: '1: sacuri.zip'. A mouse cursor is hovering over the '1: sacuri.zip' entry.
- Workflow Diagrams:** At the bottom, there are three workflow diagrams labeled 'LC/MS', 'MS', and 'Common', each showing a sequence of processing steps.

The Windows taskbar at the bottom shows the system clock as 10:23 AM on 6/5/2015, and the language is set to FR. A watermark 'Active 4' is visible in the bottom right corner of the browser window.

Data import

From local files

The screenshot displays the Galaxy workflow4metabolomics.org v2.0 web interface. The browser address bar shows the URL galaxy.workflow4metabolomics.org. The main navigation bar includes 'Galaxy / 4 / Metabolomics', 'Analyze Data', 'Workflow', 'Shared Data', 'Visualization', 'Admin', 'Help', and 'User'. A sidebar on the left lists various tools categorized by LC-MS, NMR, and COMMON TOOLS. The central content area features a green 'Welcome to workflow4metabolomics.org v2.0' message with publication details and a 'Latest news' section. A right-hand 'History' panel shows a job named '1: sacuri.zip' with a status of 'This job is currently running'. The bottom of the interface shows a Windows taskbar with various application icons and a system tray with the date 6/5/2015 and time 10:23 AM.

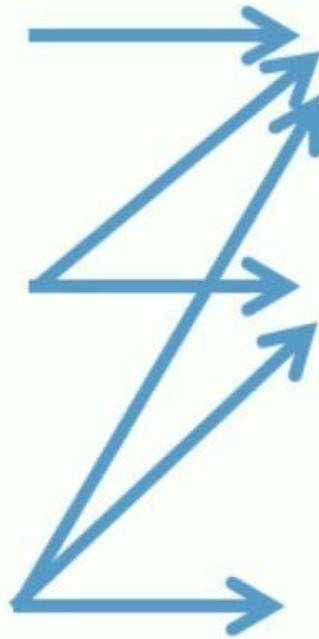
Step 1: Choose a FTP Client

DATA IMPORT USING FTP

STEP 1: CHOOSE A FTP CLIENT



STEP 1: CHOOSE A FTP CLIENT



Avoid:
FileZilla
Malwares inside



Cyberduck



WinSCP

STEP 1: CHOOSE A FTP CLIENT



FileZilla



Cyberduck



WinSCP

Step 2: Easy!

DATA IMPORT USING FTP

Data import using FTP

The screenshot shows a web browser window with the URL `galaxy.sb-roscoff.fr`. The page title is "Galaxy / ABiMS". The main navigation bar includes "Analyze Data", "Workflow", "Shared Data", "Visualization", "Admin", "Help", and "User". A status bar in the top right corner indicates "Using 46%".

The left sidebar contains a "Tools" section with a search bar and a "Download from URL or upload files from disk" button. Below this, there are several categories of tools: "Get Data", "COMMON TOOLS", "Convert Formats", "FASTA manipulation", "Filter and Sort", "Join, Subtract and Group", "Text Manipulation", "Graphics", "Statistics", "EMBOSS 5 Suite", "SEARCHING TOOLS", "NCBI BLAST+", "Diamond", "Primer/Microsatellite", "NGS TOOLS", "NGS: BedTools", "NGS: Mapping", "NGS: Picard", and "NGS: QC and manipulation".

The main content area features a green banner that says "Welcome to galaxy.sb-roscoff.fr". Below this is an "Information" box with an "i" icon, stating: "For any question or request for tools or account, send an email at support.abims@sb-roscoff.fr".

The center of the page displays the "ABiMS" logo, with "4" in orange above "BIMS", and the text "Analyses and Bioinformatics for Marine Science". To the right of the logo is the logo for "CNRS UPRC Station Biologique Roscoff".

Below the logo are two expandable sections: "Changelog" and "Tutorials".

At the bottom of the main content area, there is a paragraph of text: "Galaxy is an open, web-based platform for data intensive biomedical research. The [Galaxy team](#) is a part of [BX](#) at Penn State, and the [Biology](#) and [Mathematics and Computer Science](#) departments at [Emory University](#). The [Galaxy Project](#) is supported in part by [NHGRI](#), [NSF](#), [The Huck Institutes of the Life Sciences](#), [The Institute for CyberScience at Penn State](#), and [Emory University](#)."

The right sidebar contains a "History" section with a search bar and a "Refresh" button. Below this is an "Unnamed history" section showing "0 bytes" and a message: "This history is empty. You can [load your own data](#) or [get data from an external source](#)".

The Windows taskbar at the bottom shows various application icons, including Internet Explorer, Chrome, File Explorer, Word, Excel, PowerPoint, and a yellow duck icon. The system tray in the bottom right corner shows the date and time: "FR 11:19 AM 7/31/2015".

Data import using FTP

The screenshot shows a web browser window with the URL `galaxy.sb-roscoff.fr`. The page title is "Galaxy / ABiMS". The navigation menu includes "Analyze Data", "Workflow", "Shared Data", "Visualization", "Admin", "Help", and "User". A sidebar on the left lists various tools under categories like "COMMON TOOLS", "SEARCHING TOOLS", and "NGS TOOLS".

A modal dialog box is open in the center, titled "Download data directly from web or upload files from your disk". It contains the text "You can Drag & Drop files into this box." and a large dashed rectangular area for file upload. Below this area are two dropdown menus: "Type (set all):" with "Auto-detect" selected, and "Genome (set all):" with "unspecified (?)" selected. At the bottom of the dialog are several buttons: "Choose local file", "Choose FTP file", "Paste/Fetch data", "Start", "Pause", "Reset", and "Close".

The Windows taskbar at the bottom shows the system tray with the date and time "11:19 AM 7/31/2015" and the language "FR".

Data import using FTP

The screenshot shows a web browser window with the URL `galaxy.sb-roscoff.fr`. The page title is "Galaxy / ABiMS". The navigation menu includes "Analyze Data", "Workflow", "Shared Data", "Visualization", "Admin", "Help", and "User". A sidebar on the left lists various tool categories such as "COMMON TOOLS", "FASTA manipulation", "Filter and Sort", "Text Manipulation", "Graphics", "Statistics", "EMBOSS 5 Suite", "SEARCHING TOOLS", "NCBI BLAST+", "Diamond", "Primer/Microsatelli", "NGS TOOLS", "NGS: BedTools", "NGS: Mapping", "NGS: Picard", and "NGS: QC and manip".

A modal dialog box is open in the center of the screen with the title "Download data directly from web or upload files from your disk". Inside the dialog, there is a large dashed box with the text "You can Drag & Drop files into this box." Below this box, there are two dropdown menus: "Type (set all):" with "Auto-detect" selected, and "Genome (set all):" with "unspecified (?)" selected. At the bottom of the dialog, there are several buttons: "Choose local file", "Choose FTP file" (which is being clicked by a mouse cursor), "Paste/Fetch data", "Start", "Pause", "Reset", and "Close".

The Windows taskbar at the bottom shows the system tray with the date and time "11:19 AM 7/31/2015" and the language "FR".

Data import using FTP

The screenshot shows a web browser window with the URL `galaxy.sb-roscoff.fr`. The page title is "Galaxy / ABiMS". The main navigation bar includes "Analyze Data", "Workflow", "Shared Data", "Visualization", "Admin", "Help", and "User". A "Tools" sidebar is visible on the left. A central dialog box titled "Download data directly from web or upload files from your disk" is open. It contains a dashed box for file upload and a smaller "FTP files" dialog box. The "FTP files" dialog box contains the text: "This Galaxy server allows you to upload files via FTP. To upload some files, log in to the FTP server at **galaxy.sb-roscoff.fr** using your Galaxy credentials (email address and password)." Below this text is a yellow warning box that says "Your FTP directory does not contain any files." At the bottom of the main dialog box, there are buttons: "Choose local file", "Choose FTP file", "Paste/Fetch data", "Start", "Pause", "Reset", and "Close". The "Choose FTP file" button is highlighted with a mouse cursor. The Windows taskbar at the bottom shows various application icons and the system clock indicating 11:19 AM on 7/31/2015.

Data import using FTP

The screenshot shows a web browser window with the URL `galaxy.sb-roscoff.fr`. The page title is "Galaxy / ABiMS". The main content area is titled "Download data directly from web or upload files from your disk" and contains a large dashed box for file uploads. A modal dialog box titled "FTP files" is open, displaying the following text: "This Galaxy server allows you to upload files via FTP. To upload some files, log in to the FTP server at **galaxy.sb-roscoff.fr** using your Galaxy credentials (email address and password)." Below this text is a yellow warning box that says "Your FTP directory does not contain any files." At the bottom of the dialog are buttons for "Choose local file", "Choose FTP file", "Paste/Fetch data", "Start", "Pause", "Reset", and "Close". The background interface includes a navigation menu with categories like "Tools", "Get Data", "COMMON TOOLS", "SEARCHING TOOLS", and "NGS TOOLS". The Windows taskbar at the bottom shows various application icons and the system clock indicating 11:19 AM on 7/31/2015.

Data import using FTP

The screenshot shows a Windows desktop environment. In the background, a web browser window displays the Galaxy/ABiMS interface at galaxy.sb-roscoff.fr. The browser's address bar shows the URL, and the page title is "Galaxy / ABiMS". The interface includes a navigation menu with options like "Analyze Data", "Workflow", "Shared Data", "Visualization", "Admin", "Help", and "User". A central panel titled "Download data directly from web or upload files from your disk" contains a large dashed box for file uploads and a message: "You can Drag & Drop files into this box." Below this, a text box explains that users can upload files via FTP by logging into galaxy.sb-roscoff.fr with their Galaxy credentials. A yellow warning box states, "Your FTP directory does not contain any files." At the bottom of the interface, there are buttons for "local file", "Choose FTP file", "Paste/Fetch data", "Start", "Pause", "Reset", and "Close".

In the foreground, the Cyberduck FTP client window is open. The title bar reads "Cyberduck" and "Unregistered". The menu bar includes "File", "Edit", "View", "Go", "Bookmark", "Window", and "Help". The main window area is currently empty, with a "Quick Connect" dropdown menu and "Action", "Get Info", and "Refresh" buttons visible. The bottom of the window shows "0 Bookmarks".

The Windows taskbar at the bottom of the screen displays various application icons, including Internet Explorer, Google Chrome, File Explorer, Microsoft Word, Microsoft Excel, Microsoft PowerPoint, a calculator, a rubber duck, and a traffic cone. The system tray on the right shows the date and time as "11:19 AM 7/31/2015" and the language as "FR".

Data import using FTP

The screenshot shows the Galaxy/ABiMS web interface in a browser window. The browser address bar shows `galaxy.sb-roscoff.fr`. The page title is "Galaxy / ABiMS" and the navigation menu includes "Analyze Data", "Workflow", "Shared Data", "Visualization", "Admin", "Help", and "User".

The main content area displays a message: "Download data directly from web or upload files from your disk". Below this, it says "You can Drag & Drop files into this box." and shows a large dashed box for file upload. A smaller message below the box states: "allows you to upload files via FTP. To upload some files, log at **galaxy.sb-roscoff.fr** using your Galaxy credentials (password). Your FTP directory does not contain any files."

Overlaid on the browser is the Cyberduck application window. The "Open connection" dialog is active, showing the "FTP (File Transfer Protocol)" protocol selected. The "Server:" field is empty, and the "Port:" is set to 21. There are also fields for "URL:", "Username:", and "Password:". The "Anonymous Login" and "Save Password" checkboxes are unchecked. "Connect" and "Cancel" buttons are visible at the bottom of the dialog.

The Windows taskbar at the bottom shows the system tray with the date and time: "FR 11:19 AM 7/31/2015".

Data import using FTP

The screenshot displays the Galaxy/ABiMS web interface. The main content area is titled "Download data directly from web or upload files from your disk" and includes a "You can Drag & Drop files into this box." instruction. A modal dialog titled "Open Connection" is open, showing the configuration for an FTP connection. The dialog includes fields for "Server" (galaxy.sb-roscoff.fr), "Port" (21), "URL" (ftp://lecorguille@galaxy.sb-roscoff.fr:21/), "Username" (lecorguille), and "Password" (masked with dots). There are checkboxes for "Anonymous Login" and "Save Password", and "Connect" and "Cancel" buttons. A message in the background states "Your FTP directory does not contain any files." The desktop environment includes a taskbar with various application icons and a system tray showing the date and time as 11:19 AM on 7/31/2015.

Galaxy / ABiMS

Analyze Data Workflow Shared Data Visualization Admin Help User

Tools

search tools

Get Data

COMMON TOOLS

Convert Formats

Download data directly from web or upload files from your disk

You can Drag & Drop files into this box.

allows you to upload files via FTP. To upload some files, log at **galaxy.sb-roscoff.fr** using your Galaxy credentials password).

Your FTP directory does not contain any files.

local file Choose FTP file Paste/Fetch data Start Pause Reset Close

Cyberduck

Unregistered

File Edit View Go Bookmark Window Help

Open Connection

FTP (File Transfer Protocol)

Server: galaxy.sb-roscoff.fr Port: 21

URL: ftp://lecorguille@galaxy.sb-roscoff.fr:21/

Username: lecorguille

Password:

Anonymous Login

Save Password

Connect Cancel

More Options

0 Bookmarks

FR 11:19 AM 7/31/2015

Data import using FTP

The screenshot displays the Galaxy/ABiMS web interface. The main content area is titled "Download data directly from web or upload files from your disk" and includes a "You can Drag & Drop files into this box." area. Below this, there are instructions: "allows you to upload files via FTP. To upload some files, log in at galaxy.sb-roscoff.fr using your Galaxy credentials (password)." and a message: "Your FTP directory does not contain any files." At the bottom of the interface, there are buttons for "local file", "Choose FTP file", "Paste/Fetch data", "Start", "Pause", "Reset", and "Close".

An "Unsecured FTP connection" warning dialog is overlaid on the interface. The dialog text reads: "Unsecured FTP connection. Password will be sent in plaintext. Please contact your web hosting service provider for assistance." It features two buttons: "Continue" and "Disconnect", and a checkbox for "Don't show again". A "Help" link is also present.

The browser's address bar shows "galaxy.sb-roscoff.fr". The desktop background includes icons for "lecorguille" and "left_kept_r...". The taskbar at the bottom shows various application icons, including Internet Explorer, Chrome, File Explorer, Word, Excel, PowerPoint, and a yellow duck icon. The system tray in the bottom right corner shows the date and time: "FR 11:19 AM 7/31/2015".

Data import using FTP

The screenshot displays the Galaxy/ABiMS web interface. The main content area features a large box titled "Download data directly from web or upload files from your disk" with the instruction "You can Drag & Drop files into this box." Below this, a text box explains that users can upload files via FTP by logging into galaxy.sb-roscoff.fr. A yellow warning box states, "Your FTP directory does not contain any files." At the bottom of the interface, there are buttons for "local file", "Choose FTP file", "Paste/Fetch data", "Start", "Pause", and "Reset".

An FTP client window titled "lecorguille@galaxy.sb-roscoff.fr - FTP" is open in the foreground. It shows a menu bar (File, Edit, View, Go, Bookmark, Window, Help) and a toolbar with "Quick Connect", "Action", "Get Info", and "Refresh" buttons. The main area is a file list table with columns for "Filename", "Size", and "Modified", which is currently empty. A "Copy to /" button is visible at the bottom of the file list.

A system notification bubble in the bottom right corner reads "Connection opened galaxy.sb-roscoff.fr".

Data import using FTP

The screenshot displays a Windows desktop environment. In the background, a web browser window is open to the Galaxy web interface at `galaxy.sb-roscoff.fr`. The browser shows a "Drop files from your disk" area with a message: "Drop files into this box." Below this, a notification states: "Files cannot be uploaded via FTP. To upload some files, log off galaxy.sb-roscoff.fr using your Galaxy credentials." A yellow box below the notification says "This folder does not contain any files." At the bottom of the browser window, there are buttons for "FTP file", "Paste/Fetch data", "Start", "Pause", "Reset", and "Cancel".

In the foreground, an "Transfers" window is open, showing the upload progress of a file named `left_kept_reads.bam`. The progress bar is approximately 55% full. The status text reads: "50.6 MiB (53,018,624 bytes) of 91.6 MiB (55%, 70.1 MB/sec, 1 seconds remaining) Uploading left_kept_reads.bam". The window also includes controls for "Resume", "Reload", "Stop", "Remove", "Open", and "Show". At the bottom of the window, the URL and local file paths are displayed: "URL: ftp://galaxy.sb-roscoff.fr/left_kept_reads.bam" and "Local File: C:\Users\lecorguille\Desktop\left_kept_reads.bam".

In the bottom right corner of the desktop, a notification bubble says "Connection opened galaxy.sb-roscoff.fr". The taskbar at the bottom shows various application icons, including Internet Explorer, Chrome, File Explorer, Word, Excel, PowerPoint, and a notification icon. The system tray in the bottom right corner shows the date and time: "FR 11:19 AM 7/31/2015".

Data import using FTP

The screenshot shows the Galaxy/ABiMS web interface with a file upload dialog open. The dialog title is 'lecorguille@galaxy.sb-roscoff.fr - FTP' and it lists one file: 'left_kept_reads.bam' with a size of 91.6 MiB and a modification date of 7/31/2015 9:19:00 AM. The background web page has a header 'Galaxy / ABiMS' and a main area with the text 'Download data directly from web or upload files from your disk'. A notification bubble in the bottom right corner indicates 'Upload complete left_kept_reads.bam'.

Galaxy / ABiMS

Analyze Data Workflow Shared Data Visualization Admin Help User

Tools

search tools

Get Data

COMMON TOOLS

Convert Formats

Download data directly from web or upload files from your disk

You can Drag & Drop files into this box.

allows you to upload files via FTP. To upload some files, log in at **galaxy.sb-roscoff.fr** using your Galaxy credentials (username and password).

Your FTP directory does not contain any files.

local file Choose FTP file Paste/Fetch data Start Pause Reset

1 Files

Upload complete left_kept_reads.bam

Filename	Size	Modified
left_kept_reads.bam	91.6 MiB	7/31/2015 9:19:00 AM

Data import using FTP

Galaxy / ABiMS

galaxy.sb-roscoff.fr

Analyze Data Workflow Shared Data Visualization Admin Help User

Using 46%

Tools

search tools

Get Data

COMMON TOOLS

Convert Formats

FASTA manipulation

Filter and Sort

Join, Subtract and C

Text Manipulation

Graphics

Statistics

EMBOSS 5 Suite

SEARCHING TOOLS

NCBI BLAST+

Diamond

Primer/Microsatelli

NGS TOOLS

NGS: BedTools

NGS: Mapping

NGS: Picard

NGS: QC and manip

Download data directly from web or upload files from your disk

You can Drag & Drop files into this box.

FTP files

This Galaxy server allows you to upload files via FTP. To upload some files, log in to the FTP server at **galaxy.sb-roscoff.fr** using your Galaxy credentials (email address and password).

Your FTP directory does not contain any files.

Type (set a

specified (?)

Choose local file Choose FTP file Paste/Fetch data Start Pause Reset Close

Upload complete

left_kept_reads.bam

11:19 AM 7/31/2015

Data import using FTP

The screenshot shows a Windows desktop environment with a web browser window open to the Galaxy/ABiMS website. The browser's address bar shows the URL `galaxy.sb-roscoff.fr`. The website's navigation menu includes options like "Analyze Data", "Workflow", "Shared Data", "Visualization", "Admin", "Help", and "User".

A modal dialog box is displayed in the center of the browser window, titled "Download data directly from web or upload files from your disk". The dialog contains a large dashed-line box with the text "You can Drag & Drop files into this box." Below this box, there are two dropdown menus: "Type (set all):" set to "Auto-detect" and "Genome (set all):" set to "unspecified (?)". At the bottom of the dialog, there are several buttons: "Choose local file", "Choose FTP file" (which is highlighted by a mouse cursor), "Paste/Fetch data", "Start", "Pause", "Reset", and "Close".

In the background, the Galaxy/ABiMS interface is partially visible, showing a sidebar with tool categories such as "COMMON TOOLS", "SEARCHING TOOLS", and "NGS TOOLS". A taskbar at the bottom of the screen shows various application icons, including Internet Explorer, Chrome, and several office applications. The system tray in the bottom right corner shows the date and time as "11:19 AM 7/31/2015".

Data import using FTP

The screenshot shows the Galaxy/ABiMS web interface. A modal window titled "Download data directly from web or upload files from your disk" is open. It contains a "Get Data" section with a "Choose FTP file" button selected. An "FTP files" dialog box is also open, displaying a list of available files.

Download data directly from web or upload files from your disk

You can Drag & Drop files into this box.

Get Data

COMMON TOOLS

- Convert Formats
- FASTA manipulation
- Filter and Sort
- Join, Subtract and C
- Text Manipulation
- Graphics
- Statistics
- EMBOSS 5 Suite

SEARCHING TOOLS

- NCBI BLAST+
- Diamond
- Primer/Microsatelli

NGS TOOLS

- NGS: BedTools
- NGS: Mapping
- NGS: Picard
- NGS: QC and manip

FTP files

This Galaxy server allows you to upload files via FTP. To upload some files, log in to the FTP server at **galaxy.sb-roscoff.fr** using your Galaxy credentials (email address and password).

Available files: 1 files 96 MB

<input type="checkbox"/>	Name	Size	Created
<input type="checkbox"/>	left_kept_reads.bam	96 MB	07/31/2015 11:19:45 AM

Buttons: Choose local file, **Choose FTP file**, Paste/Fetch data, Start, Pause, Reset, Close

Data import using FTP

The screenshot shows the Galaxy/ABiMS web interface. A modal window titled "Download data directly from web or upload files from your disk" is open. It displays a table of available files for upload via FTP. The table has columns for Name, Size, Type, Genome, Settings, and Status. One file, "left_kept_reads.bam", is selected. Below the table, there are buttons for "Choose local file", "Choose FTP file", "Paste/Fetch data", "Start", "Pause", "Reset", and "Close".

Download data directly from web or upload files from your disk

You added 1 file(s) to the queue. Add more files or click 'Start' to proceed.

Name	Size	Type	Genome	Settings	Status
left_kept_reads.bam	96 MB	Auto-det...	unspecified (?)		

FTP files

This Galaxy server allows you to upload files via FTP. To upload some files, log in to the FTP server at **galaxy.sb-roscoff.fr** using your Galaxy credentials (email address and password).

Available files: 1 files 96 MB

<input checked="" type="checkbox"/>	Name	Size	Created
<input checked="" type="checkbox"/>	left_kept_reads.bam	96 MB	07/31/2015 11:19:45 AM

Buttons: Choose local file, Choose FTP file, Paste/Fetch data, Start, Pause, Reset, Close

Data import using FTP

Galaxy / ABiMS

galaxy.sb-roscoff.fr

Galaxy / ABiMS

Analyze Data Workflow Shared Data Visualization Admin Help User

Tools

search tools

Get Data

COMMON TOOLS

- Convert Formats
- FASTA manipulation
- Filter and Sort
- Join, Subtract and C
- Text Manipulation
- Graphics
- Statistics
- EMBOSS 5 Suite

SEARCHING TOOLS

- NCBI BLAST+
- Diamond
- Primer/Microsatelli

NGS TOOLS

- NGS: BedTools
- NGS: Mapping
- NGS: Picard
- NGS: QC and manip

Using 46%

Assets

History

History is empty. You can [download data](#) or [get data from an external source](#)

Download data directly from web or upload files from your disk

You added 1 file(s) to the queue. Add more files or click 'Start' to proceed.

Name	Size	Type	Genome	Settings	Status
left_kept_reads.bam	96 MB	Auto-det...	unspecified (?)		

Type (set all): Auto-detect

Genome (set all): unspecified (?)

Choose local file Choose FTP file Paste/Fetch data **Start** Pause Reset Close

FR 11:20 AM 7/31/2015

Data import using FTP

The screenshot shows the Galaxy/ABiMS web interface. A dialog box titled "Download data directly from web or upload files from your disk" is open. The dialog box contains a table with the following data:

Name	Size	Type	Genome	Settings	Status
left_kept_reads.bam	96 MB	Auto-det...	unspecified (?)		100%

Below the table, there are two dropdown menus: "Type (set all):" set to "Auto-detect" and "Genome (set all):" set to "unspecified (?)". At the bottom of the dialog box, there are several buttons: "Choose local file", "Choose FTP file", "Paste/Fetch data", "Start", "Pause", "Reset", and "Close". The "Close" button is being clicked by the mouse.

Data import using FTP

The screenshot displays the Galaxy / ABiMS web interface in a browser window. The address bar shows the URL `galaxy.sb-roscoff.fr`. The main navigation bar includes links for **Analyze Data**, **Workflow**, **Shared Data**, **Visualization**, **Admin**, **Help**, and **User**. A green banner at the top reads "Welcome to galaxy.sb-roscoff.fr". Below this, an information box provides contact details for support at `support.abims@sb-roscoff.fr`. The central content area features the ABiMS logo (Analyses and Bioinformatics for Marine Science) and the Station Biologique Roscoff logo. A sidebar on the left lists various tool categories such as "Get Data", "COMMON TOOLS", "Convert Formats", "FASTA manipulation", "Filter and Sort", "Join, Subtract and Group", "Text Manipulation", "Graphics", "Statistics", "EMBOSS 5 Suite", "SEARCHING TOOLS", "NCBI BLAST+", "Diamond", "Primer/Microsatellite", "NGS TOOLS", "NGS: BedTools", "NGS: Mapping", "NGS: Picard", and "NGS: QC and manipulation". A right-hand sidebar shows the "History" section with a search bar and a list of datasets, including "left_kept_reads.bam". The Windows taskbar at the bottom shows the system clock as 11:20 AM on 7/31/2015.

Data import using FTP

The screenshot displays the Galaxy/ABiMS web interface in a browser window. The address bar shows the URL `galaxy.sb-roscoff.fr`. The main navigation bar includes links for **Analyze Data**, **Workflow**, **Shared Data**, **Visualization**, **Admin**, **Help**, and **User**. A status indicator in the top right corner shows **Using 46%**.

The left sidebar contains a **Tools** section with a search bar and a list of tool categories:

- Get Data**
- COMMON TOOLS**
- Convert Formats**
- FASTA manipulation**
- Filter and Sort**
- Join, Subtract and Group**
- Text Manipulation**
- Graphics**
- Statistics**
- EMBOSS 5 Suite**
- SEARCHING TOOLS**
- NCBI BLAST+**
- Diamond**
- Primer/Microsatellite**
- NGS TOOLS**
- NGS: BedTools**
- NGS: Mapping**
- NGS: Picard**
- NGS: QC and manipulation**

The main content area features a green **Welcome to galaxy.sb-roscoff.fr** message and an information box with contact details: `support.abims@sb-roscoff.fr`. Below this is the **ABiMS** logo and the text **Analyses and Bioinformatics for Marine Science**, accompanied by the **Station Biologique Roscoff** logo. A menu on the left includes **Changelog** and **Tutorials**.

The right sidebar shows a **History** section with a search bar and a list of datasets, including **left_kept_reads.bam**.

The Windows taskbar at the bottom shows various application icons and the system clock indicating **11:20 AM 7/31/2015**.

Data import using FTP

The screenshot shows the Galaxy/ABiMS web interface in a browser window. The address bar shows 'galaxy.sb-roscoff.fr'. The main navigation bar includes 'Analyze Data', 'Workflow', 'Shared Data', 'Visualization', 'Admin', 'Help', and 'User'. A green banner at the top says 'Welcome to galaxy.sb-roscoff.fr'. Below it is an information box with contact details for support.abims@sb-roscoff.fr. The central area features the ABiMS logo and the text 'Analyses and Bioinformatics for Marine Science'. A sidebar on the left lists various tools under categories like 'Get Data', 'COMMON TOOLS', and 'NGS TOOLS'. On the right, the 'History' panel shows a single entry: '1: left_kept_reads.bam', which is 91.6 MB and in BAM format. The desktop background is blue with icons for 'lecorguille' and 'left_kept_r...'. The taskbar at the bottom shows various application icons and the system clock indicating 11:20 AM on 7/31/2015.

~~DATA IMPORT~~

Data import

For HUGE public resources: genome, databank ...

--> Make a request to the support team

Galaxy / ABiMS Analyze Data Workflow Shared Data Visualization Admin Help User

Tools

search tools

Get Data
Send Data
Collection Operations

COMMON TOOLS
Text Manipulation
Filter and Sort
Join, Subtract and Group
Convert Formats
Extract Features
Fetch Sequences
Statistics
Graph/Display Data
Fasta Fastq Manipulation

COMMON NGS TOOLS
NGS:Samtools
NGS:Mapping
NGS:Bedtools
NGS:Picard Tools

SEARCHING TOOLS
Diamond

NCBI BLAST+ blastn Search nucleotide database with nucleotide query sequence(s) (Galaxy Version 0.1.08) Options

Nucleotide query sequence(s)
No fasta dataset available.

Subject database/sequences
Locally installed BLAST database

Nucleotide BLAST database
 Select/Unselect all

- nt
- genbank
- genbank Bacterial
- genbank Environmental sampling
- genbank EST (expressed sequence tag)
- genbank GSS (genome survey sequence)
- genbank HTC (high throughput cDNA sequencing)
- genbank HTGS (high throughput genomic sequencing)

Set expectation value cutoff
0.001

Output format
Tabular (6 columns, 125 lines)

History
search datasets

eba 2016 sartools
42 shown
1.59 MB

- 62: SARTools DESeq2 R objects (.RData)**
- 61: SARTools DESeq2 R log**
- 60: SARTools DESeq2 figures**
- 59: SARTools DESeq2 tables**
- 58: SARTools DESeq2 report**
- 57: SARTools edgeR R objects (.RData)**
- 56: SARTools edgeR R log**
- 55: SARTools edgeR figures**

Hands-on

DATA IMPORT





1. Fetch the files with your internet browser:
<https://tinyurl.com/galaxy-initiation-datasets>

2. Upload files into Galaxy (exons.bed, snps.bed, repeats.bed)
 - a. From disk
 - b. Using FTP

TOOLS

Tools - panel

The screenshot displays the Galaxy / ABiMS web interface. At the top, the navigation bar includes 'Galaxy / ABiMS', 'Analyze Data', 'Workflow', 'Shared Data', 'Visualization', 'Admin', 'Help', 'User', and a 'Using 0%' indicator. The left sidebar, titled 'Tools', contains a search bar and a list of tool categories: 'Get Data', 'Send Data', 'Collection Operations', 'COMMON TOOLS', 'Text Manipulation', 'Filter and Sort', 'Join, Subtract and Group', 'Convert Formats', 'Extract Features', 'Fetch Sequences', 'Statistics', 'Graph/Display Data', and 'Fasta Fastq Manipulation'. Under 'Fasta Fastq Manipulation', there are links for 'Filter sequences by ID from a tabular file', 'FastQC Read Quality reports', and 'FASTQ Groomer convert between various FASTQ quality formats'. The main content area features a green 'Welcome to galaxy3.sb-roscoff.fr' message, an information box with contact details for support.abims@sb-roscoff.fr, the ABiMS logo (Analyses and Bioinformatics for Marine Science), and links to 'Changelog' and 'Tutorials'. A paragraph of text describes Galaxy as an open, web-based platform for data-intensive biomedical research, mentioning its affiliation with Penn State and Emory University. The right sidebar, titled 'History', shows a search bar and a list of datasets: 'Trinity example' (2 shown, 3 deleted, 40.02 KB) and two files, '4: reads.left.fq' and '3: reads.right.fq', each with view, edit, and delete icons.

Tools - panel

The screenshot displays the Galaxy/ABiMS web interface. At the top, the navigation bar includes 'Galaxy / ABiMS', 'Analyze Data', 'Workflow', 'Shared Data', 'Visualization', 'Admin', 'Help', and 'User'. A 'Using 0%' indicator is visible in the top right. The left sidebar, titled 'Tools', contains a search bar and a list of tool categories: 'Get Data', 'Send Data', 'Collection Operations', 'COMMON TOOLS', 'Text Manipulation', 'Filter and Sort', 'Join, Subtract and Group', 'Convert Formats', 'Extract Features', 'Fetch Sequences', 'Statistics', 'Graph/Display Data', and 'Fasta Fastq Manipulation'. Under 'Fasta Fastq Manipulation', specific tools like 'Filter sequences by ID from a tabular file', 'FastQC Read Quality reports', and 'FASTQ Groomer' are listed. The main content area features a green 'Welcome to galaxy3.sb-roscoff.fr' message, an information box with contact details for support.abims@sb-roscoff.fr, and the ABiMS logo with the tagline 'Analyses and Bioinformatics for Marine Science'. A large white text box with a black border is overlaid on the main content, containing the question 'What tools are available?'. Below this box, a paragraph of text describes Galaxy as an open, web-based platform for data-intensive biomedical research, mentioning its affiliation with Penn State and Emory University.

Galaxy is an open, web-based platform for data intensive biomedical research. The Galaxy team is a part of BX at Penn State, and the Biology and Mathematics and Computer Science departments at Emory University. The Galaxy Project is supported in part by NHGRI, NSF, The Huck Institutes of the Life Sciences, The Institute for CyberScience at Penn State, and Emory University.



125+ platforms for using Galaxy: <https://galaxyproject.org/use>
[Servers, clouds, deployable resources]

125+ platforms for using Galaxy: <https://galaxyproject.org/use>
[Servers, clouds, deployable resources]

GENERAL PURPOSE:

usegalaxy.org, usegalaxy.eu, usegalaxy.fr, usegalaxy.org.au

125+ platforms for using Galaxy: <https://galaxyproject.org/use>
[Servers, clouds, deployable resources]

GENERAL PURPOSE:

usegalaxy.org, usegalaxy.eu, usegalaxy.fr, usegalaxy.org.au

DOMAIN SPECIFIC:

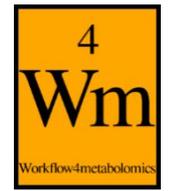


RNAseq: <http://galaxy.sb-roscoff.fr>

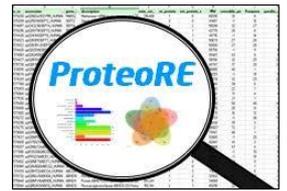
SBR tools: <http://webtools.sb-roscoff.fr>

Metagenomics: <http://galaxy4metab.sb-roscoff.fr>

Metabolomics:



Proteomics:



ChIP-seq:



125+ platforms for using Galaxy: <https://galaxyproject.org/use>
[Servers, clouds, deployable resources]

GENERAL PURPOSE:

usegalaxy.org, usegalaxy.eu, usegalaxy.fr, usegalaxy.org.au

DOMAIN SPECIFIC:

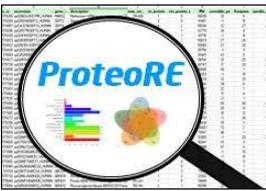


RNAseq: <http://galaxy.sb-roscoff.fr>

SBR tools: <http://webtools.sb-roscoff.fr>

Metagenomics: <http://galaxy4metab.sb-roscoff.fr>

Metabolomics: 

Proteomics: 

ChIP-seq: 




Catalog of French Galaxy tools:
<http://galaxycat.france-bioinformatique.fr>

Tools - panel

Galaxy / ABiMS Analyze Data Workflow Shared Data Visualization Admin Help User Using 0%

Tools

trinity

Trinity suite

1- ASSEMBLY:

[Trinity](#) de novo assembly of RNA-Seq data

[Trinity Statistics](#) Obtain basic stats for the number of genes and isoforms and contiguity of the assembly

[Generate gene to transcript map](#) for Trinity assembly

2- COUNTING:

[Align reads and estimate abundance](#) on a de novo assembly of RNA-Seq data

[Build expression matrix](#) for a de novo assembly of RNA-Seq data by Trinity

3- DIFFERENTIAL EXPRESSION:

[RNASeq samples quality check](#) for transcript quantification

✓ **Welcome to galaxy3.sb-roscoff.fr**

Information
For any question or request for tools or account, send an email at support.abims@sb-roscoff.fr

ABiMS
Analyses and Bioinformatics for Marine Science

Station Biologique Roscoff

Changelog

Tutorials

Galaxy is an open, web-based platform for data intensive biomedical research. The [Galaxy team](#) is a part of [BX](#) at [Penn State](#), and the [Biology and Mathematics and Computer Science](#) departments at [Emory University](#). The [Galaxy Project](#) is supported in part by [NHGRI](#), [NSF](#), [The Huck Institutes of the Life Sciences](#), [The Institute for CyberScience](#) at [Penn State](#), and [Emory University](#).

History

search datasets

Trinity example
2 shown, 3 deleted

40.02 KB

4: reads.left.fq

3: reads.right.fq

Tools - form

Galaxy / ABiMS Analyze Data Workflow Shared Data Visualization Admin Help User Using 0%

Tools

trinity

Trinity suite

1- ASSEMBLY:

[Trinity de novo assembly of RNA-Seq data](#)

[Trinity Statistics](#) Obtain basic stats for the number of genes and isoforms and contiguity of the assembly

[Generate gene to transcript map](#) for Trinity assembly

2- COUNTING:

[Align reads and estimate abundance](#) on a de novo assembly of RNA-Seq data

[Build expression matrix](#) for a de novo assembly of RNA-Seq data by Trinity

3- DIFFERENTIAL EXPRESSION:

[RNASeq samples quality check](#) for transcript quantification

Trinity de novo assembly of RNA-Seq data (Galaxy Version 2.2.0.0) Options

Paired or Single-end data?

Single

Single-end reads

4: reads.left.fq
3: reads.right.fq

(--single)

Strand specific data

Yes No

Run in silico normalization of reads

Yes No

Defaults to max. read coverage of 50. (--normalize_reads)

[Additional Options](#)

Execute

[Trinity](#) assembles transcript sequences from Illumina RNA-Seq data.

Citations [Show BibTeX](#)

Grabherr, Manfred G and Haas, Brian J and Yassour, Moran and Levin, Joshua Z and Thompson, Dawn A and Amit, Ido and Adiconis, Xian and Fan, Lin and Raychowdhury, Raktima and Zeng, Qiandong and et al. (2011). Full length

History

search datasets

Trinity example
2 shown, 2 deleted

37.53 KB

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3: reads.right.fq

Tools - form

Galaxy / ABiMS Analyze Data Workflow Shared Data Visualization Admin Help User Using 0%

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History

search datasets

Trinity example
2 shown, 2 deleted

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4: reads.left.fq

3: reads.right.fq

Tools - form

Tools can have some advanced options

Galaxy / ABiMS Analyze Data Workflow Shared Data Visualization Admin Help User Using 0%

Tools trinity

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Run in silico normalization of reads

Yes No

Defaults to max. read coverage of 50. (--normalize_reads)

Additional Options

Minimum Contig Length

200

All contigs shorter than this will be discarded (--min_contig_length)

Use the genome guided mode?

No

If you already mapped the reads to the genome, Trinity can use this information

Error-corrected or circular consensus (CCS) pac bio reads

Nothing selected

Experimental feature! Long reads must be in the same orientation as short reads if they are strand specific (--long_reads)

Minimum count for K-mers to be assembled

1

(--min_kmer_cov)

Execute

[Trinity](#) assembles transcript sequences from Illumina RNA-Seq data.

Citations Show BibTeX

History search datasets

Trinity example
2 shown, 2 deleted

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Trinity de novo assembly of RNA-Seq data (Galaxy Version 2.2.0.0) Options

Paired or Single-end data?

Paired

Left/Forward strand reads

4: reads.left.fq
3: reads.right.fq

(--left)

Right/Reverse strand reads

4: reads.left.fq
3: reads.right.fq

(--right)

Strand specific data

Yes No

Run in silico normalization of reads

Yes No

Defaults to max. read coverage of 50. (--normalize_reads)

[Additional Options](#)

Execute

History

search datasets

Trinity example
2 shown, 3 deleted

40.02 KB

4: reads.left.fq

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Tools - form

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1 job has been successfully added to the queue - resulting in the following datasets:

5: Trinity on data 3 and data 4: Assembled Transcripts

You can check the status of queued jobs and view the resulting data by refreshing the History pane. When the job has been run the status will change from 'running' to 'finished' if completed successfully or 'error' if problems were encountered.

History

search datasets

Trinity example
3 shown, 2 deleted

37.53 KB

5: Trinity on data 3 and data 4: Assembled Transcripts

4: reads.left.fq

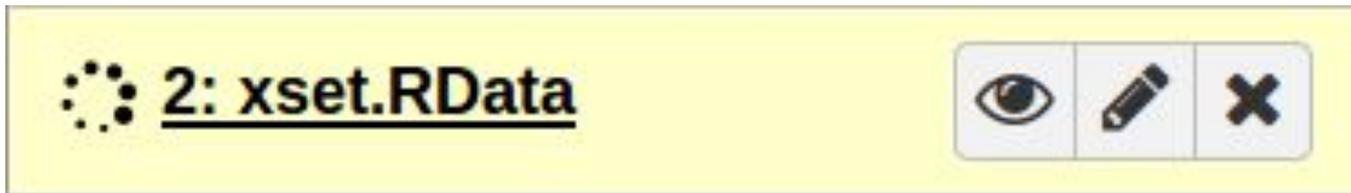
3: reads.right.fq



Job is waiting to run

= the job is in the scheduler « queue »

Duration time of this status depends on the amount of actual queued jobs and on the requested number of processors



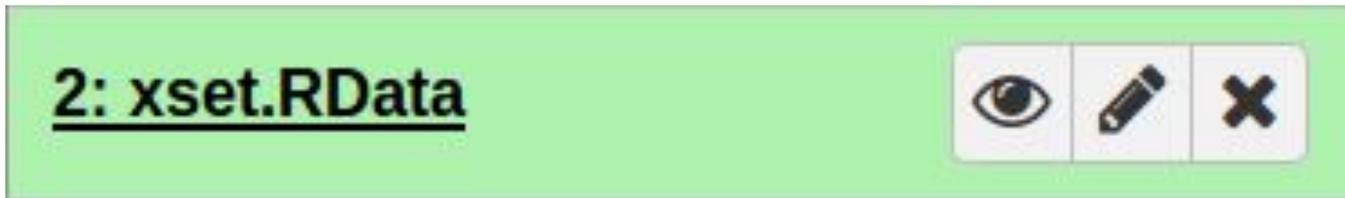
Job is currently running

= the job is being executed on the computing cluster

Duration time of this status depends on the job's attributes and the computing resources allocated.

Some programs are executed with several processors (using 4, 8 or 16 Gb of RAM).

And others are mono-threaded 😞



Job is finished and status is OK

But warnings or errors can be hidden behind!



Job is finished but with an error status

= the program sends an error

The error is often explained by the program but sometimes ...
not.



Job is finished but with an error status

= the program sends an error

Possible causes of error :

- ~~The user~~ :P
- Bad usage : input file, format or option
- Bad integration of the program into Galaxy ... sorry :/
- Non anticipated crash of the program

Tools - Handle errors

The screenshot displays the Galaxy / ABiMS web interface. At the top, the navigation bar includes 'Galaxy / ABiMS', 'Analyze Data', 'Workflow', 'Shared Data', 'Visualization', 'Admin', 'Help', and 'User'. A 'Using 0%' indicator is visible in the top right corner.

The left sidebar contains a 'Tools' section with a search bar and a list of tool categories: 'Get Data', 'Send Data', 'Collection Operations', 'COMMON TOOLS', 'Text Manipulation', 'Filter and Sort', 'Join, Subtract and Group', 'Convert Formats', 'Extract Features', 'Fetch Sequences', 'Statistics', 'Graph/Display Data', 'Fasta Fastq Manipulation', 'COMMON NGS TOOLS', 'NGS:Samtools', 'NGS:Mapping', 'NGS:Bedtools', 'NGS:Picard Tools', 'SEARCHING TOOLS', and 'Diamond'.

The main content area features a green 'Welcome to galaxy3.sb-roscoff.fr' message with a checkmark icon. Below it is an 'Information' box with an 'i' icon, stating: 'For any question or request for tools or account, send an email at support.abims@sb-roscoff.fr'. The center of the page displays the 'ABiMS 4' logo and the text 'Analyses and Bioinformatics for Marine Science', along with the 'Station Biologique Roscoff' logo. Below the logo are links for 'Changelog' and 'Tutorials'.

The bottom of the main content area contains a paragraph: 'Galaxy is an open, web-based platform for data intensive biomedical research. The Galaxy team is a part of BX at Penn State, and the Biology and Mathematics and Computer Science departments at Emory University. The Galaxy Project is supported in part by NHGRI, NSF, The Huck Institutes of the Life Sciences, The Institute for CyberScience at Penn State, and Emory University.'

The right sidebar shows a 'History' panel with a search bar and a list of datasets. The top dataset is 'eba 2016 sartools' (28 shown, 14 deleted, 1.59 MB). Below it is a red error entry: '48: group2_count2.txt' with an eye icon, a pencil icon, and a close icon. Other entries include '47: group2_count1.txt', '46: group1_count2.txt', '45: group1_count1.txt', '44: SARTools edgeR R objects (.RData)', '43: SARTools edgeR R log', '42: SARTools edgeR figures', '41: SARTools edgeR tables', and '40: SARTools edgeR'.

Tools - Handle errors

The screenshot displays the Galaxy/ABiMS web interface. The top navigation bar includes 'Galaxy / ABiMS', 'Analyze Data', 'Workflow', 'Shared Data', 'Visualization', 'Admin', 'Help', and 'User'. A 'Using 0%' indicator is in the top right. The left sidebar contains a 'Tools' section with a search bar and various tool categories like 'Get Data', 'Send Data', 'Collection Operations', 'COMMON TOOLS', 'Text Manipulation', 'Filter and Sort', 'Join, Subtract and Group', 'Convert Formats', 'Extract Features', 'Fetch Sequences', 'Statistics', 'Graph/Display Data', 'Fasta Fastq Manipulation', 'COMMON NGS TOOLS', 'NGS:Samtools', 'NGS:Mapping', 'NGS:Bedtools', 'NGS:Picard Tools', 'SEARCHING TOOLS', and 'Diamond'.

The main content area is titled 'Dataset generation errors' and shows details for 'Dataset 48: group2_count2.txt'. It states: 'The Galaxy framework encountered the following error while attempting to run the tool:'. A code block contains a traceback:

```
Traceback (most recent call last):
  File "/w/galaxy/galaxy3/galaxy/lib/galaxy/jobs/runners/local.py", line
    stdout_file.close()
  File "/opt/python/lib/python2.7/tempfile.py", line 403, in close
    self.unlink(self.name)
OSError: [Errno 2] No such file or directory: '/w/gala
```

The error message is 'failure running job'. Below this, a section titled 'Report this error to the local Galaxy administrators' explains that local administrators review errors and provides a form to report the error, including a field for 'Your email' with the value 'e.gueguen@sb-roscoff.fr'.

The right sidebar shows a 'History' panel with a search bar and a list of datasets. The selected dataset is '48: group2_count2.txt', which is marked as a 'tool error' with the message 'An error occurred with this dataset: failure running job'. Below it, a table shows gene counts:

gene1	1353
gene10	72
gene100	496
gene1000	50

Other datasets in the history include '47: group2_count1.txt', '46: group1_count2.txt', and '45: group1_count1.txt'. A tooltip 'View or report this error' is visible over the error report icon.

Tools - Handle errors

Sent to the support team

The screenshot shows the Galaxy/ABiMS interface. At the top, there is a navigation bar with 'Galaxy / ABiMS' and various menu items: 'Analyze Data', 'Workflow', 'Shared Data', 'Visualization', 'Admin', 'Help', and 'User'. A 'Using 0%' indicator is in the top right. On the left, a 'Tools' sidebar lists categories like 'Get Data', 'Send Data', 'Collection Operations', 'COMMON TOOLS', and 'COMMON NGS TOOLS'. The main area displays an error message: 'Tool execution generated the following error message: failure running job'. Below this is a large heading: 'Report this error to the local Galaxy administrators'. A paragraph explains that local administrators review errors, but users can provide more info and an email address for better support. An 'Error Report' form is shown with a 'Your email' field containing 'loraine.gueguen@sb-roscoff.fr' and a 'Message' text area. A 'Report' button is at the bottom. On the right, a 'History' panel shows a list of datasets. The selected dataset is 'eba 2016 sartools' (1.59 MB). Below it, a red error entry '48: group2_count2.txt' is highlighted, showing a 'tool error' and the message 'An error occurred with this dataset: failure running job'. A table of gene counts is visible below the error message.

Galaxy / ABiMS Analyze Data Workflow Shared Data Visualization Admin Help User Using 0%

Tools

search tools

Get Data
Send Data
Collection Operations

COMMON TOOLS
Text Manipulation
Filter and Sort
Join, Subtract and Group
Convert Formats
Extract Features
Fetch Sequences
Statistics
Graph/Display Data
Fasta Fastq Manipulation

COMMON NGS TOOLS
NGS:Samtools
NGS:Mapping
NGS:Bedtools
NGS:Picard Tools

SEARCHING TOOLS
Diamond

Tool execution generated the following error message:

```
failure running job
```

Report this error to the local Galaxy administrators

Usually the local Galaxy administrators regularly review errors that occur on the server. However, if you would like to provide additional information (such as what you were trying to do when the error occurred) and a contact e-mail address, we will be better able to investigate your problem and get back to you.

Error Report

Your email

Message

Report

History

search datasets

eba 2016 sartools
28 shown, 14 deleted
1.59 MB

48:
group2_count2.txt
tool error
An error occurred with this dataset:
failure running job

1.gene0	2,1813
gene1	1353
gene10	72
gene100	496
gene1000	50

47:
group2_count1.txt

46:
group1_count2.txt

45:
group1_count1.txt

DATASET

Dataset

Both inputs and outputs

The screenshot displays the Galaxy / ABiMS web interface. The top navigation bar includes 'Galaxy / ABiMS', 'Analyze Data', 'Workflow', 'Shared Data', 'Visualization', 'Admin', 'Help', 'User', and a 'Using 0%' indicator. The left sidebar lists various tool categories such as 'Get Data', 'Send Data', 'Collection Operations', 'COMMON TOOLS', 'COMMON NGS TOOLS', and 'SEARCHING TOOLS'. The main content area features a green welcome banner for 'galaxy3.sb-roscoff.fr', an information box with contact details, and the ABiMS logo with the text 'Analyses and Bioinformatics for Marine Science'. A red box highlights a dataset entry in the 'History' panel on the right, labeled '5: Trinity on data 3 and data 4: Assembled Transcripts'. Other entries in the history include '4: reads.left.fq' and '3: reads.right.fq'. The 'Dataset' label is overlaid in red text on the main content area.

Dataset

Informations

Galaxy / ABiMS Analyze Data Workflow Shared Data Visualization Admin Help User Using 0%

Tool: Trinity

Number:	5
Name:	Trinity on data 3 and data 4: Assembled Transcripts
Created:	Wed 01 Mar 2017 03:52:36 PM (UTC)
Filesize:	2.5 KB
Dbkey:	?
Format:	fasta
Galaxy Tool ID:	toolshed.g2.bx.psu.edu/repos/iuc/trinity/trinity/2.2.0.0
Galaxy Tool Version:	2.2.0.0
Tool Version:	
Tool Standard Output:	stdout
Tool Standard Error:	stderr
Tool Exit Code:	0

Input Parameter	Value	Note for rerun
Paired or Single-end data?	paired	
Left/Forward strand reads	4: reads.left.fq	
Right/Reverse strand reads	3: reads.right.fq	
Strand specific data	true	
Strand-specific Library Type	Reverse-Forward	
Jaccard Clip options	Not available.	
Run in silico normalization of reads	True	
additional_params		
Minimum Contig Length	200	
Use the genome guided mode?	no	

galaxy3.sb-roscoff.fr/datasets/c10ec933dc50450a/show_params

History   

search datasets 

Trinity example
3 shown, 3 [deleted](#)
40.3 KB   

5: Trinity on data 3 and data 4: Assembled Transcripts   
7 sequences
format: **fasta**, database: ?
    

View details [2_g1_i1 len=541 path=\[519:0-544](#)

```
GTCTGAATTGCGCATGTAATGCAAGCTTTCCGAGACACAAGTATGG  
TCGCCATTGTGCAAAATATGTGTCTGATAGACCCGCGCTTTCA  
TGACATGAGCGTGSCACCTGAAGACAGGCTGTGGGTGAGAGGSETC  
TGAGTTGTCTTGTATCATCAATAGATGCAAAATTAGATGTAAGAAK
```

4: reads.left.fq   

3: reads.right.fq   

Dataset

Informations

Galaxy / ABiMS Analyze Data Workflow Shared Data Visualization Admin Help User Using 0%

Tool: Trinity

Number:	5
Name:	Trinity on data 3 and data 4: Assembled Transcripts
Created:	Wed 01 Mar 2017 03:52:36 PM (UTC)
Filesize:	2.5 KB
Dbkey:	?
Format:	fasta
Galaxy Tool ID:	toolshed.g2.bx.psu.edu/repos/iuc/trinity/trinity/2.2.0.0
Galaxy Tool Version:	2.2.0.0
Tool Version:	
Tool Standard Output:	stdout
Tool Standard Error:	stderr
Tool Exit Code:	0

Input Parameter	Value	Note for rerun
Paired or Single-end data?	paired	
Left/Forward strand reads	4: reads.left.fq	
Right/Reverse strand reads	3: reads.right.fq	
Strand specific data	true	
Strand-specific Library Type	Reverse-Forward	
Jaccard Clip options	Not available.	
Run in silico normalization of reads	True	
additional_params		
Minimum Contig Length	200	
Use the genome guided mode?	no	

[galaxy3.sb-roscoff.fr/datasets/c10ec933dc50450a/show_params](#)

History search datasets

Trinity example
3 shown, 3 deleted
40.3 KB

5: Trinity on data 3 and data 4: Assembled Transcripts
7 sequences
format: **fasta**, database: ?

[View details](#) @_g1_i1 len=541 path=[519:0-544

```
GTCTGAATTCGCATGTAATGCAGCTTTCCGACAGACACAAGTATGG  
TCGCCATTGTGCAAAATATGTGTCTGATAGACCCGACGCTTTCA  
TGACATGAGCGTGSCACCTGAAGACAGGCTGTGGGTGAGAGGSETC  
TGAGTTGTCTTGTATCATCAATAGATGCAAAATTAGATGTAAGAAK
```

4: reads.left.fq

3: reads.right.fq

Dataset

Download

Galaxy / ABiMS Analyze Data Workflow Shared Data Visualization Admin Help User Using 0%

Tool: Trinity

Number: 5
Name: Trinity on data 3 and data 4: Assembled Transcripts
Created: Wed 01 Mar 2017 03:52:36 PM (UTC)
Filesize: 2.5 KB
Dbkey: ?
Format: fasta
Galaxy Tool ID: toolshed.g2.bx.psu.edu/repos/iuc/trinity/trinity/2.2.0.0
Galaxy Tool Version: 2.2.0.0
Tool Version:
Tool Standard Output: [stdout](#)
Tool Standard Error: [stderr](#)
Tool Exit Code: 0

Input Parameter	Value	Note for rerun
Paired or Single-end data?	paired	
Left/Forward strand reads	4: reads.left.fq	
Right/Reverse strand reads	3: reads.right.fq	
Strand specific data	true	
Strand-specific Library Type	Reverse-Forward	
Jaccard Clip options	Not available.	
Run in silico normalization of reads	True	
additional_params		
Minimum Contig Length	200	
Use the genome guided mode?	no	

galaxy3.sb-roscoff.fr/datasets/c10ec933dc50450a/display?to_ext=fasta

History search datasets Trinity example 3 shown, 3 deleted 40.3 KB

5: Trinity on data 3 and data 4: Assembled Transcripts 7 sequences format: fasta, database: ?

4: reads.left.fq

3: reads.right.fq

Download `DN0_c0_g1_i1 len=541 path=[519:0-544`

```
GTCTGAATTCGCATGTAATGCAGCTTTCCAGACACAAGTATGG  
TCGCCATTGTGCAAAATATGTGTCTGATAGACCCGACGCTTTCA  
TGACATGAGCGTGGCACCTGAAGACAGGCTGTGGGTGAGAGGCT  
TGAGTTGTCTTGATCATCAATAGATGCAAAATTAGATGTAAGAAK
```

Dataset

Re-run a job

The screenshot displays the Galaxy ABiMS interface for running a Trinity de novo assembly job. The main panel shows the tool configuration for "Trinity de novo assembly of RNA-Seq data (Galaxy Version 2.2.0.0)".

Trinity de novo assembly of RNA-Seq data (Galaxy Version 2.2.0.0) Options

Paired or Single-end data?
Paired

Left/Forward strand reads
5: Trinity on data 3 and data 4: Assembled Transcripts
4: reads.left.fq
3: reads.right.fq
(--left)

Right/Reverse strand reads
5: Trinity on data 3 and data 4: Assembled Transcripts
4: reads.left.fq
3: reads.right.fq
(--right)

Strand specific data
Yes No

Strand-specific Library Type
Reverse-Forward
(--SS_lib_type)

Jaccard Clip options
Yes No

... expect high gene density with UTR overlap (--jaccard_clip)

The History panel on the right shows the job history. The top entry is "Trinity example" (3 shown, 3 deleted, 40.3 KB). Below it, the job "5: Trinity on data 3 and data 4: Assembled Transcripts" is highlighted in green, showing 7 sequences in fasta format. A tooltip "Run this job again" is visible over the job entry. Below it, the input datasets "4: reads.left.fq" and "3: reads.right.fq" are also highlighted in green.

galaxy3.sb-roscoff.fr/tool_runner/rerun?id=c10ec933dc50450a

Dataset

Dataset display : text, tabular, pdf, picture, html ...

Galaxy / ABiMS Analyze Data Workflow Shared Data Visualization Admin Help User Using 0%

Tools search tools

Get Data Send Data Collection Operations COMMON TOOLS Text Manipulation Filter and Sort Join, Subtract and Group Convert Formats Extract Features Fetch Sequences Statistics Graph/Display Data Fasta Fastq Manipulation COMMON NGS TOOLS NGS:Samtools NGS:Mapping NGS:Bedtools NGS:Picard Tools SEARCHING TOOLS Diamond

1	2	3	4	5	6
	wt_37_2	wt_37_3	wt_37_1	wt_GSNO_3	wt_GSNO_1
TR24 c0_g1_i1	90.00	67.00	85.00	36.00	35.00
TR2779 c0_g1_i1	186.00	137.00	217.00	147.00	186.00
TR127 c1_g1_i1	9.00	23.00	16.00	2.00	0.00
TR2107 c1_g1_i1	59.00	65.00	47.00	6.00	6.00
TR2011 c5_g1_i1	11.00	4.00	4.00	8.00	5.00
TR4163 c0_g1_i1	368.00	422.00	425.00	172.00	216.00
TR5055 c0_g2_i1	36.00	17.00	27.00	4.00	7.00
TR1449 c0_g1_i1	196.00	230.00	207.00	66.00	113.00
TR1982 c2_g1_i1	7.00	7.00	6.00	4.00	3.00
TR1859 c3_g1_i1	0.00	0.00	1.00	0.00	0.00
TR1492 c0_g1_i2	1895.00	1906.00	1921.00	1104.00	1263.00
TR1122 c0_g1_i1	2.00	3.00	0.00	3.00	0.00
TR2278 c0_g1_i1	497.00	610.00	598.00	333.00	406.00
TR4084 c0_g1_i1	95.00	148.00	86.00	77.00	111.00
TR4761 c0_g1_i1	2089.00	1746.00	1875.00	155.00	174.00
TR3638 c0_g1_i1	647.00	676.00	712.00	117.00	184.00
TR2090 c0_g1_i1	0.00	0.00	0.00	22.00	0.00
TR3854 c0_g1_i1	1878.00	1734.00	1864.00	1775.00	2173.00
TR131 c0_g1_i1	32.00	28.00	31.00	1001.00	1233.00
TR5075 c0_g1_i1	13.00	22.00	21.00	6.00	8.00
TR2182 c3_g2_i6	1.44	2.70	3.84	3.35	0.00
TR3788 c0_g1_i1	17.00	30.00	22.00	91.00	132.00
TR4859 c0_g1_i1	6.00	12.00	8.00	4.00	1.00
TR2487 c0_g1_i1	386.00	383.00	424.00	689.00	866.00

History

- 6: **de results** a list of 3 datasets
- 5: **matrix.counts.matrix** View data
- 4: **input.matrix.wt GSNO vs wt ph8.DESeq2.DE results**
- 3: **input.matrix.wt 37 vs wt ph8 .DESeq2.DE results**
- 2: **input.matrix.wt 37 vs wt GS NO.DESeq2.DE results**
- 1: **samples.txt**

galaxy3.sb-roscoff.fr/datasets/4437d546e349a03a/display/?preview=True

Dataset

Renaming and annotation

Galaxy / ABiMS Analyze Data Workflow Shared Data Visualization Admin Help User Using 0%

Tools search tools

Get Data
Send Data
Collection Operations

COMMON TOOLS
Text Manipulation
Filter and Sort
Join, Subtract and Group
Convert Formats
Extract Features
Fetch Sequences
Statistics
Graph/Display Data
Fasta Fastq Manipulation

COMMON NGS TOOLS
NGS:Samtools
NGS:Mapping
NGS:Bedtools
NGS:Picard Tools

SEARCHING TOOLS
Diamond

Attributes **Convert Format** **Datatype** **Permissions**

Edit Attributes

Name:
matrix.counts.matrix

Info:
uploaded tabular file

Annotation / Notes:
This is my expression matrix.

Add an annotation or notes to a dataset; annotations are available when a history is viewed.

Database/Build:
unspecified (?)

Save
Auto-detect

This will inspect the dataset and attempt to correct the above column values if they are not accurate.

History

8: Extract and cluster differentially expressed transcripts on data 2, data 3, and others

7: Extract and cluster differentially expressed transcripts on data 2, data 3, and others: extracted differentially expressed genes
a list of datasets

6: de results
a list of 3 datasets

5: matrix.counts.matrix
41 lines
format: **txt**, database: ?
uploaded tabular file
Tags: **trinity**
Annotation: This is my expression matrix.
wt_37_2 wt_37_3 wt_37_1 wt_GSMO_3

Edit attributes

galaxy3.sb-roscoff.fr/datasets/4437d546e349a03a/edit

Dataset

Renaming and annotation

The screenshot displays the Galaxy ABiMS interface for editing a dataset. The main panel is titled "Edit Attributes" and contains the following sections:

- Name:** A text input field containing "matrix.counts.matrix".
- Info:** A text input field containing "uploaded tabular file".
- Annotation / Notes:** A text input field containing "This is my expression matrix." Below this field is a note: "Add an annotation or notes to a dataset; annotations are available when a history is viewed."
- Database/Build:** A dropdown menu currently set to "unspecified (?)".

At the bottom of the "Edit Attributes" panel are two buttons: "Save" and "Auto-detect". Below these buttons is a note: "This will inspect the dataset and attempt to correct the above column values if they are not accurate."

On the left side, there is a "Tools" sidebar with a search bar and a list of tool categories including "Get Data", "Send Data", "Collection Operations", "COMMON TOOLS", "Text Manipulation", "Filter and Sort", "Join, Subtract and Group", "Convert Formats", "Extract Features", "Fetch Sequences", "Statistics", "Graph/Display Data", "Fasta Fastq Manipulation", "COMMON NGS TOOLS", "NGS:Samtools", "NGS:Mapping", "NGS:Bedtools", "NGS:Picard Tools", "SEARCHING TOOLS", and "Diamond".

On the right side, there is a "History" panel showing a list of dataset operations. The current dataset is highlighted in green and includes the following information:

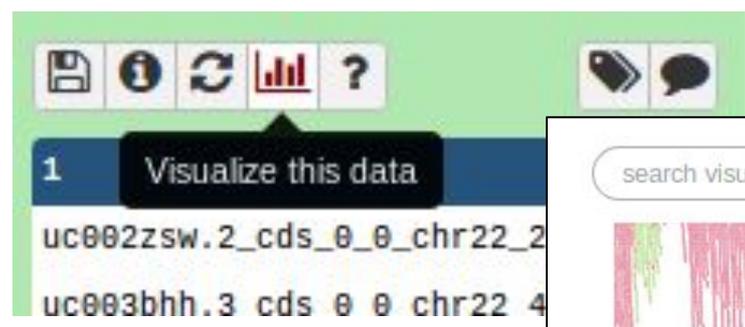
- 5: matrix.counts.matrix** (with an "Edit attributes" tooltip)
- 41 lines
- format: **txt**, database: ?
- uploaded tabular file
- Tags: **trinity** (highlighted with a red box)
- Annotation: This is my expression matrix.

The top navigation bar includes "Galaxy / ABiMS", "Analyze Data", "Workflow", "Shared Data", "Visualization", "Admin", "Help", "User", and a "Using 0%" indicator.

Dataset

Change the Datatype of the Dataset

The screenshot shows the Galaxy / ABiMS interface. The top navigation bar includes 'Galaxy / ABiMS', 'Analyze Data', 'Workflow', 'Shared Data', 'Visualization', 'Admin', 'Help', 'User', and 'Using 0%'. The left sidebar lists tool categories: 'Tools', 'Get Data', 'Send Data', 'Collection Operations', 'COMMON TOOLS', 'Text Manipulation', 'Filter and Sort', 'Join, Subtract and Group', 'Convert Formats', 'Extract Features', 'Fetch Sequences', 'Statistics', 'Graph/Display Data', 'Fasta Fastq Manipulation', 'COMMON NGS TOOLS', 'NGS:Samtools', 'NGS:Mapping', 'NGS:Bedtools', 'NGS:Picard Tools', 'SEARCHING TOOLS', and 'Diamond'. The main panel has tabs for 'Attributes', 'Convert Format', 'Datatype', and 'Permissions'. The 'Datatype' tab is active, showing a 'Change data type' section with a 'New Type:' dropdown menu. The dropdown menu is open, showing options: 'txt', 'supermatcher', 'svg', 'swiss', 'syco', 'tabix', 'table', 'tabular' (highlighted), 'tagseq', and 'tandem'. Below the dropdown, there is a text box with the instruction: 'Changing dataset but *not* modify its contents. Change the type of your dataset.' The right sidebar shows a 'History' panel with a list of datasets: '8: Extract and cluster differentially expressed transcripts on data 2, data 3, and others', '7: Extract and cluster differentially expressed transcripts on data 2, data 3, and others: extracted differentially expressed genes', '6: de results', '5: matrix.counts.matrix', '4: input.matrix.wt GSNO vs wt ph8.DESeq2.DE results', '3: input.matrix.wt 37 vs wt ph8 .DESeq2.DE results', '2: input.matrix.wt 37 vs wt GS NO.DESeq2.DE results', and '1: samples.txt'. Each history entry has icons for view, edit, and delete.







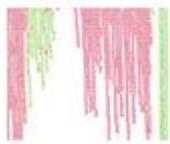


1 Visualize this data

uc002zsw.2_cds_0_0_chr22_2

uc003bhh.3 cds 0 0 chr22 4

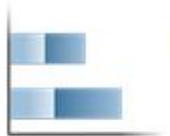
search visualizations



Trackster
Fast, interactive visualization for large, NGS/HTS datasets using only a web browser.



Bar Horizontal (NVD3)
Renders a horizontal bar diagram using NVD3 hosted at <http://www.nvd3.org>.



Bar Horizontal Stacked (NVD3)
Renders a stacked horizontal bar diagram using NVD3 hosted at <http://www.nvd3.org>.



Bar Stacked (NVD3)
Renders a scatter plot using NVD3 hosted at <http://www.nvd3.org>.



Bar diagram (NVD3)
Renders a regular bar diagram using NVD3 hosted at <http://www.nvd3.org>.



Bar diagram (jqPlot)
Renders a bar diagram using jqPlot hosted at <http://www.jqplot.com>.



Box plot (jqPlot)
Processes tabular data using R and renders a box plot using jqPlot hosted at

7: Select first on data 6   

5 lines
 format: **tabular**, database: **hg38**









1	2
uc010gqp.3_cds_0_0_chr22_15690078_f	63
uc011agd.3_cds_0_0_chr22_15528159_f	48
uc062bek.1_cds_0_0_chr22_15690246_f	46
uc003bhh.4_cds_0_0_chr22_46256561_r	30
uc062bej.1_cds_1_0_chr22_15690426_f	26

Tool help for Select first

What it does

This tool outputs specified number of lines from the **beginning** of a dataset

Example

Selecting 2 lines from this:

```
chr7 56632 56652 D17003_CTCF_R6
chr7 56736 56756 D17003_CTCF_R7
chr7 56761 56781 D17003_CTCF_R4
chr7 56772 56792 D17003_CTCF_R7
chr7 56775 56795 D17003_CTCF_R4
```

will produce:

```
chr7 56632 56652 D17003_CTCF_R6
chr7 56736 56756 D17003_CTCF_R7
```

HISTORY

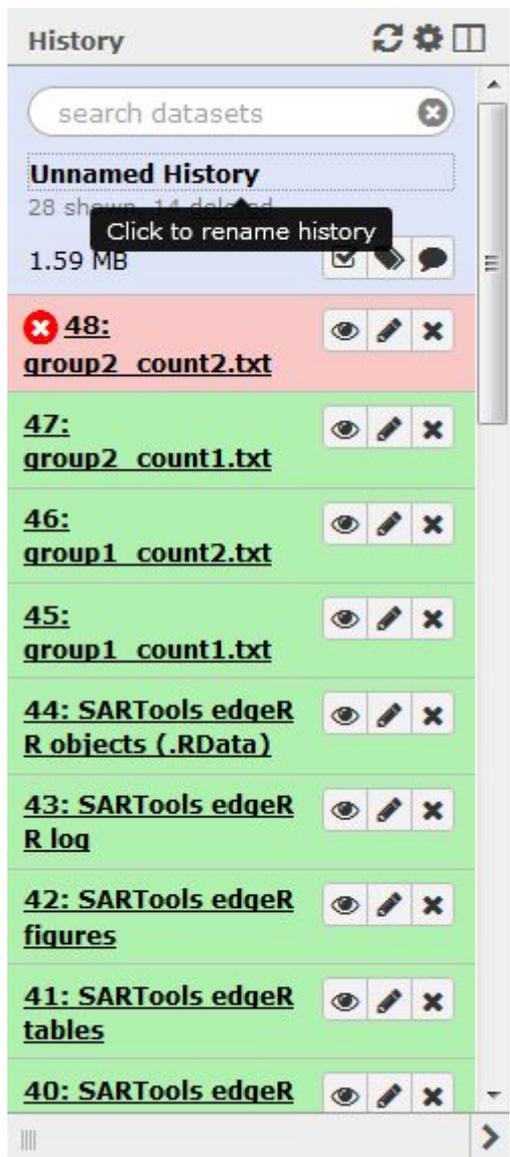
History panel

Both inputs and outputs

The screenshot shows the Galaxy / ABiMS interface. The top navigation bar includes 'Galaxy / ABiMS', 'Analyze Data', 'Workflow', 'Shared Data', 'Visualization', 'Admin', 'Help', 'User', and 'Using 0%'. The left sidebar contains a 'Tools' section with a search bar and various tool categories like 'Get Data', 'Send Data', 'Collection Operations', 'COMMON TOOLS', 'COMMON NGS TOOLS', and 'SEARCHING TOOLS'. The main content area features a 'Welcome to galaxy3.sb-roscoff.fr' message, an 'Information' box, the 'ABiMS 4' logo, and a 'History' label in red. The right sidebar shows the 'History' panel with a search bar and a list of datasets. The dataset '48: group2_count2.txt' is highlighted with a red box.

History

ID	Name	Actions
48	group2_count2.txt	View, Edit, Delete
47	group2_count1.txt	View, Edit, Delete
46	group1_count2.txt	View, Edit, Delete
45	group1_count1.txt	View, Edit, Delete
44	SARTools edgeR R objects (.RData)	View, Edit, Delete
43	SARTools edgeR R log	View, Edit, Delete
42	SARTools edgeR figures	View, Edit, Delete
41	SARTools edgeR tables	View, Edit, Delete
40	SARTools edgeR	View, Edit, Delete



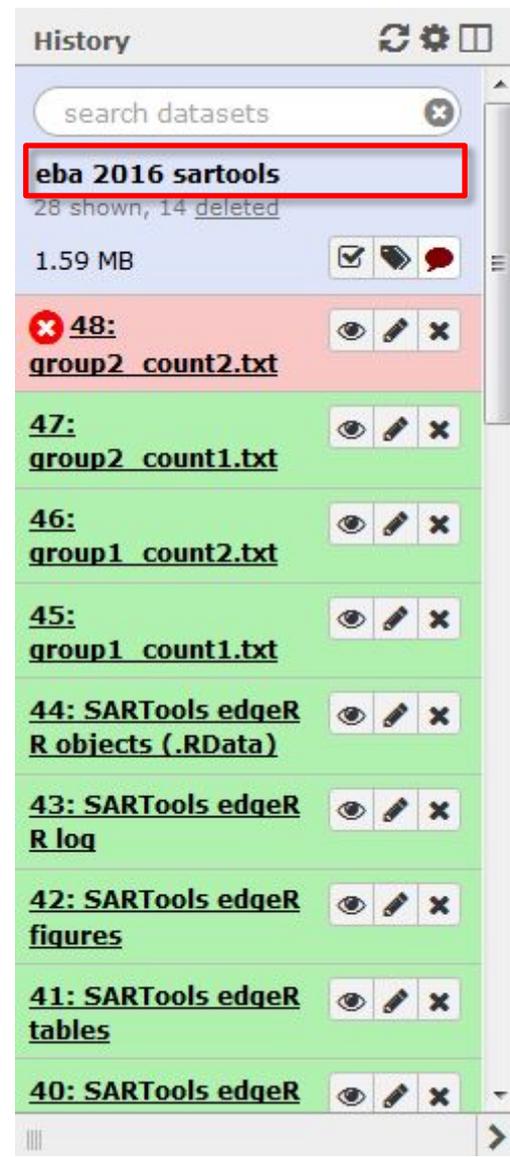
History

search datasets

Unnamed History
28 shown, 14 deleted
1.59 MB

Click to rename history

- 48: group2_count2.txt
- 47: group2_count1.txt
- 46: group1_count2.txt
- 45: group1_count1.txt
- 44: SARTools edgeR R objects (.RData)
- 43: SARTools edgeR R log
- 42: SARTools edgeR figures
- 41: SARTools edgeR tables
- 40: SARTools edgeR

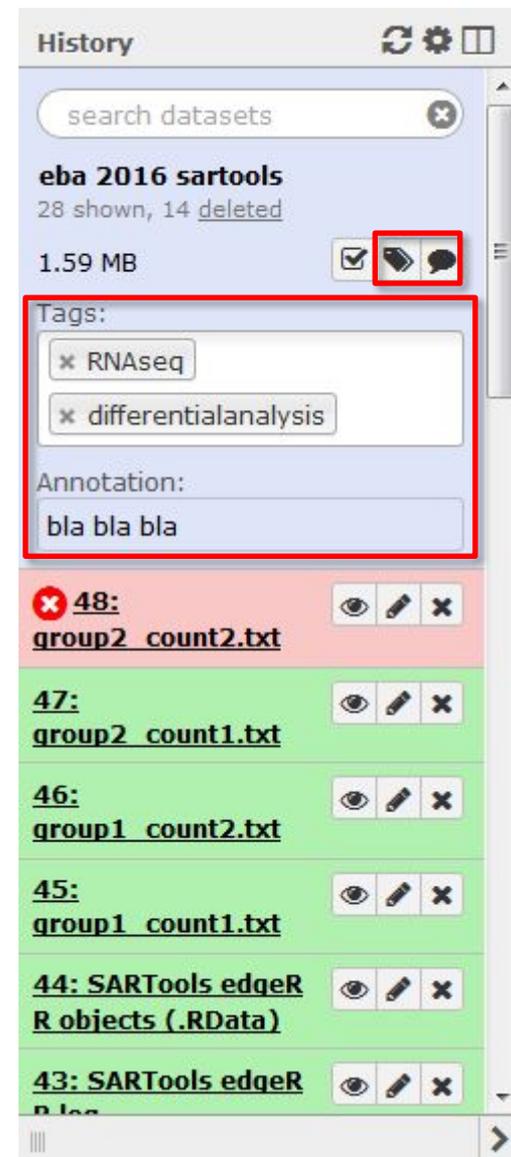


History

search datasets

eba 2016 sartools
28 shown, 14 deleted
1.59 MB

- 48: group2_count2.txt
- 47: group2_count1.txt
- 46: group1_count2.txt
- 45: group1_count1.txt
- 44: SARTools edgeR R objects (.RData)
- 43: SARTools edgeR R log
- 42: SARTools edgeR figures
- 41: SARTools edgeR tables
- 40: SARTools edgeR



History

search datasets

eba 2016 sartools
28 shown, 14 deleted
1.59 MB

Tags:
 x RNAseq
 x differentialanalysis

Annotation:
 bla bla bla

- 48: group2_count2.txt
- 47: group2_count1.txt
- 46: group1_count2.txt
- 45: group1_count1.txt
- 44: SARTools edgeR R objects (.RData)
- 43: SARTools edgeR R log

History panel

History menu: Create new, Rename, Delete, **Delete Permanently**

The screenshot displays the Galaxy / ABiMS web interface. The top navigation bar includes 'Galaxy / ABiMS', 'Analyze Data', 'Workflow', 'Shared Data', 'Visualization', 'Admin', 'Help', and 'User'. A 'Using 0%' indicator is visible in the top right. The left sidebar contains a 'Tools' section with a search bar and various tool categories like 'Get Data', 'Send Data', 'Collection Operations', 'COMMON TOOLS', 'COMMON NGS TOOLS', and 'SEARCHING TOOLS'. The main content area features a green 'Welcome to galaxy3.sb-roscoff.fr' message, an 'Information' box with contact details, and the 'ABiMS 4' logo with the tagline 'Analyses and Bioinformatics for Marine Science'. Below the logo are links for 'Changelog' and 'Tutorials', followed by a paragraph of text about the Galaxy platform. On the right, the 'History' panel is open, showing a menu with categories: 'HISTORY LISTS', 'HISTORY ACTIONS', 'DATASET ACTIONS', 'DOWNLOADS', and 'OTHER ACTIONS'. The 'HISTORY ACTIONS' list includes 'Create New', 'Copy History', 'Share or Publish', 'Show Structure', 'Extract Workflow', 'Delete', and 'Delete Permanently'. The 'DATASET ACTIONS' list includes 'Copy Datasets', 'Dataset Security', 'Resume Paused Jobs', 'Collapse Expanded Datasets', 'Unhide Hidden Datasets', 'Delete Hidden Datasets', and 'Purge Deleted Datasets'. The 'DOWNLOADS' list includes 'Export Tool Citations' and 'Export History to File'. The 'OTHER ACTIONS' list includes 'Import from File'. A red box highlights the gear icon in the top right of the History panel.

History panel

Saved histories

Galaxy / ABiMS Analyze Data Workflow Shared Data Visualization Admin Help User Using 0%

Tools search tools

Get Data Send Data Collection Operations COMMON TOOLS Text Manipulation Filter and Sort Join, Subtract and Group Convert Formats Extract Features Fetch Sequences Statistics Graph/Display Data Fasta Fastq Manipulation COMMON NGS TOOLS NGS:Samtools NGS:Mapping NGS:Bedtools NGS:Picard Tools SEARCHING TOOLS Diamond

search history names and tags

Advanced Search

Name	Datasets	Tags	Sharing	Size on Disk	Created	Last
eba2016 deseq2	8	0 Tags		829.2 KB	Nov 14, 2016	Nov 1
Copy of TP_ITMO2016 shared by xi.liu@sb- roscoff.fr (active items only)	32 2	0 Tags		28.5 MB	Nov 10, 2016	Nov 1
eba 2016 macs2	3 2	0 Tags		602.8 KB	Nov 09, 2016	Nov C
eba 2016 sickle	10	0 Tags		4.2 MB	Oct 04, 2016	Nov C

Page: 1 2 | Show All

For 0 selected histories: Rename Delete Delete Permanently Undele

History

HISTORY LISTS

- Saved Histories
- Histories Shared with Me

HISTORY ACTIONS

- Create New
- Copy History
- Share or Publish
- Show Structure
- Extract Workflow
- Delete
- Delete Permanently

DATASET ACTIONS

- Copy Datasets
- Dataset Security
- Resume Paused Jobs
- Collapse Expanded Datasets
- Unhide Hidden Datasets
- Delete Hidden Datasets
- Purge Deleted Datasets

DOWNLOADS

- Export Tool Citations
- Export History to File

OTHER ACTIONS

- Import from File

Histories that have been deleted for more than a time period specified by the Galaxy administrator(s) may be permanently deleted.

galaxy3.sb-roscoff.fr/history/list

History panel

Saved histories: Switch histories

Galaxy / ABiMS Analyze Data Workflow Shared Data Visualization Admin Help User Using 0%

Tools search tools

Get Data Send Data Collection Operations COMMON TOOLS Text Manipulation Filter and Sort Join, Subtract and Group Convert Formats Extract Features Fetch Sequences Statistics Graph/Display Data Fasta Fastq Manipulation COMMON NGS TOOLS NGS:Samtools NGS:Mapping NGS:Bedtools NGS:Picard Tools SEARCHING TOOLS Diamond

search history names and tags

Advanced Search

Name	Datasets	Tags	Sharing	Size on Disk	Created	Last
eba2016 deseq2	8	0 Tags		829.2 KB	Nov 14, 2016	Nov 14, 2016
Copy of TP_ITMO2016 shared by xi.liu@sb-roscoff.fr (active items only)	32 2	0 Tags		28.5 MB	Nov 10, 2016	Nov 10, 2016
eba 2016 macs2	3 2	0 Tags		602.8 KB	Nov 09, 2016	Nov 09, 2016
eba 2016 sickle	10	0 Tags		4.2 MB	Oct 04, 2016	Nov 04, 2016

Page: 1 2 | Show All

For 0 selected histories: Rename Delete Delete Permanently Undelete

Histories that have been deleted for more than a time period specified by the Galaxy administrator(s) may be permanently deleted.

History HISTORY LISTS Saved Histories Histories Shared with Me HISTORY ACTIONS Create New Copy History Share or Publish Show Structure Extract Workflow Delete Delete Permanently DATASET ACTIONS Copy Datasets Dataset Security Resume Paused Jobs Collapse Expanded Datasets Unhide Hidden Datasets Delete Hidden Datasets Purge Deleted Datasets DOWNLOADS Export Tool Citations Export History to File OTHER ACTIONS Import from File

galaxy3.sb-roscoff.fr/history/list

History panel

History menu: Create new, Rename, Delete, Delete Permanently

The screenshot displays the Galaxy/ABiMS web interface. The top navigation bar includes 'Galaxy / ABiMS', 'Analyze Data', 'Workflow', 'Shared Data', 'Visualization', 'Admin', 'Help', and 'User'. A 'Using 0%' indicator is visible in the top right. The left sidebar contains a 'Tools' section with a search bar and various tool categories like 'Get Data', 'Send Data', 'Collection Operations', 'COMMON TOOLS', 'Text Manipulation', 'Filter and Sort', 'Join, Subtract and Group', 'Convert Formats', 'Extract Features', 'Fetch Sequences', 'Statistics', 'Graph/Display Data', 'Fasta Fastq Manipulation', 'COMMON NGS TOOLS', 'NGS:Samtools', 'NGS:Mapping', 'NGS:Bedtools', 'NGS:Picard Tools', 'SEARCHING TOOLS', and 'Diamond'. The main content area features a green 'Welcome to galaxy3.sb-roscoff.fr' message, an 'Information' box with contact details for support.abims@sb-roscoff.fr, and the ABiMS logo (Analyses and Bioinformatics for Marine Science) with links to 'Changelog' and 'Tutorials'. A paragraph of text describes Galaxy as an open, web-based platform for data-intensive biomedical research, supported by various institutions. On the right, the 'History' panel is open, showing a menu with categories: 'HISTORY LISTS' (Saved Histories, Histories Shared with Me), 'HISTORY ACTIONS' (Create New, Copy History, Share or Publish, Show Structure, Extract Workflow, Delete, Delete Permanently), 'DATASET ACTIONS' (Copy Datasets, Dataset Security, Resume Paused Jobs, Collapse Expanded Datasets, Unhide Hidden Datasets, Delete Hidden Datasets, Purge Deleted Datasets), 'DOWNLOADS' (Export Tool Citations, Export History to File), and 'OTHER ACTIONS' (Import from File). The 'Create New' option in the 'HISTORY ACTIONS' section is highlighted with a red box.

History panel

The screenshot displays the Galaxy / ABiMS web interface. The top navigation bar includes 'Galaxy / ABiMS', 'Analyze Data', 'Workflow', 'Shared Data', 'Visualization', 'Admin', 'Help', 'User', and a 'Using 0%' indicator. The left sidebar contains a 'Tools' section with a search bar and various tool categories: 'Get Data', 'Send Data', 'Collection Operations', 'COMMON TOOLS' (Text Manipulation, Filter and Sort, Join, Subtract and Group, Convert Formats, Extract Features, Fetch Sequences, Statistics, Graph/Display Data, Fasta Fastq Manipulation), and 'COMMON NGS TOOLS' (NGS:Samtools, NGS:Mapping, NGS:Bedtools, NGS:Picard Tools). The main content area features a green welcome message for 'galaxy3.sb-roscoff.fr', an information box with contact details, the ABiMS logo (Analyses and Bioinformatics for Marine Science), and links to 'Changelog' and 'Tutorials'. A paragraph of text describes Galaxy as an open, web-based platform for data-intensive biomedical research, supported by various institutions. The right sidebar shows the 'History' panel, which lists datasets with their names, sizes, and actions. The top of the history panel has a search bar and icons for refresh, settings, and a red-bordered icon. The history list includes: 'eba 2016 sartools' (1.59 MB), '48: group2_count2.txt' (red background), '47: group2_count1.txt' (green background), '46: group1_count2.txt' (green background), '45: group1_count1.txt' (green background), '44: SARTools edgeR R objects (.RData)', '43: SARTools edgeR R log', '42: SARTools edgeR figures', '41: SARTools edgeR tables', and '40: SARTools edgeR'.

History panel

Galaxy / ABiMS Analyze Data Workflow Shared Data Visualization Admin Help User Using 0%

Done search histories search all datasets Create new

Current History

Switch to

eba 2016 sartools
28 shown, 14 deleted
1.59 MB

search datasets

Drag datasets here to copy them to the current history

- 48: group2_count2.txt
- 47: group2_count1.txt
- 46: group1_count2.txt
- 45: group1_count1.txt
- 44: SARTools edgeR R objects (.RData)
- 43: SARTools edgeR R log
- 42: SARTools edgeR figures
- 41: SARTools edgeR tables
- 40: SARTools edgeR report

Switch to

Trinity example
3 shown, 3 deleted
40.3 KB

search datasets

- 5: Trinity on data 3 and data 4: Assembled Transcripts
- 4: reads.left.fq
- 3: reads.right.fq

Switch to

trinity_contig_exn50_statistic
12 shown, 15 deleted
47.01 KB

search datasets

- 14: Build expression matrix on data 7 and data 6: matrix of UpperQuartile-normalized expression values
- 13: Build expression matrix on data 7 and data 6: matrix of TPM expression values (not cross-sample normalized)
- 12: Build expression matrix on data 7 and data 6: estimated RNA-Seq fragment counts (raw counts)
- 9: Build expression matrix on data 7 and data 6: matrix of TPM expression values (not cross-sample normalized)
- 8: Build expression matrix on data 7 and data 6: estimated

Switch to

eba 2016 tr
16 shown
21.92 KB

search da

- 16: Extra differentially transcripts of RData file
- 15: Extract expressed to data 3, and depleted cat a list of data
- 14: Extract expressed to data 3, and a list of data
- 13: Extra differentially transcripts of
- 12: Extra differentially

History panel

The screenshot displays the Galaxy / ABiMS interface with a focus on the history panel. At the top, navigation tabs include 'Analyze Data', 'Workflow', 'Shared Data', 'Visualization', 'Admin', 'Help', and 'User'. A search bar at the top left contains 'search histories' and 'search all datasets'. The main area is divided into several history panels, each with a 'Switch to' button. The first panel, 'eba 2016 sartools', shows a list of datasets from 40 to 48. The second panel, 'Trinity example', shows datasets 3, 4, and 5. The third panel, 'trinity_contig_exn50_statistic', shows datasets 8 to 14. The fourth panel, 'eba 2016 tr...', shows datasets 12 to 16. A red box highlights the 'Switch to' button of the 'Trinity example' panel.

Galaxy / ABiMS Analyze Data Workflow Shared Data Visualization Admin Help User Using 0%

Done search histories search all datasets Create new

Current History Switch to Switch to Switch to Switch to

eba 2016 sartools
28 shown, 14 deleted
1.59 MB
search datasets
Drag datasets here to copy them to the current history

- 48: group2_count2.txt
- 47: group2_count1.txt
- 46: group1_count2.txt
- 45: group1_count1.txt
- 44: SARTools edgeR R objects (.RData)
- 43: SARTools edgeR R log
- 42: SARTools edgeR figures
- 41: SARTools edgeR tables
- 40: SARTools edgeR report

Trinity example
3 shown, 3 deleted
40.3 KB
search datasets

- 5: Trinity on data 3 and data 4: Assembled Transcripts
- 4: reads.left.fq
- 3: reads.right.fq

trinity_contig_exn50_statistic
12 shown, 15 deleted
47.01 KB
search datasets

- 14: Build expression matrix on data 7 and data 6: matrix of UpperQuartile-normalized expression values
- 13: Build expression matrix on data 7 and data 6: matrix of TPM expression values (not cross-sample normalized)
- 12: Build expression matrix on data 7 and data 6: estimated RNA-Seq fragment counts (raw counts)
- 9: Build expression matrix on data 7 and data 6: matrix of TPM expression values (not cross-sample normalized)
- 8: Build expression matrix on data 7 and data 6: estimated

eba 2016 tr...
16 shown
21.92 KB
search da

- 16: Extra differentially transcripts of RData file
- 15: Extract expressed to data 3, and depleted cat a list of data
- 14: Extract expressed to data 3, and a list of data
- 13: Extra differentially transcripts of
- 12: Extra differentially

History panel

Galaxy / ABiMS Analyze Data Workflow Shared Data Visualization Admin Help User Using 0%

Done search histories search all datasets Create new

Current History

Switch to

eba 2016 sartools
28 shown, 14 deleted
1.59 MB

search datasets

Drag datasets here to copy them to the current history

- 48: group2_count2.txt
- 47: group2_count1.txt
- 46: group1_count2.txt
- 45: group1_count1.txt
- 44: SARTools edgeR R objects (.RData)
- 43: SARTools edgeR R log
- 42: SARTools edgeR figures
- 41: SARTools edgeR tables
- 40: SARTools edgeR report

Switch to

Trinity example
3 shown, 3 deleted
40.3 KB

search datasets

- 5: Trinity on data 3 and data 4: Assembled Transcripts
- 4: reads.left.fq
- 3: reads.right.fq

Switch to

trinity_contig_exn50_statistic
12 shown, 15 deleted
47.01 KB

search datasets

- 14: Build expression matrix on data 7 and data 6: matrix of UpperQuartile-normalized expression values
- 13: Build expression matrix on data 7 and data 6: matrix of TPM expression values (not cross-sample normalized)
- 12: Build expression matrix on data 7 and data 6: estimated RNA-Seq fragment counts (raw counts)
- 9: Build expression matrix on data 7 and data 6: matrix of TPM expression values (not cross-sample normalized)
- 8: Build expression matrix on data 7 and data 6: estimated

Switch to

eba 2016 tr
16 shown
21.92 KB

search datasets

- 16: Extra differentially transcripts of RData file
- 15: Extract expressed to data 3, and depleted cat a list of dataset
- 14: Extract expressed to data 3, and a list of dataset
- 13: Extra differentially transcripts of
- 12: Extra differentially

History panel

The screenshot displays the Galaxy / ABiMS interface. At the top, the navigation bar includes 'Galaxy / ABiMS', 'Analyze Data', 'Workflow', 'Shared Data', 'Visualization', 'Admin', 'Help', and 'User'. A 'Using 0%' indicator is on the right. Below the navigation bar, a search bar is highlighted with a red box, containing 'search histories' and 'search all datasets' options. The main area is divided into three panels, each with a 'Switch to' dropdown. The left panel, titled 'Current History', shows a list of datasets including 'eba 2016 sartools' (1.59 MB) and a list of files from 40 to 48. The middle panel, titled 'Trinity example' (40.3 KB), shows files 3, 4, and 5. The right panel, titled 'trinity_contig_exn50_statistic' (47.01 KB), shows files 8 through 16. Each dataset entry includes a search bar, a 'Drag datasets here to copy them to the current history' instruction, and icons for viewing, editing, and deleting.

History panel

Galaxy / ABiMS Analyze Data Workflow Shared Data Visualization Admin Help User

Using 0%

Done search histories search all datasets

Create new

Current History

Switch to

eba 2016 sartools
28 shown, 14 deleted
1.59 MB

search datasets

Drag datasets here to copy them to the current history

- 48: group2_count2.txt
- 47: group2_count1.txt
- 46: group1_count2.txt
- 45: group1_count1.txt
- 44: SARTools edgeR R objects (.RData)
- 43: SARTools edgeR R log
- 42: SARTools edgeR figures
- 41: SARTools edgeR tables
- 40: SARTools edgeR report

Trinity example
3 shown, 3 deleted
40.3 KB

search datasets

- 5: Trinity on data 3 and data 4: Assembled Transcripts
- 4: reads.left.fq
- 3: reads.right.fq

trinity_contig_exn50_statistic
12 shown, 15 deleted
47.01 KB

search datasets

- 14: Build expression matrix on data 7 and data 6: matrix of UpperQuartile-normalized expression values
- 13: Build expression matrix on data 7 and data 6: matrix of TPM expression values (not cross-sample normalized)
- 12: Build expression matrix on data 7 and data 6: estimated RNA-Seq fragment counts (raw counts)
- 9: Build expression matrix on data 7 and data 6: matrix of TPM expression values (not cross-sample normalized)
- 8: Build expression matrix on data 7 and data 6: estimated

eba 2016 tr
16 shown
21.92 KB

search da

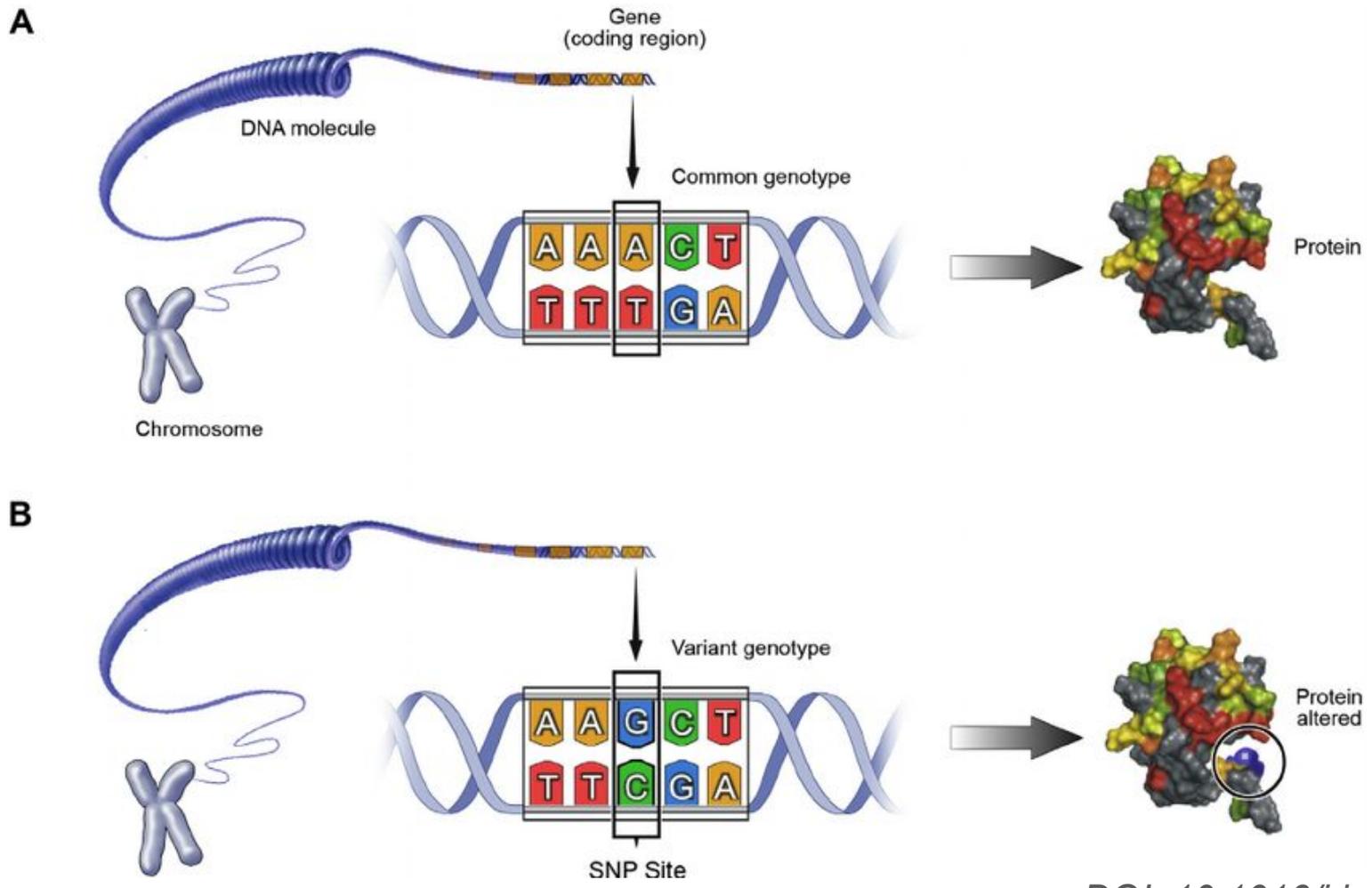
- 16: Extra differentially transcripts of RData file
- 15: Extract expressed to data 3, and depleted cat a list of datase
- 14: Extract expressed to data 3, and a list of datase
- 13: Extra differentially transcripts of
- 12: Extra differentially

Hands-on
TOOLS





Which coding exon has the highest number of single nucleotide polymorphisms on chromosome 22?





Which coding exon has the highest number of single nucleotide polymorphisms on chromosome 22?

exons.bed

Chrom	Start	End	Name	Score	Strand
chr22	16258185	16258303	uc002zlh.1_cds_1_0_chr22_16258186_r	0	-
chr22	16266928	16267095	uc002zlh.1_cds_2_0_chr22_16266929_r	0	-
chr22	16268136	16268181	uc002zlh.1_cds_3_0_chr22_16268137_r	0	-

snps.bed

Chrom	Start	End	Name	Score	Strand
chr22	16266919	16266920	rs757764551	0	+
chr22	16266920	16266920	rs375488594	0	+
chr22	16267014	16267015	rs544633418	0	+
chr22	16267029	16267030	rs563306354	0	+
chr22	16267081	16267082	rs568292779	0	+



Which coding exon has the highest number of single nucleotide polymorphisms on chromosome 22?

exons.bed

Chrom	Start	End	Name	Score	Strand
chr22	16258185	16258303	uc002zlh.1_cds_1_0_chr22_16258186_r	0	-
chr22	16266928	16267095	uc002zlh.1_cds_2_0_chr22_16266929_r	0	-
chr22	16268136	16268181	uc002zlh.1_cds_3_0_chr22_16268137_r	0	-

snps.bed

Chrom	Start	End	Name	Score	Strand
chr22	16266919	16266920	rs757764551	0	+
chr22	16266920	16266920	rs375488594	0	+
chr22	16267014	16267015	rs544633418	0	+
chr22	16267029	16267030	rs563306354	0	+
chr22	16267081	16267082	rs568292779	0	+

exons



snps





Which coding exon has the highest number of single nucleotide polymorphisms on chromosome 22?

exons.bed

Chrom	Start	End	Name	Score	Strand
chr22	16258185	16258303	uc002zlh.1_cds_1_0_chr22_16258186_r	0	-
chr22	16266928	16267095	uc002zlh.1_cds_2_0_chr22_16266929_r	0	-
chr22	16268136	16268181	uc002zlh.1_cds_3_0_chr22_16268137_r	0	-

snps.bed

Chrom	Start	End	Name	Score	Strand
chr22	16266919	16266920	rs757764551	0	+
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chr22	16267081	16267082	rs568292779	0	+

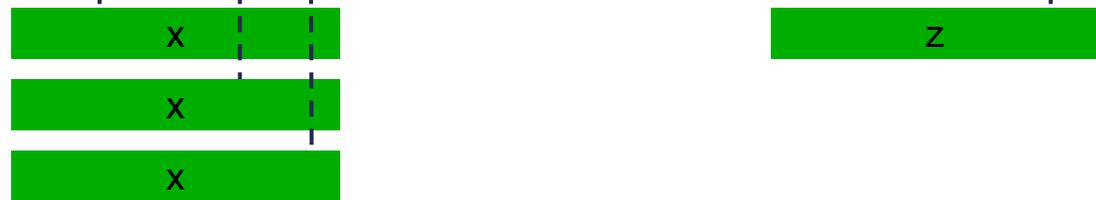
exons



snps



intersect





Which coding exon has the highest number of single nucleotide polymorphisms on chromosome 22?

exons.bed

Chrom	Start	End	Name	Score	Strand
chr22	16258185	16258303	uc002zlh.1_cds_1_0_chr22_16258186_r	0	-
chr22	16266928	16267095	uc002zlh.1_cds_2_0_chr22_16266929_r	0	-
chr22	16268136	16268181	uc002zlh.1_cds_3_0_chr22_16268137_r	0	-

snps.bed

Chrom	Start	End	Name	Score	Strand
chr22	16266919	16266920	rs757764551	0	+
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chr22	16267029	16267030	rs563306354	0	+
chr22	16267081	16267082	rs568292779	0	+

exons



snps



intersect



exon	count
x	3
z	1
...	...



Which coding exon has the highest number of single nucleotide polymorphisms on chromosome 22?

exons.bed

Chrom	Start	End	Name	Score	Strand
chr22	16258185	16258303	uc002zlh.1_cds_1_0_chr22_16258186_r	0	-
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chr22	16268136	16268181	uc002zlh.1_cds_3_0_chr22_16268137_r	0	-

snps.bed

Chrom	Start	End	Name	Score	Strand
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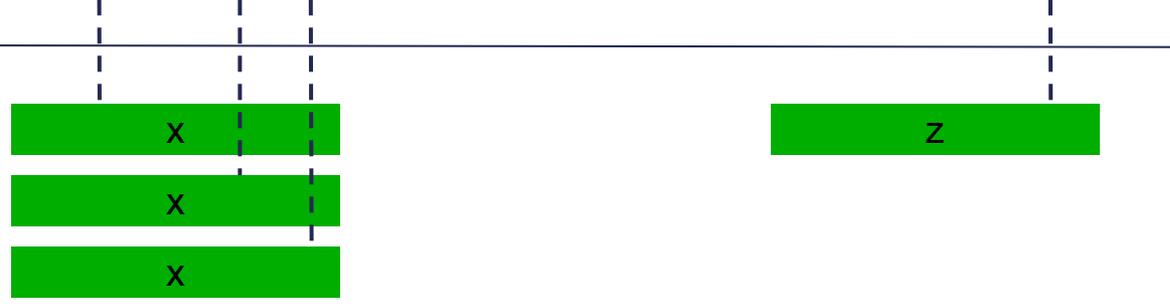
exons



snps



intersect



exon	count
x	3
z	1
...	...

↑
sort



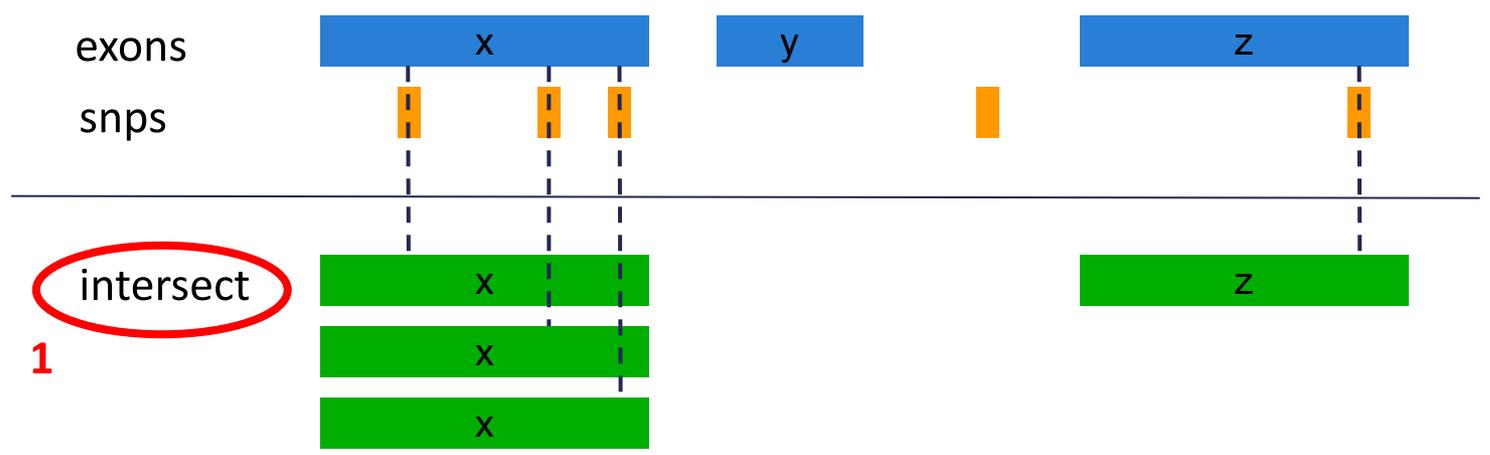
Which coding exon has the highest number of single nucleotide polymorphisms on chromosome 22?

exons.bed

Chrom	Start	End	Name	Score	Strand
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chr22	16266928	16267095	uc002zlh.1_cds_2_0_chr22_16266929_r	0	-
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snps.bed

Chrom	Start	End	Name	Score	Strand
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chr22	16267029	16267030	rs563306354	0	+
chr22	16267081	16267082	rs568292779	0	+



2

exon	count
x	3
z	1
...	...

sort

3



Which coding exon has the highest number of single nucleotide polymorphisms on chromosome 22?

1. Get the data in a new history
2. Intersect exons with SNPs
3. Count the number of SNPs per exon
4. Sort exons by SNP count
5. Select top five
6. Recover exon info
7. Display data in UCSC genome browser



Which coding exon has the highest number of single nucleotide polymorphisms on chromosome 22?

1. Get the data in a new history
2. Intersect exons with SNPs *Bedtools -> Intersect intervals*
3. Count the number of SNPs per exon
4. Sort exons by SNP count
5. Select top five
6. Recover exon info
7. Display data in UCSC genome browser

Q : For the first 3 exons in your file, what is the number of SNPs that fall into that exon?



Which coding exon has the highest number of single nucleotide polymorphisms on chromosome 22?

1. Get the data in a new history
2. Intersect exons with SNPs *Bedtools -> Intersect intervals*
3. Count the number of SNPs per exon *Join, Subtract, and Group -> Group*
4. Sort exons by SNP count
5. Select top five
6. Recover exon info
7. Display data in UCSC genome browser

Q : How many exons are there in total in your file?



Which coding exon has the highest number of single nucleotide polymorphisms on chromosome 22?

1. Get the data in a new history
2. Intersect exons with SNPs *Bedtools -> Intersect intervals*
3. Count the number of SNPs per exon *Join, Subtract, and Group -> Group*
4. Sort exons by SNP count *Filter and Sort -> Sort*
5. Select top five
6. Recover exon info
7. Display data in UCSC genome browser

Q : Which exon has the highest number of SNPs in your file?



Which coding exon has the highest number of single nucleotide polymorphisms on chromosome 22?

1. Get the data in a new history
2. Intersect exons with SNPs *Bedtools -> Intersect intervals*
3. Count the number of SNPs per exon *Join, Subtract, and Group -> Group*
4. Sort exons by SNP count *Filter and Sort -> Sort*
5. Select top five *Text Manipulation -> Select First*
6. Recover exon info
7. Display data in UCSC genome browser



Which coding exon has the highest number of single nucleotide polymorphisms on chromosome 22?

1. Get the data in a new history
2. Intersect exons with SNPs *Bedtools -> Intersect intervals*
3. Count the number of SNPs per exon *Join, Subtract, and Group -> Group*
4. Sort exons by SNP count *Filter and Sort -> Sort*
5. Select top five *Text Manipulation -> Select First*
6. Recover exon info *Join, Subtract and Group -> Compare two Datasets*
7. Display data in UCSC genome browser

BED file

Chrom	Start	End	Name	Score	Strand
chr22	16258185	16258303	uc002zlh.1_cds_1_0_chr22_16258186_r	0	-
chr22	16266928	16267095	uc002zlh.1_cds_2_0_chr22_16266929_r	0	-



Cleanup

DATASET



Dataset

Delete a dataset

The screenshot shows the Galaxy / ABiMS web interface. The top navigation bar includes 'Galaxy / ABiMS', 'Analyze Data', 'Workflow', 'Shared Data', 'Visualization', 'Admin', 'Help', 'User', and a 'Using 0%' indicator. The left sidebar contains a 'Tools' section with a search bar and various tool categories like 'Get Data', 'Send Data', 'Collection Operations', 'COMMON TOOLS', 'COMMON NGS TOOLS', and 'SEARCHING TOOLS'. The main content area features a green welcome banner for 'galaxy3.sb-roscoff.fr', an information box with contact details, the ABiMS logo (Analyses and Bioinformatics for Marine Science), and a list of links for 'Changelog' and 'Tutorials'. The right sidebar shows the 'History' panel with a search bar and a list of datasets. The dataset '62: SARTools DESeq2 R objects (.RData)' is highlighted in green, and a 'Delete' button is overlaid on its 'x' icon. Other datasets in the history include '61: SARTools DESeq2 R log', '60: SARTools DESeq2 figures', '59: SARTools DESeq2 tables', '58: SARTools DESeq2 report', '57: SARTools edgeR R objects (.RData)', '56: SARTools edgeR R log', and '55: SARTools edgeR figures'. The bottom of the page shows a JavaScript console with 'javascript:void(0);'.

Dataset



The dataset isn't really deleted. It's in the Trash

Galaxy / ABiMS Analyze Data Workflow Shared Data Visualization Admin Help User Using 0%

Tools search tools

- Get Data
- Send Data
- Collection Operations
- COMMON TOOLS
 - Text Manipulation
 - Filter and Sort
 - Join, Subtract and Group
 - Convert Formats
 - Extract Features
 - Fetch Sequences
 - Statistics
 - Graph/Display Data
 - Fasta Fastq Manipulation
- COMMON NGS TOOLS
 - NGS:Samtools
 - NGS:Mapping
 - NGS:Bedtools
 - NGS:Picard Tools
- SEARCHING TOOLS
 - Diamond

javascript:void(0);

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ABiMS⁴
Analyses and Bioinformatics for Marine Science

Station Biologique Roscoff

- Changelog
- Tutorials

Galaxy is an open, web-based platform for data intensive biomedical research. The [Galaxy team](#) is a part of [BX](#) at [Penn State](#), and the [Biology](#) and [Mathematics and Computer Science](#) departments at [Emory University](#). The [Galaxy Project](#) is supported in part by [NHGRI](#), [NSF](#), [The Huck Institutes of the Life Sciences](#), [The Institute for CyberScience](#) at Penn State, and [Emory University](#).

History search datasets

eba 2016 sartools
42 shown, [hide deleted](#)
1.59 MB

⚠ This dataset has been deleted
[Undelete it](#)
[Permanently remove it from disk](#)

- 62: SARTools DESeq2 R objects (.RData)
- 61: SARTools DESeq2 R log
- 60: SARTools DESeq2 figures
- 59: SARTools DESeq2 tables
- 58: SARTools DESeq2 report
- 57: SARTools edgeR R objects (.RData)
- 56: SARTools edgeR

Dataset



“Empty Trash” : to free up disk space

Galaxy / ABiMS Analyze Data Workflow Shared Data Visualization Admin Help User Using 0%

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History

- HISTORY LISTS
 - Saved Histories
 - Histories Shared with Me
- HISTORY ACTIONS
 - Create New
 - Copy History
 - Share or Publish
 - Show Structure
 - Extract Workflow
 - Delete
 - Delete Permanently
- DATASET ACTIONS
 - Copy Datasets
 - Dataset Security
 - Resume Paused Jobs
 - Collapse Expanded Datasets
 - Unhide Hidden Datasets
 - Delete Hidden Datasets
 - Purge Deleted Datasets
- DOWNLOADS
 - Export Tool Citations
 - Export History to File
- OTHER ACTIONS
 - Import from File

galaxy3.sb-roscoff.fr/history/purge_deleted_datasets

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Datatypes

DATASET

Dataset - Datatypes

- Every Galaxy dataset is associated with a datatype.
- Datatype can be detected or user specified.

The screenshot shows the Galaxy web interface. On the left, the 'Tools' sidebar is visible with a red box around the 'Upload from disk' icon. The main panel is titled 'Download from web or upload from disk' and shows a table of files in the queue. The file 'AR-80-50K.m80.s2.fa' (5.1 MB) is listed with 'Auto-detect' as its type. A dropdown menu for the 'Type' column is open, showing a list of datatypes: 'Auto-detect', 'ab1', 'affybatch', 'arff', 'asn1', 'asn1-binary', 'axt', and 'bam'. The 'Auto-detect' option is highlighted with a red box, and the entire dropdown menu is circled in red. At the bottom, there are buttons for 'Choose local file', 'Paste/Fetch data', 'Pause', 'Reset', 'Start', and 'Close'.

Name	Size	Type	Genome	Settings	Status
AR-80-50K.m80.s2.fa	5.1 MB	Auto-detect	unspecified (?)		

Common text formats:

- ***txt***: plain text ('.txt')
- ***tabular***: tab delimited ('.tab', '.txt', etc.)

	wt_37_2	wt_37_3	wt_37_1
TR24 c0_g1_i1	90.00	67.00	85.00
TR2779 c0_g1_i1	186.00	137.00	217.00
TR127 c1_g1_i1	9.00	23.00	16.00

- ***csv***: comma-separated values ('.csv')
- ***html***: standard language for web pages

```
Year,Make,Model  
1997,Ford,E350  
2000,Mercury,Cougar
```

```
<!DOCTYPE html>  
<html>  
  <head>  
    <title>This is a  
title</title>  
  </head>  
  <body>  
    <p>Hello world!</p>  
  </body>  
</html>
```

Galaxy / ABiMS Analyze Data Workflow Shared Data Visualization Admin Help User Using 0%

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NGS:Mapping

NGS:Bedtools

NGS:Picard Tools

SEARCHING TOOLS

Diamond

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Text Manipulation

- [Add column to an existing dataset](#)
- [Concatenate datasets tail-to-head](#)
- [Cut columns from a table](#)
- [Merge Columns together](#)
- [Convert delimiters to TAB](#)
- [Create single interval as a new dataset](#)
- [Change Case of selected columns](#)
- [Paste two files side by side](#)
- [Remove beginning of a file](#)
- [Select random lines from a file](#)

Changelog

Tutorials

Galaxy is an open source platform for collaborative and reproducible biomedical research. The Galaxy team is a multidisciplinary team of Biologists, Computer Scientists, and Mathematicians. The Galaxy Project is supported by the French National Research Agency (ANR), the National Center for Genome Research (NCGR), the National Science Foundation (NSF), the National Institutes of Health (NIH), the National Natural Science Foundation of China (NSFC), the National Institute of Standards and Technology (NIST), the National Institute of Environmental Health Sciences (NIEHS), the National Institute of Diabetes and Digestive and Kidney Diseases (NIDDK), the National Institute of Mental Health (NIMH), the National Institute on Drug Abuse (NIDA), the National Institute on Aging (NIA), the National Institute on Cancer Research (NICR), the National Institute on Child Health and Human Development (NICHD), the National Institute on Deafness and Other Communication Disorders (NIDCD), the National Institute on Drug Abuse (NIDA), the National Institute on Environmental Health Sciences (NIEHS), the National Institute on Health and Human Development (NIHHD), the National Institute on Mental Health (NIMH), the National Institute on Neurological Disorders and Stroke (NINDS), the National Institute on Nursing Research (NINR), the National Institute on Substance Abuse (NISA), the National Institute on Tobacco Use (NITU), the National Institute on Vision Research (NIVR), the National Institute on Aging (NIA), the National Institute on Cancer Research (NICR), the National Institute on Child Health and Human Development (NICHD), the National Institute on Deafness and Other Communication Disorders (NIDCD), the National Institute on Drug Abuse (NIDA), the National Institute on Environmental Health Sciences (NIEHS), the National Institute on Health and Human Development (NIHHD), the National Institute on Mental Health (NIMH), the National Institute on Neurological Disorders and Stroke (NINDS), the National Institute on Nursing Research (NINR), the National Institute on Substance Abuse (NISA), the National Institute on Tobacco Use (NITU), the National Institute on Vision Research (NIVR).

History

search datasets

eba 2016 sartools
42 shown
1.59 MB

- 62: SARTools DESeq2 R objects (.RData)**
- 61: SARTools DESeq2 R log**
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- 56: SARTools edgeR R log**
- 55: SARTools edgeR figures**

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SEARCHING TOOLS

Diamond

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Filter and Sort

Filter data on any column using simple expressions

Sort data in ascending or descending order

Select lines that match an expression

GFF

Extract features from GFF data

Filter GFF data by attribute using simple expressions

Filter GFF data by feature count using simple expressions

Filter GTF data by attribute

Changelog

Tutorials

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Join, Subtract and Group

Join two Datasets side by side on a specified field

Compare two Datasets to find common or distinct rows

Group data by a column and perform aggregate operation on other columns.

Analyses and Bioinformatics for Marine Science

► Changelog

► Tutorials

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- 56: SARTools edgeR R log
- 55: SARTools edgeR figures

Common binary formats:

- ***data***: generic binary format
- ***zip, tar***: archives
- ***pdf, png, jpg, bmp, tiff, gif***: images
- ***rdata***: statistical computing program R
- ***bam, wig, bigwig***: sequence alignment

Sequence file formats:

- ***fasta***: a single-line description with '>', followed by lines of sequence data ('.fasta', '.fas')

```
>sequence1
atgcgtttgcgtgcatgCGtttgcgtgcatgCGtttgcgtgcatgCGtttgcgtg
c
atgCGtttgcgtgc
>sequence2
tttcgtgCGtatagtttcgtgCGtatagtttcgtgCGtatagtttcgtgCGtata
g
tggcgcggt
```

- ***fastq***: sequence + quality score ('.fastq', '.fq')

```
@SEQ_ID
GATTTGGGGTTCAAAGCAGTATCGATCAAATAGTAAATCCATTTGTTCAACTCACAGTT
T
+
!' '*((( (***) )%%%++) (%%%) .1***-+*'') ) **55CCF>>>>>CCCCCCC6
5
@SEQ_ID2
GATTTGGGGTTCAAAGCAGTATCGATCAAATAGTAAATCCATTTGTTCAACTCACAGTT
T
+
!' '*((( (***) )%%%++) (%%%) .1***-+*'') ) **55CCF>>>>>CCCCCCC6
```

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Galaxy

Comput

in part

CyberS

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- 58: SARTools DESeq2 report**
- 57: SARTools edgeR R objects (.RData)**
- 56: SARTools edgeR R log**
- 55: SARTools edgeR figures**

Fasta Fastq Manipulation

- [faSplit](#) Split a FASTA file
- [FastQC](#) Read Quality reports
- [Filter sequences by length](#)
- [FASTQ Groomer](#) convert between various FASTQ quality formats
- [Filter sequences by ID](#) from a tabular file

Sequence file formats:

- ***gff3, bed, genbank***: sequence + annotations

bed

```
track name=pairedReads description="Clone Paired Reads"  
useScore=1  
chr22 1000 5000 cloneA 960 + 1000 5000 0 2 567,488, 0,3512  
chr22 2000 6000 cloneB 900 - 2000 6000 0 2 433,399, 0,3601
```

gff3

```
##gff-version 3  
ctg123 . exon 1300 1500 . + .  
ID=exon00001  
ctg123 . exon 1050 1500 . + .  
ID=exon00002  
ctg123 . exon 3000 3902 . + .  
ID=exon00003  
##FASTA  
>ctg123  
cttctgggcgtacccgattctcggagaacttgccgcaccattccgcctt  
g  
tgttcattgctgctgcatgttcattgtctacctcggctacgtgtggct
```

<https://genome.ucsc.edu/FAQ/FAQformat.html#format3>

...

Dataset - Datatypes

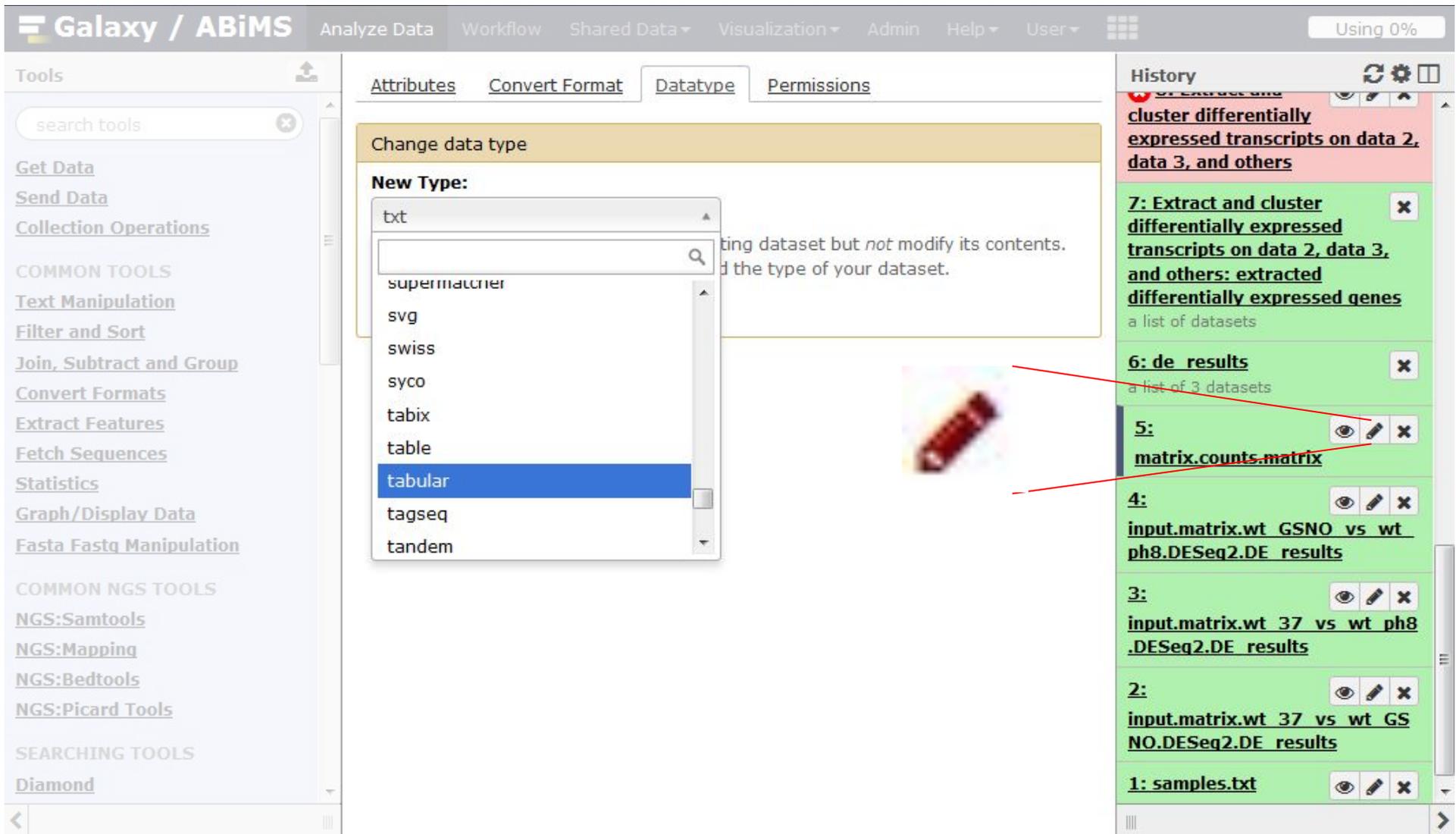
- Every Galaxy dataset is associated with a datatype.
- Datatype can be detected or user specified.

The screenshot shows the Galaxy web interface. On the left, the 'Tools' sidebar is visible with a red box around the 'Upload from disk' icon. The main panel is titled 'Download from web or upload from disk' and shows a table of files in the queue. The file 'AR-80-50K.m80.s2.fa' (5.1 MB) is listed with 'Auto-detect' as its type. A dropdown menu for the 'Type' column is open, showing a list of datatypes: 'Auto-detect', 'ab1', 'affybatch', 'arff', 'asn1', 'asn1-binary', 'axt', and 'bam'. The 'Auto-detect' option is highlighted with a red box, and the entire dropdown menu is circled in red. At the bottom, there are buttons for 'Choose local file', 'Paste/Fetch data', 'Pause', 'Reset', 'Start', and 'Close'.

Name	Size	Type	Genome	Settings	Status
AR-80-50K.m80.s2.fa	5.1 MB	Auto-detect	unspecified (?)		

Dataset - Datatypes

- Every Galaxy dataset is associated with a datatype.
- Datatype can be detected or user specified.



Galaxy / ABiMS Analyze Data Workflow Shared Data Visualization Admin Help User Using 0%

Tools

search tools

Get Data
Send Data
Collection Operations

COMMON TOOLS
Text Manipulation
Filter and Sort
Join, Subtract and Group
Convert Formats
Extract Features
Fetch Sequences
Statistics
Graph/Display Data
Fasta Fastq Manipulation

COMMON NGS TOOLS
NGS:Samtools
NGS:Mapping
NGS:Bedtools
NGS:Picard Tools

SEARCHING TOOLS
Diamond

Attributes Convert Format **Datatype** Permissions

Change data type

New Type:

txt
supermatcrier
svg
swiss
syco
tabix
table
tabular
tagseq
tandem

...ing dataset but *not* modify its contents.
...d the type of your dataset.

History

6: Extract and cluster differentially expressed transcripts on data 2, data 3, and others

7: Extract and cluster differentially expressed transcripts on data 2, data 3, and others: extracted differentially expressed genes
a list of datasets

6: de results
a list of 3 datasets

5: **matrix.counts.matrix**

4: input.matrix.wt GSNO vs wt ph8.DESeq2.DE results

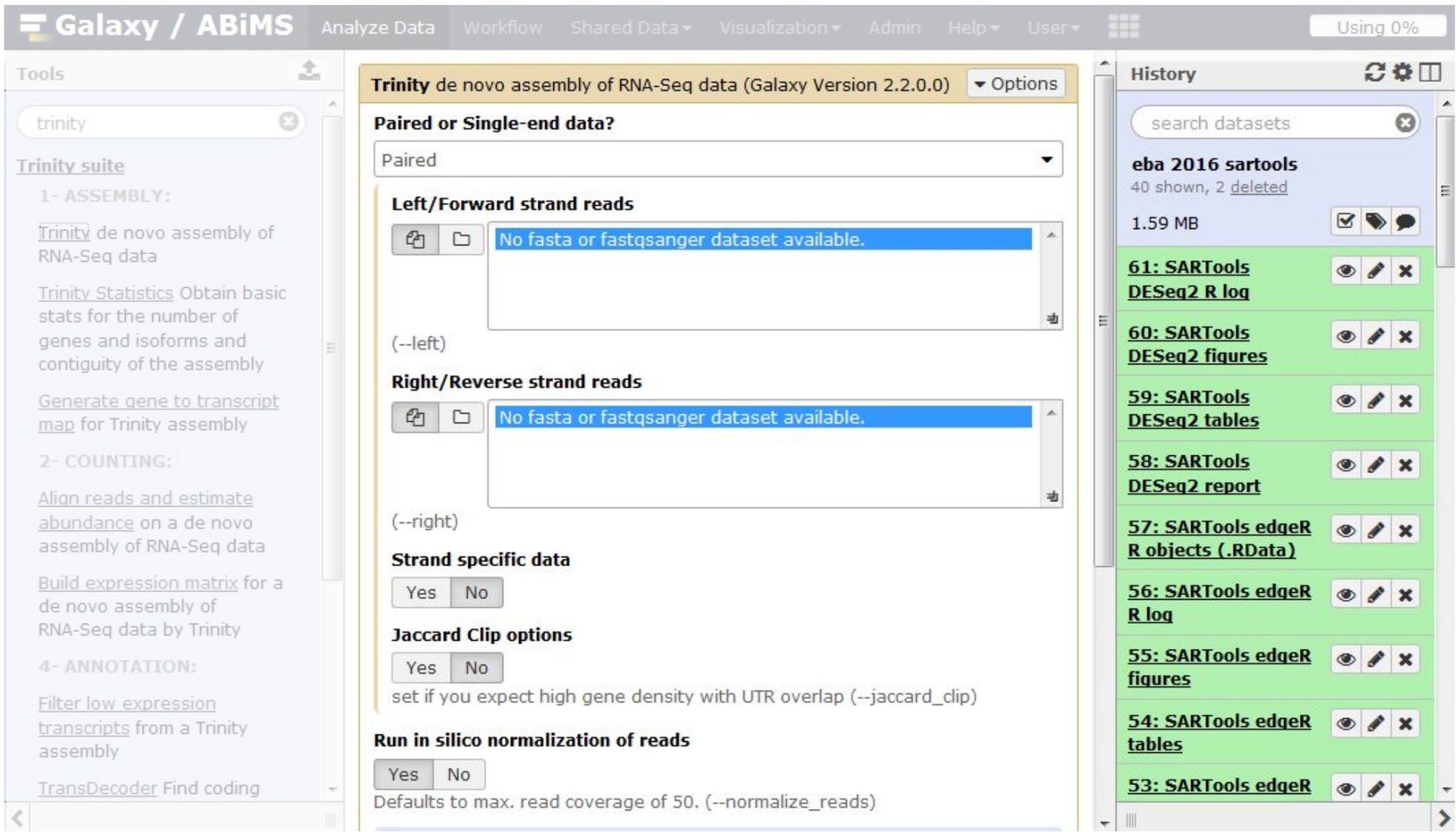
3: input.matrix.wt 37 vs wt ph8.DESeq2.DE results

2: input.matrix.wt 37 vs wt GSNO.DESeq2.DE results

1: samples.txt

Dataset - Datatypes

- Many tools will only accept input datasets with the appropriate datatype assigned.



Galaxy / ABiMS Analyze Data Workflow Shared Data Visualization Admin Help User Using 0%

Tools

trinity

Trinity suite

1- ASSEMBLY:

[Trinity](#) de novo assembly of RNA-Seq data

[Trinity Statistics](#) Obtain basic stats for the number of genes and isoforms and contiguity of the assembly

[Generate gene to transcript map](#) for Trinity assembly

2- COUNTING:

[Align reads and estimate abundance](#) on a de novo assembly of RNA-Seq data

[Build expression matrix](#) for a de novo assembly of RNA-Seq data by Trinity

4- ANNOTATION:

[Filter low expression transcripts](#) from a Trinity assembly

[TransDecoder](#) Find coding

Trinity de novo assembly of RNA-Seq data (Galaxy Version 2.2.0.0) Options

Paired or Single-end data?

Paired

Left/Forward strand reads

No fasta or fastqsanger dataset available.

(--left)

Right/Reverse strand reads

No fasta or fastqsanger dataset available.

(--right)

Strand specific data

Yes No

Jaccard Clip options

Yes No

set if you expect high gene density with UTR overlap (--jaccard_clip)

Run in silico normalization of reads

Yes No

Defaults to max. read coverage of 50. (--normalize_reads)

History

search datasets

eba 2016 sartools
40 shown, 2 deleted
1.59 MB

61: SARTools DESeq2 R log

60: SARTools DESeq2 figures

59: SARTools DESeq2 tables

58: SARTools DESeq2 report

57: SARTools edgeR R objects (.RData)

56: SARTools edgeR R log

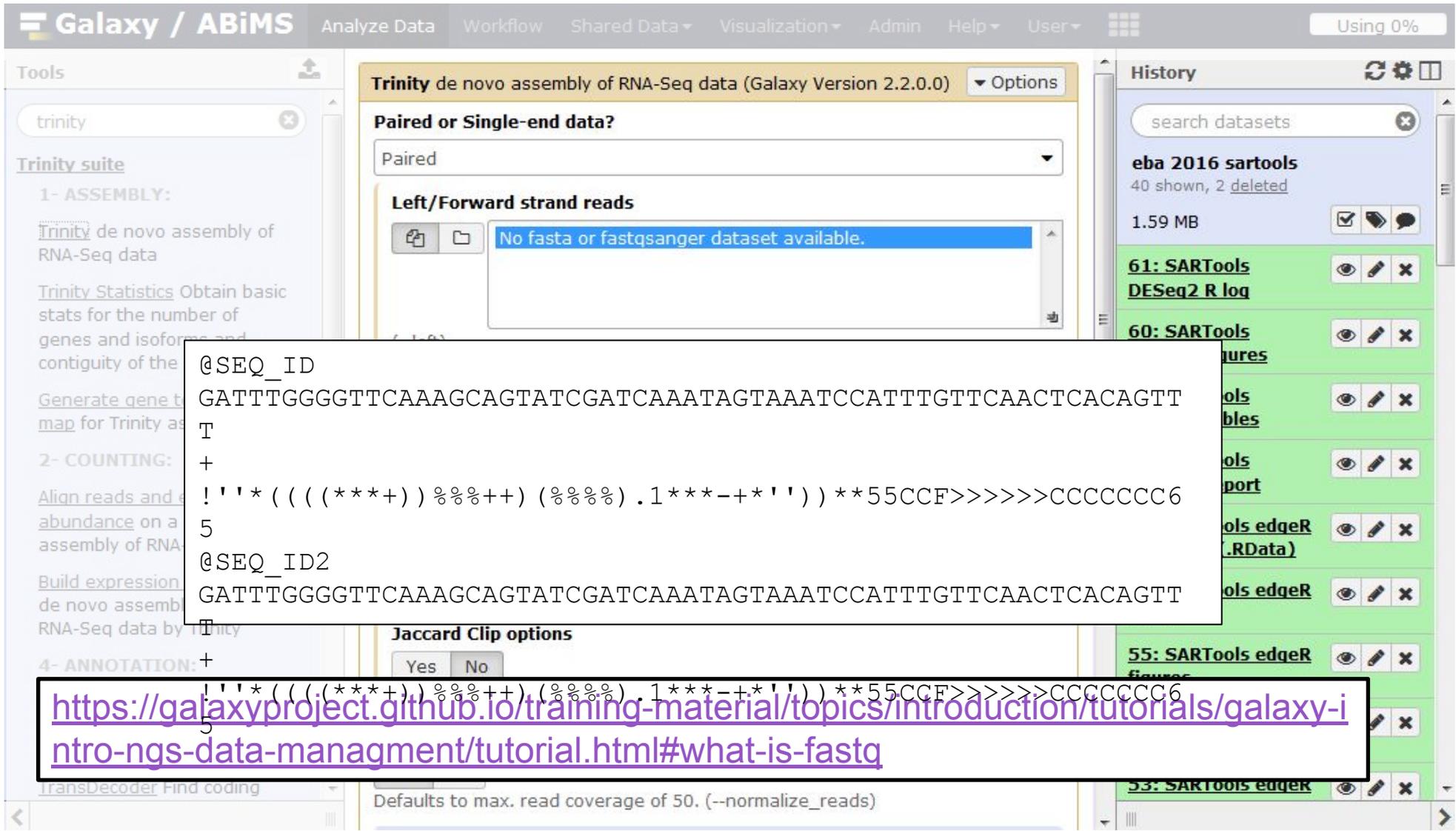
55: SARTools edgeR figures

54: SARTools edgeR tables

53: SARTools edgeR

Dataset - Datatypes

- Many tools will only accept input datasets with the appropriate datatype assigned.



Galaxy / ABiMS Analyze Data Workflow Shared Data Visualization Admin Help User Using 0%

Tools

trinity

Trinity suite

1- ASSEMBLY:

Trinity de novo assembly of RNA-Seq data

Trinity Statistics Obtain basic stats for the number of genes and isoforms and contiguity of the

Generate gene t map for Trinity as

2- COUNTING:

Align reads and abundance on a assembly of RNA

Build expression de novo assem RNA-Seq data by Trinity

4- ANNOTATION:

TransDecoder Find coding

Trinity de novo assembly of RNA-Seq data (Galaxy Version 2.2.0.0) Options

Paired or Single-end data?

Paired

Left/Forward strand reads

No fasta or fastqsanger dataset available.

Jaccard Clip options

Yes No

History

search datasets

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40 shown, 2 deleted
1.59 MB

61: SARTools
DESeq2 R log

60: SARTools

55: SARTools edgeR

53: SARTools edgeR

```

@SEQ_ID
GATTTGGGGTTCAAAGCAGTATCGATCAAATAGTAAATCCATTTGTTCAACTCACAGTT
T
+
!''*((( (***) )%%%++) (%%%) .1***-+*'' ) **55CCF>>>>>CCCCCCC6
5
@SEQ_ID2
GATTTGGGGTTCAAAGCAGTATCGATCAAATAGTAAATCCATTTGTTCAACTCACAGTT
T
    
```

<https://galaxyproject.github.io/training-material/topics/introduction/tutorials/galaxy-intro-ngs-data-managment/tutorial.html#what-is-fastq>

Defaults to max. read coverage of 50. (--normalize_reads)

Hands-on

TOOLS (sequence files manipulation)





Part 1:

You have sequencing data from your favorite species. You want to check the quality of your sequences and to map on the reference genome.

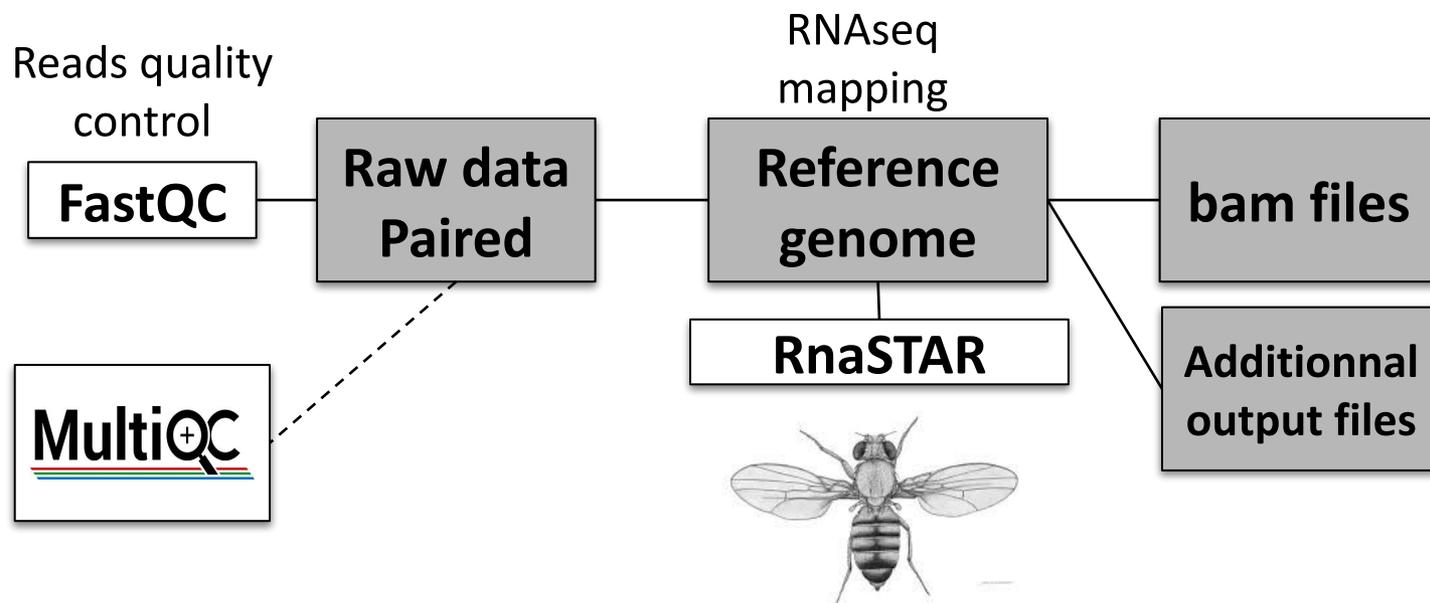
Case of *Drosophila melanogaster* (dm)



Part 1:

You have sequencing data from your favorite species. You want to check the quality of your sequences and to map on the reference genome.

Case of *Drosophila melanogaster* (dm)





Part 1:

You have sequencing data from your favorite species. You want to check the quality of your sequences and to map on the reference genome.

Case of *Drosophila melanogaster* (dm)

1. Look into the description of the tool FastQC, what is its purpose ?
2. Does it takes compressed files ? ".gz"
3. Which encoding of the file (in FastQC results, basic statistics) ? => edit if needed the datatype of your reads files
4. Look at mapping tools, map your reads on genome with RNA STAR.
5. What is the default parameter of the option "Maximum ratio of mismatches to mapped length" in RNA STAR ?
6. How to map on another reference genome or assembly release ?



Part 2:

You have new sequences that you want to compare with the gene and protein databases from your favorite species (BLAST).

Case of *Drosophila melanogaster* (dm)

1. Create new history
2. Import CDS and peptide sequences databases
3. Import query sequences
4. Make BLAST databases
5. Run BLAST against the CDS database
6. Run BLAST against the protein database



Part 2:

You have new sequences that you want to compare with the gene and protein databases from your favorite species (BLAST).

Case of *Drosophila melanogaster* (dm)

1. Create new history
2. Import CDS and peptide sequences databases
3. Import query sequences
4. Make BLAST databases *Blast --> NCBI BLAST+ makeblastdb*
5. Run BLAST against the CDS database
6. Run BLAST against the protein database



Part 2:

You have new sequences that you want to compare with the gene and protein databases from your favorite species (BLAST).

Case of *Drosophila melanogaster* (dm)

1. Create new history
2. Import CDS and peptide sequences databases
3. Import query sequences
4. Make BLAST databases *Blast --> NCBI BLAST+ makeblastdb*
5. Run BLAST against the CDS database *Blast --> NCBI BLAST+ blastn*
6. Run BLAST against the protein database



Part 2:

You have new sequences that you want to compare with the gene and protein databases from your favorite species (BLAST).

Case of *Drosophila melanogaster* (dm)

1. Create new history
2. Import CDS and peptide sequences databases
3. Import query sequences
4. Make BLAST databases *Blast --> NCBI BLAST+ makeblastdb*
5. Run BLAST against the CDS database *Blast --> NCBI BLAST+ blastn*
6. Run BLAST against the protein database *Blast --> NCBI BLAST+ blastx*

DATASET COLLECTION



- Problematic: you have a large numbers of datasets to send through the same analysis

- Problematic: you have a large numbers of datasets to send through the same analysis
- Solution 1: select multiple datasets as input

Dataset collection

Select multiple datasets as input

Galaxy Analyze Data Workflow Shared Data Visualization Help User Using 0%

Workflow: Find exons with highest number of SNPs

Run workflow

History Options

Send results to a new history

Yes No

1: Input dataset

2: Exons

2: Input dataset

4: Repeats
3: SNPs
2: Exons

Multiple datasets

This is a batch mode input field. Separate jobs will be triggered for each dataset selection.

Batch options:

3: Join the intervals of two datasets side-by-side (Galaxy Version 1.0.0)

4: Group data by a column and perform aggregate operation on other columns. (Galaxy Version 2.1.1)

History

search datasets

Galaxy initiation - multiple datasets

3 shown, 1 deleted

10.41 MB

4: Repeats

3: SNPs

2: Exons

Dataset collection

Select multiple datasets as input

The screenshot displays the Galaxy web interface. At the top, the navigation bar includes 'Galaxy', 'Analyze Data', 'Workflow', 'Shared Data', 'Visualization', 'Help', and 'User'. A 'Using 0%' indicator is visible in the top right. On the left, a 'Tools' sidebar lists various categories such as 'Get Data', 'Send Data', 'Text Manipulation', and 'NGS: DeepTools'. The main workspace contains a green notification box with a checkmark icon, stating: 'Successfully invoked workflow **Find exons with highest number of SNPs 2 times.** You can check the status of queued jobs and view the resulting data by refreshing the History pane. When the job has been run the status will change from 'running' to 'finished' if completed successfully or 'error' if problems were encountered.' On the right, the 'History' pane shows a search bar and a list of datasets. The top entry is 'Galaxy initiation - multiple datasets' (7 shown, 1 deleted, 6 hidden, 11.68 MB). Below it, several datasets are listed, each with a green background and a red border: '14: Top exon genetic location', '13: Top exons', '12: Top exon genetic location', '10: Top exons', '4: Repeats', '3: SNPs', and '2: Exons'. Each dataset entry includes an eye icon, a pencil icon, and a close icon.

Galaxy

Analyze Data Workflow Shared Data Visualization Help User

Using 0%

Tools

search tools

Get Data

Send Data

Lift-Over

Text Manipulation

Datamash

Convert Formats

Filter and Sort

Join, Subtract and Group

Fetch Alignments/Sequences

NGS: QC and manipulation

NGS: DeepTools

NGS: Mapping

NGS: RNA Analysis

NGS: SAMtools

NGS: BamTools

NGS: Picard

NGS: VCF Manipulation

NGS: Peak Calling

NGS: Variant Analysis

NGS: RNA Structure

NGS: Du Novo

NGS: Gemini

History

search datasets

Galaxy initiation - multiple datasets

7 shown, 1 deleted, 6 hidden

11.68 MB

14: Top exon genetic location

13: Top exons

12: Top exon genetic location

10: Top exons

4: Repeats

3: SNPs

2: Exons

- Problematic: you have a large numbers of datasets to send through the same analysis
- Solution 1: select multiple datasets as input
- Solution 2: create a dataset collection

- Problematic: you have a large numbers of datasets to send through the same analysis
- Solution 1: select multiple datasets as input
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 - Dataset list: set of files of the same type
 - Dataset pairs: pairs of read files (forward, reverse)
 - List of dataset pairs

- Problematic: you have a large numbers of datasets to send through the same analysis
- Solution 1: select multiple datasets as input
- Solution 2: create a dataset collection
 - Dataset list: set of files of the same type
 - Dataset pairs: pairs of read files (forward, reverse)
 - List of dataset pairs
- Galaxy runs the tool automatically on each dataset in the collection using the same settings

Dataset collection

Create a dataset collection

The screenshot displays the Galaxy / ABiMS web interface. The top navigation bar includes 'Galaxy / ABiMS', 'Analyze Data', 'Workflow', 'Shared Data', 'Visualization', 'Admin', 'Help', 'User', and a 'Using 0%' indicator. The left sidebar lists various tool categories such as 'Get Data', 'Send Data', 'Collection Operations', and 'COMMON TOOLS'. The main content area features a green welcome message for 'galaxy3.sb-roscoff.fr', an information box with contact details, and the ABiMS logo (Analyses and Bioinformatics for Marine Science) with links to 'Changelog' and 'Tutorials'. The right sidebar shows a 'History' panel with a search bar and a list of datasets under the heading 'Galaxy initiation - collection'. The list includes '3: Repeats', '2: SNPs', and '1: Exons'. A red box highlights a checkmark icon next to the '3: Repeats' entry, with a tooltip that reads 'Operations on multiple datasets'.

Dataset collection

Create a dataset collection

The screenshot displays the Galaxy / ABiMS web interface. The main content area features a green welcome banner for 'galaxy3.sb-roscoff.fr' and an information box with contact details for support.abims@sb-roscoff.fr. Below this is the ABiMS logo and navigation links for Changelog and Tutorials. A paragraph of text describes Galaxy as an open, web-based platform for data-intensive biomedical research, supported by various institutions.

The left sidebar contains a 'Tools' section with a search bar and a list of tool categories: Get Data, Send Data, Collection Operations, COMMON TOOLS, Text Manipulation, Filter and Sort, Join, Subtract and Group, Convert Formats, Extract Features, Fetch Sequences, Statistics, Graph/Display Data, Fasta Fastq Manipulation, COMMON NGS TOOLS, NGS:Samtools, NGS:Mapping, NGS:Bedtools, NGS:Picard Tools, and SEARCHING TOOLS with Diamond.

The right sidebar shows the 'History' section with a search bar and a list of datasets. A context menu is open over the 'Galaxy initiation - collection' dataset, listing actions such as Hide datasets, Unhide datasets, Delete datasets, Undelete datasets, Permanently delete datasets, Build Dataset List, Build Dataset Pair, and Build List of Dataset Pairs. The 'Build Dataset List' option is highlighted with a red box.

Dataset collection

Create a dataset collection

The screenshot shows the Galaxy ABiMS web interface. At the top, there is a navigation bar with the Galaxy logo and menu items: Analyze Data, Workflow, Shared Data, Visualization, Admin, Help, and User. A 'Using 0%' indicator is visible in the top right. On the left side, there is a 'Tools' sidebar with a search bar and various tool categories like 'Get Data', 'Send Data', 'Collection Operations', 'COMMON TOOLS', 'Text Manipulation', 'Filter and Sort', 'Join, Subtract and Merge', 'Convert Formats', 'Extract Features', 'Fetch Sequences', 'Statistics', 'Graph/Display', 'Fasta Fastq Manipulation', 'COMMON NGS TOOLS', 'NGS:Samtools', 'NGS:Mapping', 'NGS:Bedtools', 'NGS:Picard Tools', 'SEARCHING TOOLS', and 'Diamond'. The main content area is partially obscured by a dialog box titled 'Create a collection from a list of datasets'. The dialog box contains the following elements: a title bar, a text area with the text 'Collections of datasets are permanent, ordered lists of datasets that can be passed to tools and workfl... More help', a 'Start over' link, two rows of dataset selection buttons labeled 'Repeats' and 'SNPs', each with a 'Discard' button to its right, a 'Name' label followed by a text input field containing 'Collection of different features', a 'Cancel' button, and a 'Create list' button. The 'Create list' button is highlighted with a red border. In the background, a 'Tutorials' section is visible, containing a paragraph of text about the Galaxy project and its support.

Galaxy / ABiMS Analyze Data Workflow Shared Data Visualization Admin Help User Using 0%

Create a collection from a list of datasets

Collections of datasets are permanent, ordered lists of datasets that can be passed to tools and workfl... [More help](#)

[Start over](#)

[Repeats](#) [Discard](#)

[SNPs](#) [Discard](#)

Name

[Cancel](#) [Create list](#)

Tutorials

Galaxy is an open, web-based platform for data intensive biomedical research. The Galaxy team is a part of BX at Penn State, and the Biology and Mathematics and Computer Science departments at Emory University. The Galaxy Project is supported in part by NHGRI, NSF, The Huck Institutes of the Life Sciences, The Institute for CyberScience at Penn State, and Emory University.

Dataset collection

Create a dataset collection

The screenshot displays the Galaxy / ABiMS web interface. The top navigation bar includes 'Galaxy / ABiMS', 'Analyze Data', 'Workflow', 'Shared Data', 'Visualization', 'Admin', 'Help', 'User', and a 'Using 0%' indicator. The left sidebar lists various tool categories such as 'Get Data', 'Send Data', 'Collection Operations', 'COMMON TOOLS', 'Text Manipulation', 'Filter and Sort', 'Join, Subtract and Group', 'Convert Formats', 'Extract Features', 'Fetch Sequences', 'Statistics', 'Graph/Display Data', 'Fasta Fastq Manipulation', 'COMMON NGS TOOLS', 'NGS:Samtools', 'NGS:Mapping', 'NGS:Bedtools', 'NGS:Picard Tools', 'SEARCHING TOOLS', and 'Diamond'. The main content area features a green welcome banner for 'galaxy3.sb-roscoff.fr', an information box with contact details for support.abims@sb-roscoff.fr, and the ABiMS logo (Analyses and Bioinformatics for Marine Science) with logos for CNRS UPR1098 and Station Biologique Roscoff. Below the logo are links for 'Changelog' and 'Tutorials'. A paragraph of text describes Galaxy as an open, web-based platform for data intensive biomedical research, supported by NHGRI, NSF, The Huck Institutes of the Life Sciences, The Institute for CyberScience at Penn State, and Emory University. The right sidebar shows a 'History' section with a search bar and a list of dataset collections. The first collection, '4: Collection of different features', is highlighted with a red border and includes a sub-item 'a list of datasets'. Other collections listed are '3: Repeats', '2: SNPs', and '1: Exons'. The total size of the collections is 11.32 MB.

Galaxy / ABiMS Analyze Data Workflow Shared Data Visualization Admin Help User Using 0%

Tools search tools

Get Data Send Data Collection Operations

COMMON TOOLS Text Manipulation Filter and Sort Join, Subtract and Group Convert Formats Extract Features Fetch Sequences Statistics Graph/Display Data Fasta Fastq Manipulation

COMMON NGS TOOLS NGS:Samtools NGS:Mapping NGS:Bedtools NGS:Picard Tools

SEARCHING TOOLS Diamond

Welcome to galaxy3.sb-roscoff.fr

Information For any question or request for tools or account, send an email at support.abims@sb-roscoff.fr

ABiMS Analyses and Bioinformatics for Marine Science

Station Biologique Roscoff

Changelog Tutorials

Galaxy is an open, web-based platform for data intensive biomedical research. The Galaxy team is a part of BX at Penn State, and the Biology and Mathematics and Computer Science departments at Emory University. The Galaxy Project is supported in part by NHGRI, NSF, The Huck Institutes of the Life Sciences, The Institute for CyberScience at Penn State, and Emory University.

History search datasets

Galaxy initiation - collection 4 shown

11.32 MB

All None For all selected...

4: Collection of different features
a list of datasets

3: Repeats

2: SNPs

1: Exons

Dataset collection

Create a dataset collection

The screenshot displays the Galaxy / ABiMS web interface. The top navigation bar includes 'Galaxy / ABiMS', 'Analyze Data', 'Workflow', 'Shared Data', 'Visualization', 'Admin', 'Help', 'User', and a 'Using 0%' indicator. The left sidebar lists various tool categories such as 'Get Data', 'Send Data', 'Collection Operations', 'COMMON TOOLS', 'Text Manipulation', 'Filter and Sort', 'Join, Subtract and Group', 'Convert Formats', 'Extract Features', 'Fetch Sequences', 'Statistics', 'Graph/Display Data', 'Fasta Fastq Manipulation', 'COMMON NGS TOOLS', 'NGS:Samtools', 'NGS:Mapping', 'NGS:Bedtools', 'NGS:Picard Tools', 'SEARCHING TOOLS', and 'Diamond'. The main content area features a green 'Welcome to galaxy3.sb-roscoff.fr' message, an 'Information' box with contact details for support.abims@sb-roscoff.fr, the ABiMS logo (Analyses and Bioinformatics for Marine Science), and links to 'Changelog' and 'Tutorials'. A paragraph of text describes Galaxy as an open, web-based platform for data-intensive biomedical research, mentioning its affiliation with Penn State and Emory University. The right sidebar shows a 'History' section with a '< Back to Galaxy initiation - collection' link and a highlighted 'Collection of different features' section, which includes 'Repeats' and 'SNPs' with eye and pencil icons.

Dataset collection

Use a collection as input

BED-to-GFF converter (Galaxy Version 2.0.0) Options

Convert this dataset

5: Collection of different features

Dataset collection

Execute

What it does

This tool converts data from BED format to GFF format (scroll down for format description).

Example

The following data in BED format:

chr28	346187	388197	BC114771	0	+	346187	388197	0
-------	--------	--------	----------	---	---	--------	--------	---

Will be converted to GFF (**note** that the start coordinate is incremented by 1):

chr28	bed2gff	mRNA	346188	388197	0	+	.	mRNA	BC1147
chr28	bed2gff	exon	346188	346331	0	+	.	exon	BC1147
chr28	bed2gff	exon	370283	370363	0	+	.	exon	BC1147
chr28	bed2gff	exon	372378	372492	0	+	.	exon	BC1147
chr28	bed2gff	exon	377194	377256	0	+	.	exon	BC1147
chr28	bed2gff	exon	378319	378473	0	+	.	exon	BC1147
chr28	bed2gff	exon	379722	379817	0	+	.	exon	BC1147
chr28	bed2gff	exon	383182	383315	0	+	.	exon	BC1147
chr28	bed2gff	exon	387981	388085	0	+	.	exon	BC1147

History

search datasets

Galaxy initiation - collection
4 shown, 5 deleted, 6 hidden

11.68 MB

5: Collection of different features
a list of 2 datasets

4: Repeats

2: SNPs

1: Exons

Dataset collection

Use a collection as input

The screenshot displays the Galaxy web interface. At the top, the navigation bar includes 'Galaxy', 'Analyze Data', 'Workflow', 'Shared Data', 'Visualization', 'Help', and 'User'. A 'Using 0%' indicator is visible in the top right. On the left, a 'tools' sidebar contains a search bar and various tool categories like 'Get Data', 'Send Data', 'Lift-Over', 'Text Manipulation', 'Datamash', and 'Convert Formats'. The main workspace features a green notification box with a checkmark icon, stating: '2 jobs have been successfully added to the queue - resulting in the following datasets: 16: BED-to-GFF on data 4, 17: BED-to-GFF on data 2'. Below this, it provides instructions on checking job status. On the right, the 'History' panel shows a search bar and a list of datasets. The entry '18: BED-to-GFF on collection 5' is highlighted with a red box. Other visible entries include '17: BED-to-GFF on data 2', '16: BED-to-GFF on data 4', '5: Collection of different features', '4: Repeats', '2: SNPs', and '1: Exons'. Each entry includes icons for refresh, edit, and delete.

Hands-on
COLLECTION





Part 2:

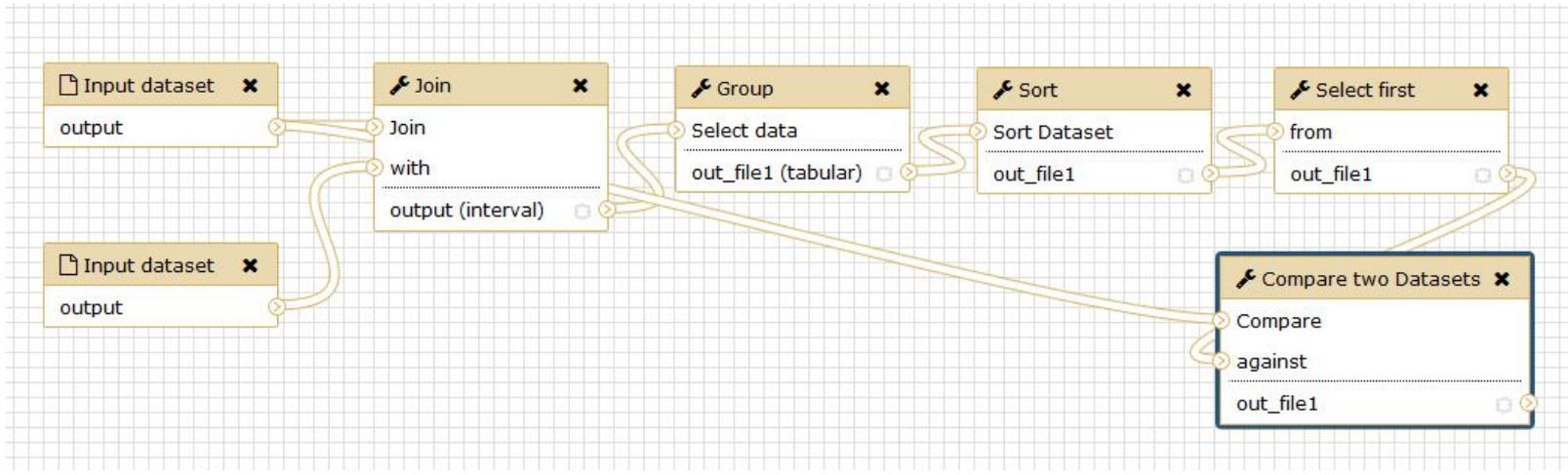
You have new sequences that you want to compare with the gene and protein databases from your favorite species (BLAST).

Case of *Drosophila melanogaster* (dm)

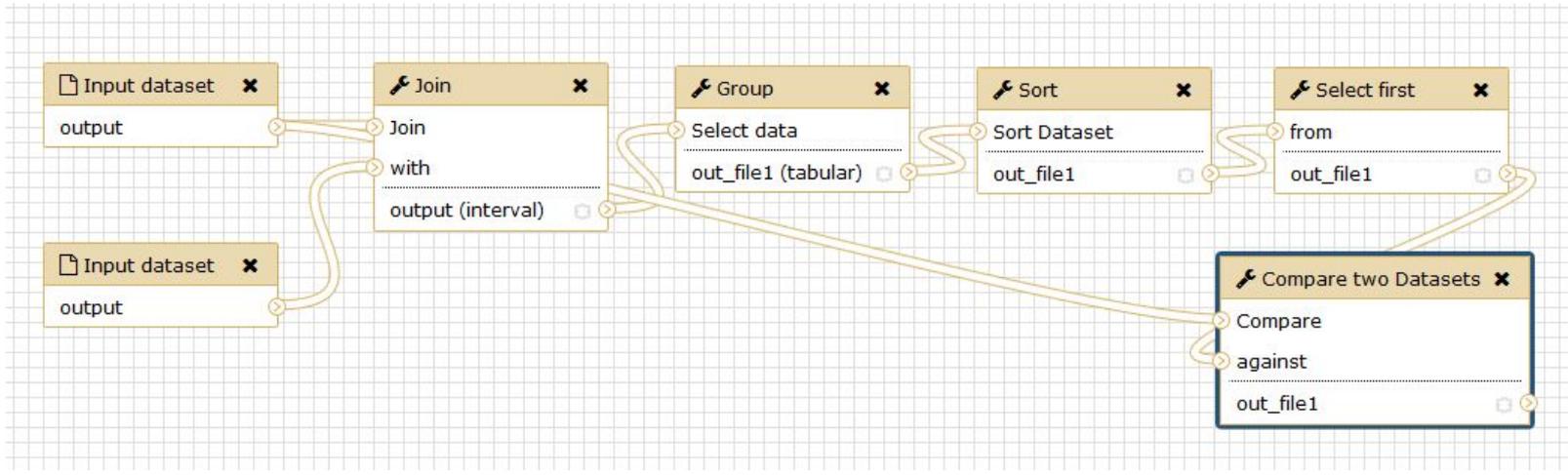
1. Create new history
2. Import CDS and peptide sequences databases
3. Import query sequences
4. Make BLAST databases
5. Run BLAST against the CDS database
6. Run BLAST against the protein database
7. Create a dataset list with all the query sequences
8. Run BLAST against the CDS database on the dataset list

WORKFLOW

Workflow



- A workflow is a sequence of tool operations and parameters
- A workflow is built to be replayed (more or less strict)

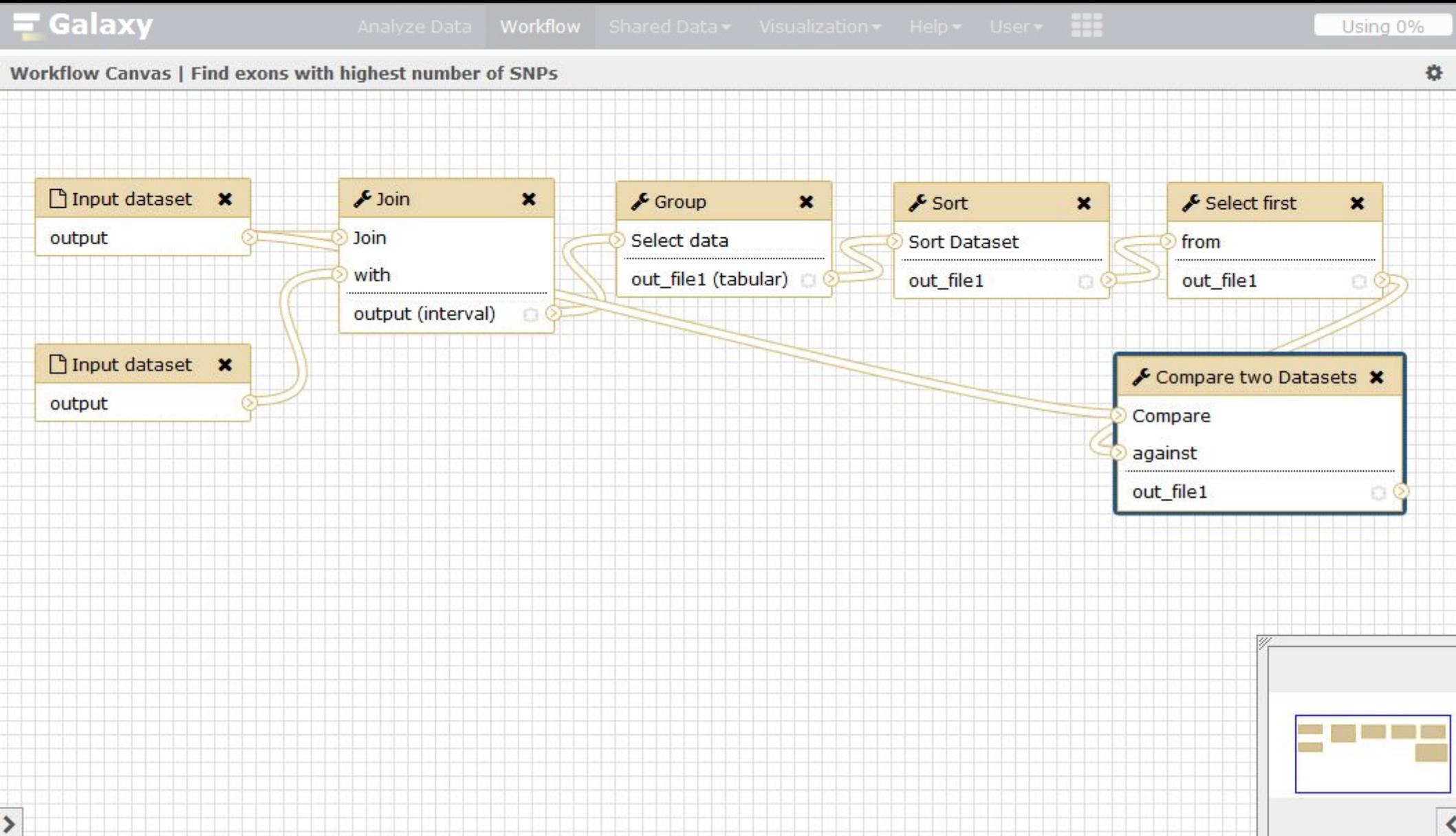


Why would you want to create workflows?

- Re-run the same analysis on different input data sets
- Change parameters before re-running a similar analysis
- Make use of the workflow job scheduling (jobs are submitted as soon as their inputs are ready)
- Share workflows for publication and with the community

Workflow

Our workflow with Galaxy



Workflow

From history

The following list contains each tool that was run to create the datasets in your current history. Please select those that you wish to include in the workflow.

Tools which cannot be run interactively and thus cannot be incorporated into a workflow will be shown in gray.

Workflow name

Workflow constructed from history 'Galaxy initiation'

Create Workflow Check all Uncheck all

Tool	History items created
UCSC Main <i>This tool cannot be used in workflows</i>	1 Exons <input checked="" type="checkbox"/> Treat as input dataset Exons
UCSC Main <i>This tool cannot be used in workflows</i>	2 SNPs <input checked="" type="checkbox"/> Treat as input dataset SNPs
Join <input checked="" type="checkbox"/> Include "Join" in workflow	3 Join on data 2 and data 1
Group <input checked="" type="checkbox"/> Include "Group" in workflow	5 Group on data 3
Sort <input checked="" type="checkbox"/> Include "Sort" in workflow	6 Sort on data 5

https://usegalaxy.org/workflow/build_from_current_history

History

- HISTORY LISTS
- Saved Histories
- Histories Shared with Me
- CURRENT HISTORY
- Create New
- Copy History
- Share or Publish
- Show Structure
- Extract Workflow**
- Delete
- Delete Permanently
- DATASET ACTIONS
- Copy Datasets
- Dataset Security
- Resume Paused Jobs
- Collapse Expanded Datasets
- Unhide Hidden Datasets
- Delete Hidden Datasets
- Purge Deleted Datasets
- DOWNLOADS
- Export Tool Citations
- Export History to File
- OTHER ACTIONS
- Import from File

Workflow

From history

Galaxy Analyze Data Workflow Shared Data Visualization Help User Using 0%

The following list contains each tool that was run to create the datasets in your current history. Please select those that you wish to include in the workflow.

Tools which cannot be run interactively and thus cannot be incorporated into a workflow will be shown in gray.

Workflow name
Find exons with the highest SNPs

Create Workflow Check all Uncheck all

Tool	History items created
UCSC Main <i>This tool cannot be used in workflows</i>	1 Exons <input checked="" type="checkbox"/> Treat as input dataset Exons
UCSC Main <i>This tool cannot be used in workflows</i>	2 SNPs <input checked="" type="checkbox"/> Treat as input dataset SNPs
Join <input checked="" type="checkbox"/> Include "Join" in workflow	3 Join on data 2 and data 1
Group <input checked="" type="checkbox"/> Include "Group" in workflow	5 Group on data 3
Sort <input checked="" type="checkbox"/> Include "Sort" in workflow	6 Sort on data 5

History search datasets Using 0%

Galaxy initiation
7 shown, 1 deleted, 1 hidden
8.77 MB

8: Compare two Datasets on data 7 and data 1

7: Select first on data 6

6: Sort on data 5

5: Group on data 3

3: Join on data 2 and data 1

2: SNPs

1: Exons

Workflow

From history

The following list contains each tool that was run to create the datasets in your current history. Please select those that you wish to include in the workflow.

Tools which cannot be run interactively and thus cannot be incorporated into a workflow will be shown in gray.

Workflow name
Find exons with the highest SNPs

Create Workflow Check all Uncheck all

Tool	History items created
UCSC Main <i>This tool cannot be used in workflows</i>	1 Exons <input checked="" type="checkbox"/> Treat as input dataset Exons
UCSC Main <i>This tool cannot be used in workflows</i>	2 SNPs <input checked="" type="checkbox"/> Treat as input dataset SNPs
Join <input checked="" type="checkbox"/> Include "Join" in workflow	3 Join on data 2 and data 1
Group <input checked="" type="checkbox"/> Include "Group" in workflow	5 Group on data 3
Sort <input checked="" type="checkbox"/> Include "Sort" in workflow	6 Sort on data 5

History
search datasets

Galaxy initiation
7 shown, 1 deleted, 1 hidden
8.77 MB

- 8: Compare two Datasets on data 7 and data 1
- 7: Select first on data 6
- 6: Sort on data 5
- 5: Group on data 3
- 3: Join on data 2 and data 1
- 2: SNPs
- 1: Exons

Workflow

Workflow manager



Your workflows

Create new workflow

Upload or import workflow

Name	# of Steps
Find exons with highest number of SNPs	7
Convert to tab (imported from API)	2
imported: ChIP-seq workflow	3

Workflows shared with you by others

No workflows have been shared with you.

Other options

Configure your workflow menu

Workflow

Workflow manager

Galaxy Analyze Data **Workflow** Shared Data Visualization Help User Using 0%

Your workflows

[Create new workflow](#) [Upload or import workflow](#)

Name	# of Steps
Find exons with highest number of SNPs ▼	7
Convert to tab (imported from ...)	2
imported: ChIP-seq workflow	3

Workflows shared by others
No workflows have been shared by others.

Other options
[Configure your workflow menu](#)

- Edit
- Run
- Share or Download
- Copy
- Rename
- View
- Delete

<https://usegalaxy.org/workflow/editor?id=17b7895387cc2214>

Workflow

Edit a workflow: add tags and annotation

The screenshot displays the Galaxy workflow editor interface. The top navigation bar includes 'Galaxy' and menu items: 'Analyze Data', 'Workflow', 'Shared Data', 'Visualization', 'Help', and 'User'. A 'Using 0%' indicator is visible in the top right. The main area is divided into three sections: 'Tools', 'Workflow Canvas', and 'Details'.

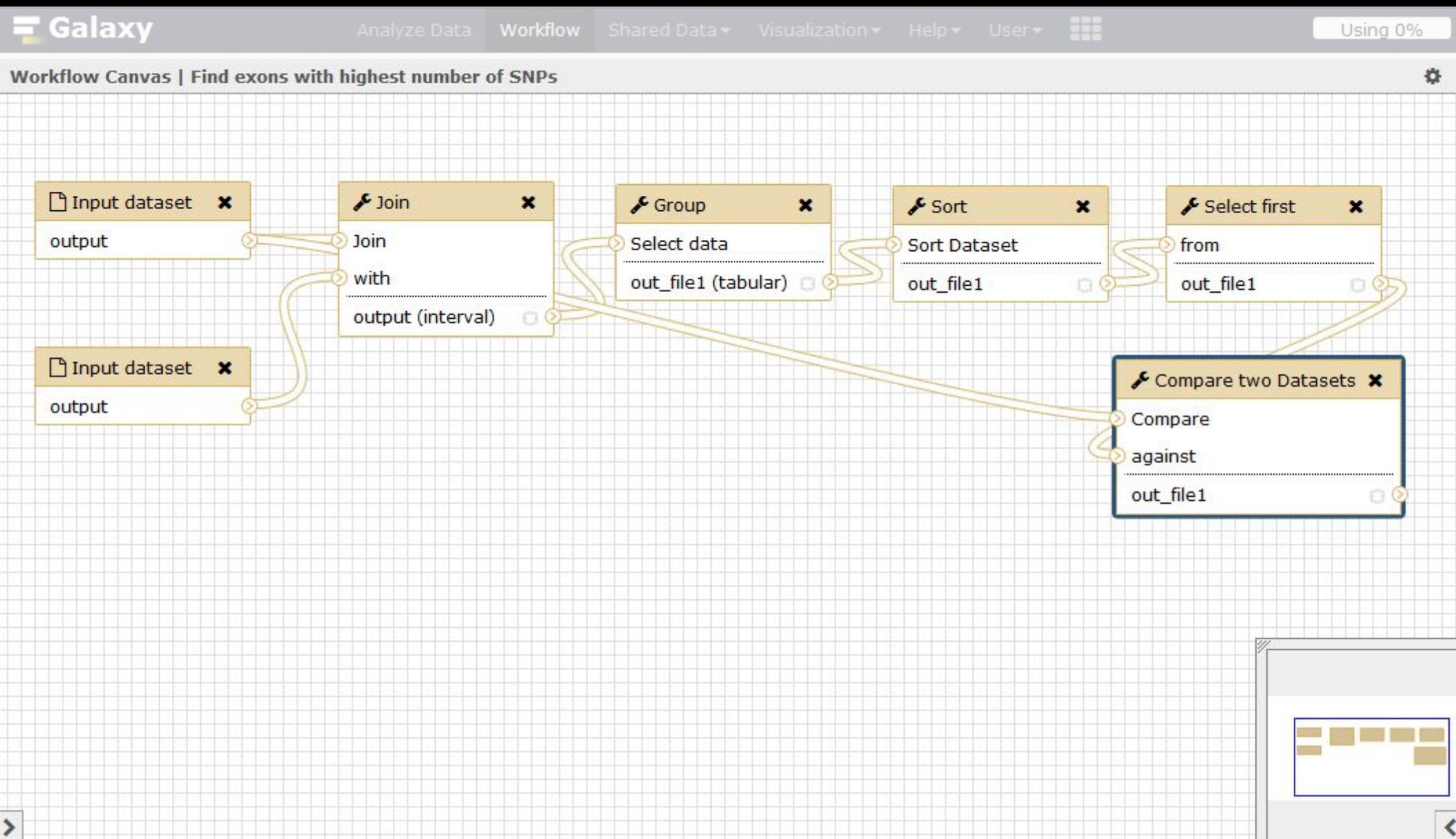
Tools: A search bar labeled 'search tools' is at the top. Below it, a list of tool categories is provided: Inputs, Get Data, Send Data, Lift-Over, Text Manipulation, Datamash, Convert Formats, Filter and Sort, Join, Subtract and Group, Fetch Alignments/Sequences, NGS: QC and manipulation, NGS: DeepTools, NGS: Mapping, NGS: RNA Analysis, NGS: SAMtools, NGS: BamTools, NGS: Picard, NGS: VCF Manipulation, NGS: Peak Calling, NGS: Variant Analysis, NGS: RNA Structure, and NGS: Du Novo.

Workflow Canvas: The title is 'Workflow Canvas | Find exons with highest number of SNPs'. It features a grid background with three main tool nodes: two 'Input dataset' nodes (each with an 'output' field) and one 'Join' node. The 'Join' node has a 'with' field containing 'output (interval)'. The 'Join' node's output is connected to a 'Group' node, which has a 'Select data' field containing 'out_file1 (tab)'. A small preview window at the bottom right shows a grid of colored rectangles.

Details: The title is 'Edit Workflow Attributes'. It contains three sections: **Name:** 'Find exons with highest number of SNPs'; **Tags:** A field with a plus icon and a dropdown menu showing 'out_file1 (tab)'; and **Annotation / Notes:** A text area with the instruction: 'Describe or add notes to workflow. Add an annotation or notes to a workflow; annotations are available when a workflow is viewed.'

Workflow

Edit a workflow



Workflow

Edit a workflow: drag and drop

The screenshot displays the Galaxy Workflow Canvas interface. At the top, the navigation bar includes the Galaxy logo, menu items for 'Analyze Data', 'Workflow', 'Shared Data', 'Visualization', 'Help', and 'User', and a 'Using 0%' indicator. The main title of the canvas is 'Workflow Canvas | Find exons with highest number of SNPs'. The workflow consists of the following steps:

- Input dataset**: Two separate input boxes, each labeled 'Input dataset' with a close button (x) and an 'output' field.
- Join**: A 'Join' tool box with a close button (x). It contains the text 'Join with output (interval)'. It receives two inputs from the 'Input dataset' boxes.
- Group**: A 'Group' tool box with a close button (x). It contains the text 'Select data out_file1 (tabular)'. It receives one input from the 'Join' tool.
- Sort**: A 'Sort' tool box with a close button (x). It contains the text 'Sort Dataset out_file1'. It receives one input from the 'Group' tool.
- Select first**: A 'Select first' tool box with a close button (x). It contains the text 'from out_file1'. It receives one input from the 'Sort' tool.
- Compare two Datasets**: A 'Compare two Datasets' tool box with a close button (x). It contains the text 'Compare against out_file1'. It receives two inputs: one from the 'Input dataset' boxes and one from the 'Select first' tool.

The workflow is visualized as a series of connected boxes on a grid background. The 'Compare two Datasets' tool is highlighted with a blue border. A small preview window in the bottom right corner shows a simplified version of the workflow diagram.

Workflow

Edit a workflow: delete a noodle

The screenshot shows the Galaxy Workflow Canvas interface. The title bar reads "Workflow Canvas | Find exons with highest number of SNPs". The main workspace contains a workflow diagram with the following tools and connections:

- Two "Input dataset" tools (output) are connected to the "Join" tool.
- The "Join" tool (with "output (interval)" selected) is connected to the "Compare two Datasets" tool.
- The "Compare two Datasets" tool (with "Compare" selected) is connected to the "Group" tool.
- The "Group" tool (with "out_file1 (tabular)" selected) is connected to the "Sort" tool.
- The "Sort" tool (with "out_file1" selected) is connected to the "Select first" tool.
- The "Select first" tool (with "out_file1" selected) is connected to the "Compare two Datasets" tool.

The "Compare two Datasets" tool is highlighted with a red box, and its "Compare" option is selected. The workflow is connected by yellow lines representing data flow.

Workflow

Edit a workflow: delete a noodle

Galaxy Analyze Data Workflow Shared Data Visualization Help User Using 0%

Workflow Canvas | Find exons with highest number of SNPs

```
graph LR; I1[Input dataset] --> J[Join]; I2[Input dataset] --> J; J --> G[Group]; G --> S[Sort]; S --> SF[Select first]; SF --> CD[Compare two Datasets];
```

The workflow consists of the following steps:

- Input dataset (output)
- Input dataset (output)
- Join (Join with output (interval))
- Group (Select data out_file1 (tabular))
- Sort (Sort Dataset out_file1)
- Select first (from out_file1)
- Compare two Datasets (Compare against out_file1)

Workflow

Edit a workflow: add a tool

The screenshot displays the Galaxy workflow editor interface. The top navigation bar includes 'Galaxy', 'Analyze Data', 'Workflow', 'Shared Data', 'Visualization', 'Help', and 'User'. A 'Using 0%' indicator is visible in the top right. The main area is titled 'Workflow Canvas | Find exons with highest number of SNPs'. The workflow consists of several tools connected by lines: two 'Input dataset' tools feeding into a 'Join' tool, which then feeds into a 'Group' tool, followed by a 'Sort' tool. A 'Merge Columns' tool is highlighted with a red box and is currently being added to the canvas. Below it, a 'Compare two Datasets' tool is also visible. The left sidebar contains a 'Tools' panel with a search bar and a list of tool categories: 'Get Data', 'Send Data', 'Lift-Over', and 'Text Manipulation'. The 'Merge Columns together' tool is highlighted in red in the 'Text Manipulation' section.

Galaxy Analyze Data Workflow Shared Data Visualization Help User Using 0%

Tools Workflow Canvas | Find exons with highest number of SNPs

search tools

Get Data

Send Data

Lift-Over

Text Manipulation

- UniProt ID mapping and retrieval
- Compute an expression on every row
- Concatenate datasets tail-to-head
- Add column to an existing dataset
- Concatenate datasets tail-to-head (cat)
- tac reverse a file (reverse cat)
- Condense consecutive characters
- Cut columns from a table
- Convert delimiters to TAB
- Merge Columns together**
- Remove beginning of a file

Input dataset x
output

Join x
Join
with
output (interval)

Group x
Select data
out_file1 (tabular)

Sort x
Sort Dataset
out_file1

Merge Columns x
Select data
out_file1 (tabular)

Compare two Datasets x
Compare
against
out_file1

Workflow

Edit a workflow: add a noodle

Galaxy Analyze Data Workflow Shared Data Visualization Help User Using 0%

Tools

search tools

Get Data

Send Data

Lift-Over

Text Manipulation

- UniProt ID mapping and retrieval
- Compute an expression on every row
- Concatenate datasets tail-to-head
- Add column to an existing dataset
- Concatenate datasets tail-to-head (cat)
- tac reverse a file (reverse cat)
- Condense consecutive characters
- Cut columns from a table
- Convert delimiters to TAB
- Merge Columns together
- Remove beginning of a file

Workflow Canvas | Find exons with highest number of SNPs

The workflow canvas displays the following tools and connections:

- Input dataset** (output) - connected to **Join** (with output (interval))
- Input dataset** (output) - connected to **Join** (with output (interval))
- Join** (with output (interval)) - connected to **Merge Columns** (Select data out_file1 (tabular))
- Merge Columns** (Select data out_file1 (tabular)) - connected to **Group** (Select data out_file1 (tabular))
- Group** (Select data out_file1 (tabular)) - connected to **Sort** (Sort Dataset out_file1)

A red box highlights the output of the first **Input dataset** tool, and a red arrow points to the **Merge Columns** tool, indicating the addition of a new 'noodle' (connection).

Workflow

Edit a workflow: hide intermediate steps

The screenshot shows the Galaxy Workflow Canvas interface. The title bar reads "Workflow Canvas | Find exons with highest number of SNPs". The workflow consists of the following steps:

- Two "Input dataset" steps, each with an "output" field.
- A "Join" step with "Join with" and "output (interval)" fields.
- A "Group" step with "Select data" and "out_file1 (tabular)" fields.
- A "Sort" step with "Sort Dataset" and "out_file1" fields.
- A "Select first" step with "from" and "out_file1" fields.
- A "Compare" step with "Compare against" and "out_file1" fields.

Two red boxes highlight the "Mark dataset as a workflow output" icon (a gear with a plus sign) in the "Select first" and "Compare" steps. A tooltip is visible over the "Compare" step, containing the text: "Mark dataset as a workflow output. All unmarked datasets will be hidden."

Workflow

Edit a workflow: set or release a parameter

The screenshot displays the Galaxy Workflow Canvas interface. The workflow is titled "Find exons with highest number of SNPs" and consists of four tools connected in a sequence:

- Group**: Select data from `out_file1 (tabular)`.
- Sort**: Sort Dataset from `out_file1`.
- Select first**: Select the first 5 lines from `out_file1`. This tool is highlighted with a red box.
- Compare two Datasets**: Compare against `out_file1`.

The configuration panel for the "Select first" tool is open on the right side of the interface. It shows the following settings:

- Select first lines from a dataset (Galaxy Version 1.0.0)**: A dropdown menu.
- Select first**: A checkbox that is checked.
- 5**: A text input field for the number of lines to select.
- lines**: A label for the input field.
- from**: A dropdown menu set to "Data input 'input' (txt)".
- Annotation / Notes**: A text area for adding notes.
- Email notification**: Radio buttons for "Yes" and "No".
- Output cleanup**: Radio buttons for "Yes" and "No".

Workflow

Edit a workflow: set or release a parameter

The screenshot displays the Galaxy Workflow Canvas for a workflow titled "Find exons with highest number of SNPs". The workflow is composed of the following steps:

- Group**: Select data (out_file1 (tabular))
- Sort**: Sort Dataset (out_file1)
- Select first**: from (out_file1) - This step is highlighted with a blue border.
- Compare two Datasets**: Compare against (out_file1)
- Select first**: from (out_file1) - This step is highlighted with a blue border.

The configuration panel for the selected "Select first" step is visible on the right, showing the following details:

- Select first lines from a dataset (Galaxy Version 1.0.0)**
- Select first** (parameter icon)
- Set at Runtime** (dropdown menu)
- lines** (parameter)
- from**: Data input 'input' (txt)
- Annotation / Notes** (text area)
- Email notification**: Yes No (radio buttons)
- Output cleanup**: Yes No (radio buttons)

Workflow

Edit a workflow: rename the outputs

The screenshot displays the Galaxy workflow editor interface. The top navigation bar includes 'Galaxy', 'Analyze Data', 'Workflow', 'Shared Data', 'Visualization', 'Help', and 'User'. The workflow canvas shows four steps: 'Group', 'Sort', 'Select first', and 'Compare two Datasets'. The 'Select first' step is highlighted with a blue border. The 'Compare two Datasets' step is also highlighted with a blue border. The 'Rename dataset' configuration panel is open, showing a text input field with the value 'Top exons'.

Workflow Canvas | Find exons with highest number of SNPs

Details
step. It will be shown with the workflow.

Email notification
 Yes No
An email notification will be sent when the job has completed.

Output cleanup
 Yes No
Upon completion of this step, delete non-starred outputs from completed workflow steps if they are no longer required as inputs.

Configure Output: 'out_file1'

Label

This will provide a short name to describe the output - this must be unique across workflows.

Rename dataset

This action will rename the output dataset. Click [here](#) for more information. Valid inputs are: **input**.

Workflow

Save

The screenshot displays the Galaxy Workflow Canvas interface. The title bar reads "Workflow Canvas | Find exons with highest number of SNPs". The workflow consists of four steps connected by yellow lines:

- Group**: Select data, out_file1 (tabular)
- Sort**: Sort Dataset, out_file1
- Select first**: from, out_file1 (highlighted with a blue border)
- Compare two Datasets**: Compare against, out_file1

A context menu is open over the "Select first" step, with the "Save" option highlighted in a blue bar. The menu options are:

- Save
- Save As
- Run
- Edit Attributes
- Auto Re-layout
- Close

The right sidebar shows the "Details" panel for the selected step, including fields for "from" (Data input 'input' (txt)), "Annotation / Notes", "Email notification" (Yes/No buttons), and "Output cleanup" (Yes/No buttons).

Workflow

Run a workflow

Galaxy Analyze Data **Workflow** Shared Data Visualization Help User Using 0%

Your workflows

Create new workflow Upload or import workflow

Name	# of Steps
Find exons with highest number of SNPs	7
Convert to tab (imported from ...)	2
imported: ChIP-seq workflow	3

Workflows shared by others
No workflows have been shared by others

Other options

Configure your workflow menu

https://usegalaxy.org/root?workflow_id=17b7895387cc2214

Workflow

Run a workflow

The screenshot displays the Galaxy workflow editor interface. The main workspace shows a workflow titled "Workflow: Find exons with highest number of SNPs" with a "Run workflow" button. The workflow steps are:

- History Options:** "Send results to a new history" with "Yes" selected.
- 1: Input dataset:** A dropdown menu showing "2: Exons" (highlighted with a red box).
- 2: Input dataset:** A dropdown menu showing "1: Repeats" (highlighted with a red box).
- 3: Join the intervals of two datasets side-by-side (Galaxy Version 1.0.0)**
- 4: Group data by a column and perform aggregate operation on other columns. (Galaxy Version 2.1.1)**
- 5: Sort data in ascending or descending order (Galaxy Version 1.0.3)**
- 6: Select first lines from a dataset (Galaxy Version 1.0.0)** (highlighted with a red box). Below this step, a "Select first" section has a text input field containing "20" and the label "lines" below it.

The left sidebar contains a "Tools" section with a search bar and a list of tool categories: Get Data, Send Data, Lift-Over, Text Manipulation, Datamash, Convert Formats, Filter and Sort, Join, Subtract and Group, Fetch Alignments/Sequences, NGS: QC and manipulation, NGS: DeepTools, NGS: Mapping, NGS: RNA Analysis, NGS: SAMtools, NGS: BamTools, NGS: Picard, NGS: VCF Manipulation, NGS: Peak Calling, NGS: Variant Analysis, NGS: RNA Structure, NGS: Du Novo, and NGS: Gemini.

The right sidebar shows the "History" section with a search bar and a list of datasets: "Galaxy initiation - workflow" (2 shown, 2.77 MB) and "2: Exons" (highlighted in green). Below it, "1: Repeats" is also highlighted in green.

Workflow

Run a workflow

Galaxy Analyze Data Workflow Shared Data Visualization Help User Using 0%

Tools search tools

Workflow: Find exons with highest number of SNPs Run workflow

History Options

Send results to a new history
Yes No

1: Input dataset
2: Exons

2: Input dataset
1: Repeats

3: Join the intervals of two datasets side-by-side (Galaxy Version 1.0.0)

Join
Output dataset 'output' from step 1
with
Output dataset 'output' from step 2
 with min overlap
Edit

Return
Only records that are joined (INNER JOIN)

History search datasets

Galaxy initiation - workflow
2 shown, 2 deleted, 3 hidden

2.92 MB

2: Exons

1: Repeats

Workflow

Run a workflow

Galaxy Analyze Data Workflow Shared Data Visualization Help User Using 0%

Workflow: Find exons with highest number of SNPs ✓ Run workflow

History Options

Send results to a new history
Yes No

1: Input dataset
2: Exons

2: Input dataset
1: Repeats

3: Join the intervals of two datasets side-by-side (Galaxy Version 1.0.0)

4: Group data by a column and perform aggregate operation on other columns. (Galaxy Version 2.1.1)

5: Sort data in ascending or descending order (Galaxy Version 1.0.3)

6: Select first lines from a dataset (Galaxy Version 1.0.0)

Select first
20
lines

History 2 shown
2.77 MB

2: Exons

1: Repeats

Workflow

Run a workflow

The screenshot displays the Galaxy web interface. At the top, the navigation bar includes 'Analyze Data', 'Workflow', 'Shared Data', 'Visualization', 'Help', and 'User'. A 'Using 0%' indicator is visible in the top right corner. On the left, a 'Tools' sidebar lists various categories such as 'Get Data', 'Send Data', 'Text Manipulation', and 'NGS: QC and manipulation'. The main workspace features a green notification box with a checkmark icon, stating: 'Successfully invoked workflow **Find exons with highest number of SNPs.** You can check the status of queued jobs and view the resulting data by refreshing the History pane. When the job has been run the status will change from 'running' to 'finished' if completed successfully or 'error' if problems were encountered.' On the right, the 'History' panel shows a search bar and a list of workflow steps. Steps 1 through 7 are listed, with steps 1 through 7 highlighted in green. Step 7, '7: Top exon genetic location', is highlighted with a red border. Each step includes an eye icon, a pencil icon, and a close icon. The 'Exons' and 'Repeats' steps are also highlighted in green.

Galaxy Analyze Data Workflow Shared Data Visualization Help User Using 0%

Tools

search tools

Get Data
Send Data
Lift-Over
Text Manipulation
Datamash
Convert Formats
Filter and Sort
Join, Subtract and Group
Fetch Alignments/Sequences
NGS: QC and manipulation
NGS: DeepTools
NGS: Mapping
NGS: RNA Analysis
NGS: SAMtools
NGS: BamTools
NGS: Picard
NGS: VCF Manipulation
NGS: Peak Calling
NGS: Variant Analysis
NGS: RNA Structure
NGS: Du Novo
NGS: Gemini

Successfully invoked workflow **Find exons with highest number of SNPs.**
You can check the status of queued jobs and view the resulting data by refreshing the History pane. When the job has been run the status will change from 'running' to 'finished' if completed successfully or 'error' if problems were encountered.

History

search datasets

Galaxy initiation - workflow
7 shown
2.77 MB

7: Top exon genetic location
6: Top exons
5: Sort on data 4
4: Group on data 3
3: Join on data 1 and data 2
2: Exons
1: Repeats

Workflow

Run a workflow

The screenshot displays the Galaxy web interface. At the top, the navigation bar includes 'Galaxy', 'Analyze Data', 'Workflow', 'Shared Data', 'Visualization', 'Help', and 'User'. A 'Using 0%' indicator is visible in the top right corner.

On the left side, there is a 'Tools' panel with a search bar and a list of tool categories such as 'Get Data', 'Send Data', 'Text Manipulation', and 'NGS: DeepTools'.

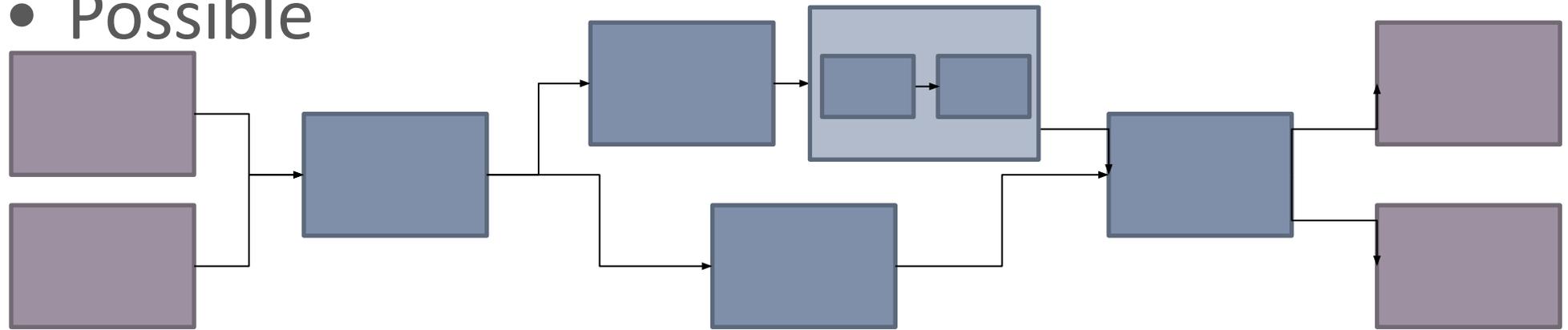
The main workspace contains a green notification box with a checkmark icon. The text reads: 'Successfully invoked workflow **Find exons with highest number of SNPs.** You can check the status of queued jobs and view the resulting data by refreshing the History pane. When the job has been run the status will change from 'running' to 'finished' if completed successfully or 'error' if problems were encountered.'

On the right side, the 'History' panel is visible. It features a search bar and a list of workflow jobs. The current workflow is 'Galaxy initiation - workflow' with a size of 2.92 MB. Below this, a list of jobs is shown, with the top three highlighted in green and enclosed in a red box:

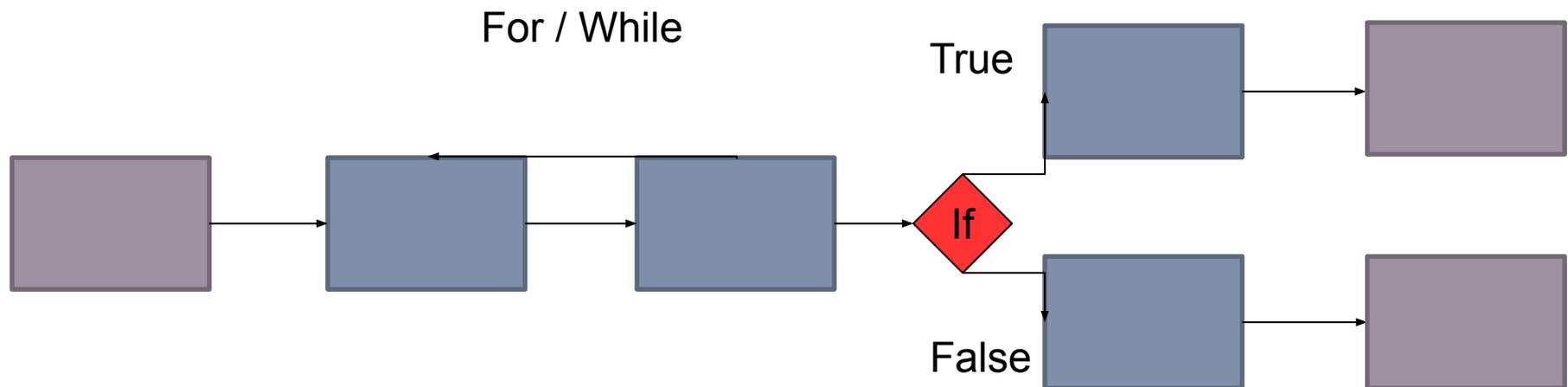
- 7: **Top exon genetic location**
- 6: **Top exons**
- 2: **Exons**
- 1: **Repeats**

Each job entry includes an eye icon (visibility), a pencil icon (edit), and an 'x' icon (delete). The '3 hidden' text in the job count is also highlighted with a red box.

- Possible



- Impossible (until now)



SHARE

biologist ↔ biologist

- Sharing histories or datasets
 - With or without linked workflow

bioanalyst ↔ biologist

- Sharing workflows
 - Pre-configured parameters
 - With or without release parameters (set at runtime)
 - According to the user-end knowledge

bioinformatician ↔ bioinformatician

- Sharing tools ,scripts and wrappers
 - Toolshed



Share

History

The screenshot displays the Galaxy web interface. The main content area is titled "Saved Histories" and contains a table of saved history items. A context menu is open over the first item, "Galaxy initiation - workflow", with the "Share or Publish" option highlighted. The right sidebar shows the "History" panel for the selected workflow, listing several datasets.

Galaxy Analyze Data Workflow Shared Data Visualization Help User Using 0%

Tools search tools

Get Data
Send Data
Lift-Over
Text Manipulation
Datamash
Convert Formats
Filter and Sort
Join, Subtract and Group
Fetch Alignments/Sequences
NGS: QC and manipulation
NGS: DeepTools
NGS: Mapping
NGS: RNA Analysis
NGS: SAMtools
NGS: BamTools
NGS: Picard
NGS: VCF Manipulation
NGS: Peak Calling
NGS: Variant Analysis
NGS: RNA Structure
NGS: Du Novo
NGS: Gemini

Saved Histories

search history names and tags

Advanced Search

Name	Datasets	Tags	Sharing	Size on Disk	Created	Last Update
Galaxy initiation - workflow	4	0 Tags		2.9 MB	~5 hours ago	~4 hours ago
imported: Galaxy 101 (2015)	7	0 Tags		8.8 MB	~11 hours ago	~11 hours ago
Unnamed history		0 Tags		0 bytes	Jun 27, 2016	Jun 27, 2016

Context Menu for "Galaxy initiation - workflow":

- Switch
- View
- Share or Publish**
- Copy
- Rename
- Delete
- Delete Permanently

History Panel:

Galaxy initiation - workflow
4 shown, 3 hidden
2.92 MB

- 7: Top exon genetic location
- 6: Top exons
- 2: Exons
- 1: Repeats

URL: https://usegalaxy.org/history/list?f-sharing=All&sort=-update_time&f-name=All&f-tags=All&f-deleted=False&operation=Share+or+Publish&id=99569b6f012ffc3c

Share

Workflow

Your workflows

Create new workflow Upload or import workflow

Name	# of Steps
Find exons with highest number of SNPs	7
Convert to tab (imported from ...)	2
imported: ChIP-seq workflow	3

- Edit
- Run
- Share or Download**
- Copy
- Rename
- View
- Delete

Workflows shared by others

No workflows have been shared by others

Other options

Configure your workflow menu

Share

Mode

Galaxy Analyze Data Workflow Shared Data Visualization Help User Using 0%

[Go back to Workflows List](#)

Workflow ' Find exons with highest number of SNPs'

Share

This workflow is currently restricted so that only you and the users listed below can access it.

→ **Restricted community**
Generates a web link that you can share with other people so that they can view and import the workflow.

→ **All the Galaxy server users**
Makes the workflow accessible via link (see above) and publishes the workflow to Galaxy's [Published Workflows](#) section, where it is publicly listed and searchable.

You have not shared this workflow with any users yet.

→ **Designated community
(login@sb-roscoff.fr)**

Export

workflow as a file so that it can be saved or imported into another Galaxy server.

This workflow must be accessible. Please use the option above to "Make Workflow Accessible and Publish" before receiving a URL for importing to another Galaxy.

of workflow in SVG format

Export to the www.myexperiment.org site.

- Get shared histories

Galaxy / METABO Analyze Data Workflow Shared Data Visualization Help User Using 216.1 MB

Tools ↑

search tools ×

Get Data

WORKFLOW 4 METABO

2-Preprocessing

3-Normalisation

4-Quality Control

5-Statistical Analysis

Individual

Histories shared with you by others

<input type="checkbox"/>	Name	Datasets	Created	Last Updated↑	Shared by
<input type="checkbox"/>	mmonsoor	6	Apr 28, 2014	~2 days ago	mmonsoor@sb-roscoff.fr

For 0 selected histories: Copy Unshare

History ↻ ⚙

- HISTORY LISTS
- Saved Histories
- Histories Shared with Me
- CURRENT HISTORY
- Create New
- Copy History
- Copy Datasets
- Share or Publish

Galaxy / METABO Analyze Data Workflow Shared Data Visualization Help User Using 70.9 MB

Public

Published Histories

search name, annotation, owner, and tags Q

Advanced Search

Name	Annotation	Owner	Community Tags	Last Updated↓
Preprocessing		mlandi		~14 seconds ago
TP1_xcms_sacuri		mmonsoor	★★★★★	~1 day ago
TP1_xcms_sacuri		jfmartin	★★★★★	Apr 28, 2014

- Data Libraries
- Data Libraries Beta
- Published Histories
- Published Workflows
- Published Visualizations
- Published Pages

- Get shared workflows

Your workflows

[Create new workflow](#) [Upload or import workflow](#)

Name	# of Steps
complete_workflow_RFMF	17

Individual

Workflows shared with you by others

Name	Owner	# of Steps
Workflow mmonsoor	mmonsoor@sb-roscoff.fr	7

Published Workflows

[Advanced Search](#)

- Data Libraries
- Data Libraries Beta
- Published Histories
- Published Workflows
- Published Visualizations
- Published Pages

Public

Name	Annotation	Owner	Rating	Community Tags	Last Updated
complete_workflow_RFMF		mmand			~17 hours ago

- Import shared

Galaxy / METABO Analyze Data Workflow Shared Data Visualization Help User Using 216.1 MB

Published Histories | [mmonsoor](#) | TP1 xcms sacuri Import history About this History

Histories

TP1 xcms sacuri
65.4 MB

search datasets

Dataset	Annotation
1: xset.RData	
2: sampleMetadata.tsv	
3: xset.TICs_raw.pdf	
4: xset.log.txt	

Author
mmonsoor

Related Histories
[All published histories](#)
[Published histories by mmonsoor](#)

Rating
 Community (0 ratings, 0.0 average) ★★★★★
 Yours ★★★★★

Tags
Community: none

Galaxy / METABO Analyze Data Workflow Shared Data Visualization Help User Using 216.1 MB

Workflows

[Create new workflow](#) [Upload or import workflow](#)

Your workflows

Name	# of Steps
complete_workflow_RFMF	17

Workflows shared with you by others

Name	Owner	# of Steps
Workflow mmonsoor	mmonsoor@sb-roscoff.fr	7

Other

[View](#) [Run](#) [Copy](#) [Remove](#)



Level 5

- Share tools and descriptions in the ToolShed



Level 4

- Launch tools autonomously
- Use advanced parameters
- Use the Galaxy API
- Provide workflow for colleagues Level 1-3



Level 3

- Launch tools autonomously
- Use workflow more or less preset



Level 2

- Use preset workflow



Level 1

- Share his data to colleagues Level 2-5

Hands-on
WORKFLOW





Which coding exon has the highest number of ~~single nucleotide polymorphisms~~ on chromosome 22?
repeats

1. Extract a workflow from your history
2. Edit the workflow (hide intermediate steps, rename inputs/outputs, set parameters at runtime, save)
3. Create a new history with the input data
4. Run the workflow
5. Share your history/workflow with your neighbour



CONCLUSION

- Easy-to-use graphical user interface for often complex command-line tools
- Keeps a full record of your analysis in a history
- Workflows enable you to repeat your analysis on different data
- Galaxy can connect to external sources for data import and visualization purposes
- Galaxy provides ways to share your results and methods with others

Tools - panel

125+ platforms for using Galaxy: <https://galaxyproject.org/use>
[Servers, clouds, deployable resources]

GENERAL PURPOSE:

usegalaxy.org, usegalaxy.eu, usegalaxy.fr, usegalaxy.org.au

DOMAIN SPECIFIC:



RNAseq: <http://galaxy.sb-roscoff.fr>

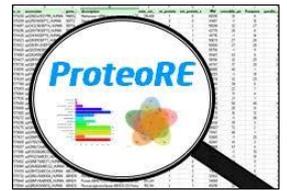
SBR tools: <http://webtools.sb-roscoff.fr>

Metagenomics: <http://galaxy4metab.sb-roscoff.fr>

Metabolomics:



Proteomics:



ChIP-seq:



Catalog of French Galaxy tools:
<http://galaxycat.france-bioinformatique.fr>

Galaxy ABiMS --> Galaxy France

<https://galaxy.sb-roscoff.fr/>



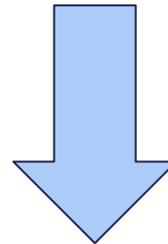

Galaxy / ABiMS | Analyze Data | Workflow | Visualize | Shared Data | Admin | Help | User | Using 34%

Tools | search tools | Get Data | Send Data | Collection Operations | COMMON TOOLS

Welcome to galaxy.sb-roscoff.fr

14-10-2020
Dear users, we plan to eventually close this <https://galaxy.sb-roscoff.fr> instance in favour of <https://usegalaxy.fr>. So, if you start a new analysis, please consider migrating to the new national instance <https://usegalaxy.fr>. This instance is more modern, supported by the IFB Core Cluster, maintained in part by ABiMS and the IFB NNCR Cluster TaskForce. If tools are missing, do not hesitate to request them on <https://community.france-bioinformatique.fr/>.

History | search datasets | Galaxy initiation training 2019 - tools hands-on | 11 shown, 4 deleted, 7 hidden | 11.73 MB



<https://usegalaxy.fr/>




Galaxy France | Analyze Data | Workflow | Visualize | Shared Data | Help | Login or Register | Using 0%

Tools | search tools | Get Data | Send Data | Collection Operations | GENERAL TEXT TOOLS | Text Manipulation | Filter and Sort | Join. Subtract and Group

Welcome to usegalaxy.fr

By using this Galaxy instance, we assume that you have read and accept the [Term Of Use](#)
For any questions or support: community.cluster.france-bioinformatique.fr/c/galaxy

History | search datasets | Unnamed history (empty) | This history is empty. You can load your own data or get data from an external source

- On your own:

- Training materials:



- <https://galaxyproject.github.io/training-material>

- Interactive tours of Galaxy:

- <http://galaxy.sb-roscoff.fr/tours>

- Training courses:

Training	What ?	Where ?	When ?
RNAseq analysis with Galaxy	RNAseq	Roscoff, France	first semester 2021
Galaxy Community Conference (GCC)	General purpose (data-intensive biology and Galaxy)	Ghent, Belgium	July 2021
Workflow4Experimenters	Metabolomics	Toulouse, France	October 2021



END



Thank you for completing the training evaluation questionnaire:

http://abims.sb-roscoff.fr/evaluation_formation

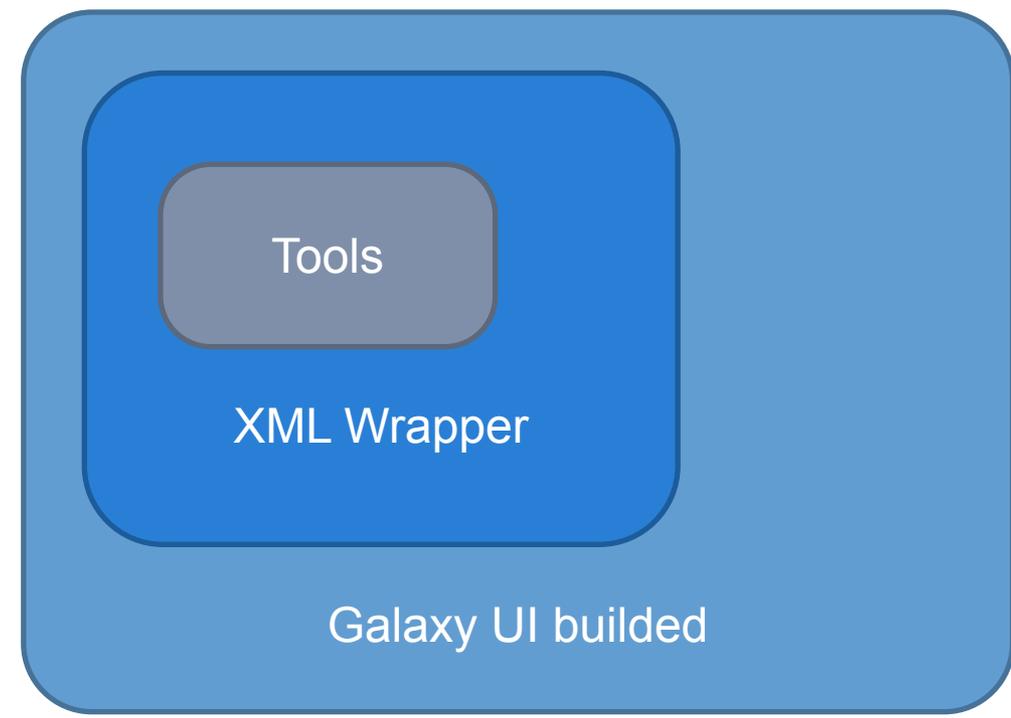
BONUS



How are tools born?

BONUS

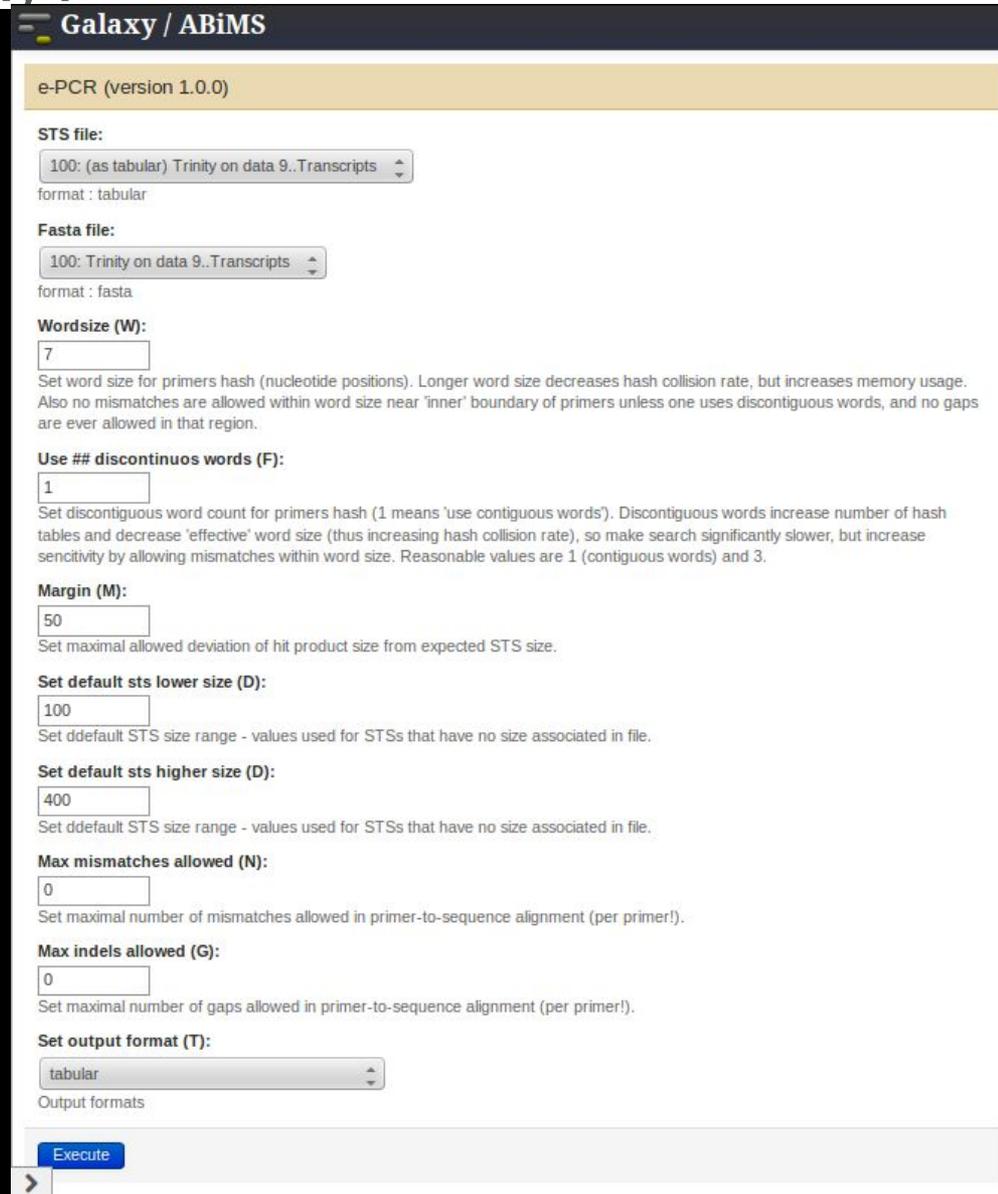
- How to import a tool in Galaxy?



• How to import a tool in Galaxy?

```

[lecorguille@n0 ~]$ e-PCR --help
e-PCR: invalid option -- -
usage: [-hV] [posix-options] stsfile [fasta ...]
[compat-options]
where posix-options are:
    -m ##      Margin (default 50)
    -w ##      Wordsize (default 7)
    -n ##      Max mismatches allowed (default 0)
    -g ##      Max indels allowed (default 0)
    -f ##      Use ## discontinuous words, slow if
                ##>1
    -o ##      Set output file
    -t ##      Set output format:
                1 - classic, range (pos1..pos2)
                2 - classic, midpoint
                3 - tabular
                4 - tabular with alignment in comments
                    (slow)
    -d##-##    Set default size range
                (default 100-350)
    -p +-      Turn hits postprocess on/off
    -v ##      Verbosity flags
    -a a|f     Use presize alignmens (only if
                gaps>0), slow
                a - Always or f - as Fallback
    -x +-      Use 5'-end lowercase masking of
                primers (default -)
    -u +-      Uppercase all primers (default -)
    
```



Galaxy / ABiMS

e-PCR (version 1.0.0)

STS file:

 format : tabular

Fasta file:

 format : fasta

Wordsize (W):

 Set word size for primers hash (nucleotide positions). Longer word size decreases hash collision rate, but increases memory usage. Also no mismatches are allowed within word size near 'inner' boundary of primers unless one uses discontinuous words, and no gaps are ever allowed in that region.

Use ## discontinuous words (F):

 Set discontinuous word count for primers hash (1 means 'use contiguous words'). Discontinuous words increase number of hash tables and decrease 'effective' word size (thus increasing hash collision rate), so make search significantly slower, but increase sensitivity by allowing mismatches within word size. Reasonable values are 1 (contiguous words) and 3.

Margin (M):

 Set maximal allowed deviation of hit product size from expected STS size.

Set default sts lower size (D):

 Set ddefault STS size range - values used for STSs that have no size associated in file.

Set default sts higher size (D):

 Set ddefault STS size range - values used for STSs that have no size associated in file.

Max mismatches allowed (N):

 Set maximal number of mismatches allowed in primer-to-sequence alignment (per primer!).

Max indels allowed (G):

 Set maximal number of gaps allowed in primer-to-sequence alignment (per primer!).

Set output format (T):

 Output formats

- How to import a tool in Galaxy?

```

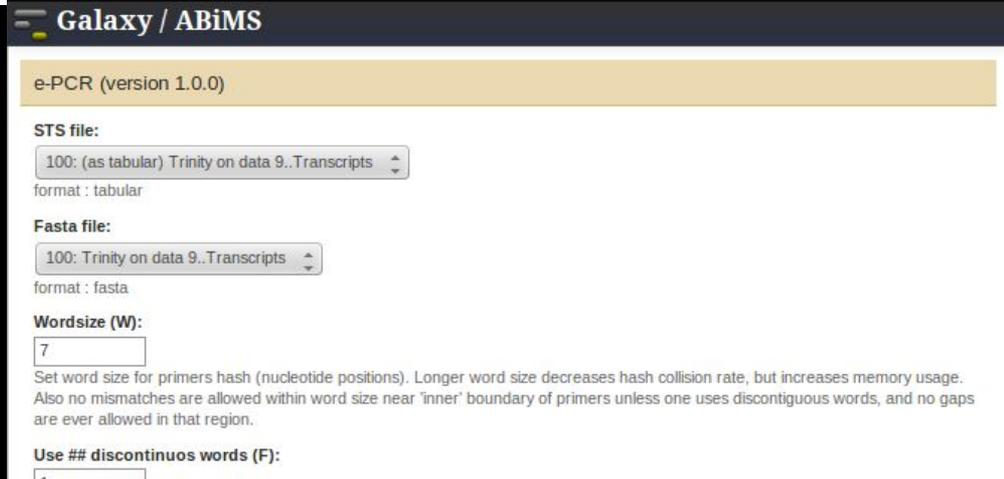
<tool id="abims_epcr" name="e-PCR">
> <!-- author : lecorguille@sb-roscoff.fr -->
> <!-- date : 11-05-12 -->
> <description>e-PCR parses stsfile in unists format, then reads nucleotide sequence data in FASTA format from files listed in commandline if
any, or from stdin otherwise. For input sequences e-PCR finds matches and prints output in one of three formats.</description>
>
> <command>e-PCR -w $wordsize -f $wordcnt -m $margin -d$sts_size_lo-$sts_size_hi -n $max_mismatch -g $max_gap -t $output_format $infile_stsfile
$infile_fasta > $output</command>
>
> <inputs>
> > <param name="infile_stsfile" type="data" label="STS file" format="tabular" help="format : tabular" />
> > <param name="infile_fasta" type="data" label="Fasta file" format="fasta" help="format : fasta" />
> > <param name="wordsize" type="integer" label="Wordsize (W)" value="7" help="Set word size for primers hash (nucleotide positions).
Longer word size decreases hash collision rate, but increases memory usage. Also no mismatches are allowed within word size near
'inner' boundary of primers unless one uses discontinuous words, and no gaps are ever allowed in that region." />
> > <param name="wordcnt" type="integer" label="Use ## discontinuos words (F)" value="1" help="Set discontinuous word count for primers
hash (1 means 'use contiguous words'). Discontinuous words increase number of hash tables and decrease 'effective' word size (thus
increasing hash collision rate), so make search significantly slower, but increase sencitivity by allowing mismatches within word
size. Reasonable values are 1 (contiguous words) and 3." />
> > <param name="margin" type="integer" label="Margin (M)" value="50" help="Set maximal allowed deviation of hit product size from
expected STS size." />
> > <param name="sts_size_lo" type="integer" label="Set default sts lower size (D)" value="100" help="Set ddefault STS size range - values
used for STSs that have no size associated in file." />
> > <param name="sts_size_hi" type="integer" label="Set default sts higher size (D)" value="400" help="Set ddefault STS size range -
values used for STSs that have no size associated in file." />
> > <param name="max_mismatch" type="integer" label="Max mismatches allowed (N)" value="0" help="Set maximal number of mismatches allowed
in primer-to-sequence alignment (per primer!)." />
> > <param name="max_gap" type="integer" label="Max indels allowed (G)" value="0" help="Set maximal number of gaps allowed in primer-to-
sequence alignment (per primer!)." />
> > <param name="output_format" type="select" help="Output formats">
> > > <label>Set output format (T)</label>
> > > <option value="1">classic, range (pos1..pos2)</option>
> > > <option value="2">classic, midpoint</option>
> > > <option value="3" selected="true">tabular</option>
> > > <option value="4">tabular with alignment in comments (slow)</option>
> > </param>
> </inputs>
>
> <outputs>
> > <data name="output" format="tabular" />
> </outputs>

```

• How to import a tool in Galaxy?

```

[lecorguille@n0 ~]$ e-PCR --help
e-PCR: invalid option -- -
usage: [-hV] [posix-options] stsfile [fasta ...]
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where posix-options are:
    -m ##      Margin (default 50)
    -w ##      Wordsize (default 7)
    -n ##      Max mismatches allowed (default 0)
    -g ##      Max indels allowed (default 0)
    -f ##      Use ## discontinuos words, slow if
                ##>1
    
```



Galaxy / ABiMS

e-PCR (version 1.0.0)

STS file:

 format : tabular

Fasta file:

 format : fasta

Wordsize (W):

 Set word size for primers hash (nucleotide positions). Longer word size decreases hash collision rate, but increases memory usage. Also no mismatches are allowed within word size near 'inner' boundary of primers unless one uses discontinuous words, and no gaps are ever allowed in that region.

Use ## discontinuos words (F):

```

<tool id="abims_epcr" name="e-PCR">
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  <description>e-PCR parses stsfile in unists format, then reads nucleotide sequence data in FASTA format from files listed in commandline if any, or from stdin otherwise. For input sequences e-PCR finds matches and prints output in one of three formats.</description>
  <command>e-PCR -w $wordsize -f $wordcnt -m $margin -d$sts_size_lo-$sts_size_hi -n $max_mismatch -g $max_gap -t $output_format $infile_stsfile $infile_fasta > $output</command>
  <inputs>
    <param name="infile_stsfile" type="data" label="STS file" format="tabular" help="format : tabular" />
    <param name="infile_fasta" type="data" label="Fasta file" format="fasta" help="format : fasta" />
    <param name="wordsize" type="integer" label="Wordsize (W)" value="7" help="Set word size for primers hash (nucleotide positions). Longer word size decreases hash collision rate, but increases memory usage. Also no mismatches are allowed within word size near 'inner' boundary of primers unless one uses discontinuous words, and no gaps are ever allowed in that region." />
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    <param name="sts_size_lo" type="integer" label="Set default sts lower size (D)" value="100" help="Set ddefault STS size range - values used for STSs that have no size associated in file." />
    <param name="sts_size_hi" type="integer" label="Set default sts higher size (D)" value="400" help="Set ddefault STS size range - values used for STSs that have no size associated in file." />
    <param name="max_mismatch" type="integer" label="Max mismatches allowed (N)" value="0" help="Set maximal number of mismatches allowed
    
```